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EXCITABILITY EFFECT OF EXTRACT OF DATURA STRAMONIUM LEAVES; SERUM BIOCHEMISTRY AND HAEMATOLOGY IN RATS

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ABSTRACT

The effect of administration of ethanol extract of the leaves of *Datura stramonium* on excitability, hematology and serum biochemistry parameters was studied in rats. Sixteen (16) experimentally matured male Wistar rats, aged 9 - 10 weeks, with an average weight ranged, 147-150g were distributed into 4 groups of 4 rats each, and group 1 were administered with distilled water (as control) groups 2, 3 and 4 were the test groups and were given 100, 200 and 400 mg/kg body weight of the leaf extract respectively. The treatment period was fourteen (14) days. Excitability scores were recorded daily immediately after administration of extract. End of treatment the rats were sacrificed and blood collected by cardiac puncture for analysis. The result showed that *D. stramonium* demonstrated increased excitability mean score of 2.50 ± 0.28 , 3.25 ± 0.25 , 3.50 ± 0.29 compared to control group 1.25 ± 0.25 which may suggest nervous system excitation in those treated rats more than the control rats. There were mild changes in haematological parameter not significantly altered but within normal reference range. However, the mild leukocytosis, lymphocytosis, neutrophilia and monocytosis were not significant and elevation was within normal reference range. The result of the serum biomarkers showed significant decreases in the liver and kidney enzymes suggesting that the leaf extract at lower doses possess hepato, nephro and cardio-protective properties It can be concluded that the extract of *Datura stramonium* exhibited excitatory effect, with no toxic effects on the blood profile and internal organs.

Index terms: Datura stramonium, Excitability, Haematology, Rats, Serum Biochemistry.

INTRODUCTION

Plants have always played a major role in the treatment of human and animal traumas and diseases worldwide. The demand for medicinal plant is increasing in both developed and developing countries due to growing recognition of natural product. Herbal medicine is an important part of both traditional and modern system of medicines, Kirtikar et al., (1994). Plants contain primary and secondary metabolites. Primary metabolites are those compounds the plants uses for its metabolism. They are protein, carbohydrates and lipids which are used by the plant for their growth and nourishment. Secondary metabolites on the other hand are compounds found in the plants that are used by the plants to protect itself, Ray et al., (2013). Animals in the wild do not have medical care and treat themselves in the wild by identifying plants with medicinal properties, this is what we try to bring home by identifying those medicinal plants for proper documentation and necessary use, Ijeh et al., (2009). D. stramonium also known by the common

names thorn apple, jimsonweed, devil's snare, devil's trumpet or Zegemi in northern and southern Nigeria is one of the widely known folklore medicinal herb of the flowering plant family Solanaceae. It is a wild growing flowering plant and was investigated as a local source for tropane alkaloids which contain a methylated nitrogen atom (N-CH₃) and include the anti-cholinergic drugs atropine, and scopolamine amongst others. The plant is a severely toxic and poisonous plant that has been and is still being used traditionally to treat ailments associated with all kinds of inflammations and all manner of burns and scalds, Glatstein et al., (2016). The leaf although toxic is generally smoked either in a cigarette or a pipe and has been stipulated to have excitatory or hyperactivity effect, Glatstein et al., (2016). There are several ways to ingest D. Stramonium, the plants can be smoked, brewed into a tea, or converted into a skin ointment. Since all parts of the plant contain variable amounts of psychedelic compounds, some people have been known to chew the seeds as well, Freye (2010). The

incidence of D. stramonium poisoning is sporadic with a cluster of poisoning cases in the 1990s and 2000s, the United States media reported some cases occurring mostly among adolescents and young adults dying or becoming seriously ill from ingesting. Some medicinal uses of the plant are its anti-inflammatory property of all parts of the plant, stimulation of the central nervous system, respiratory decongestion, treatment of dental and skin infections, alopecia and in the treatment of toothache amongst other things, Das et al., (2012). Herbal indigenous knowledge possessed by different communities on variety of human and animal medical herds has remained unwritten with the risk of being forgotten and totally lost. Therefore, research on herbs especially medicinal herbs should be documented. Prospects for human and ethno-veterinary research and development should be identified to enable researchers chart new courses of investigation and establish scientific basis of their action and develop potentials of the herb as a medicinal plant, Vijendra and Kumar (2010). D. stramonium consumed by domestic animal may have excitability or hyperactivity effects on its consumers and lead to environmental self-pollution, Ikpeazu and Igwe (2023). There is need to have a knowledge on the effects which it could have on liver, kidney and hematology since the use of this plant could be indicated in the treatment of depression or inactiveness on domestic animals especially dogs intended for warfare or security. Dog owners sometimes request for substances that can make their dogs hyperactive or aggressive and this study wants to check if D. stramonium can handle the task.



Figure 1: *Datura stramonium* plant showing the leaves, flower and fruit.

The genus name 'Datura' is derived from dhatura, the Bengali name for the plant, while the epithet 'stramonium' combines the Greek word strychnos for nightshade, and makinos meaning mad, referring to the narcotic properties of the plant (Haegi, 1976; Hadkins *et al.*, 1997). Origin is probably the tropical regions of Central and South America, *D. stramonium* has become a cosmopolitan weed in the warm regions of North, Central and South America, Europe, Asia, Africa (especially in Nigeria) and New Zealand and has been used as narcotic by British soldiers, Parsons *et al.*, (1992). *D. stramonium* also known as Jimson weed,

devil's snare or apple, thorn Apple or Devil's trumpet is an erect, annual, freely branching herb that forms a bush up to 60 to 150 cm (2 to 5 ft.) tall, Griev, e (1971). Indigenous names of the plant include; Zakami or Zegemi in Hausa, Myaramwo in Igbo, Zakedi in Kanuri, Gegemu in Yoruba and Jegemi in Igala of Nigeria (Gidado et al., 2007; Parker et al., 2021). The root is long, thick, fibrous, and white. The stem is stout, erect, leafy, smooth, and pale yellow-green to reddish purple in color. The stem forks off repeatedly into branches and each fork forms a leaf and a single erect flower, Grieve (1971). The leaves are about 8 to 20 cm (3-8 in) long, smooth, toothed, soft, and irregularly undulated. The upper surface of the leaves is a darker green, and the bottom is a light green. The leaves have a bitter and nauseating taste, which is imparted to extracts of the herb, and remains even after the leaves have been dried, Grieve (1971). The proximate analysis of Datura stramonium leaves and contain carbohydrate, protein, ash, lipid, fiber and water, vitamin C, phosphate, phosphorus, nitrate, nitrogen; metallic minerals such as manganese, calcium, sodium, potassium, iron, and trace elements cadmium, copper, zinc and lead in a negligible amount Ibiam et al. (2017). GC-MS analysis has also been used to identify compounds in plants. (Igwe et al., 2016; Ikpeazu et al 2017). Otuokere et al. (2020), in their GC-MS analysis of D. stramonium leaves revealed the presence of eighteen bioactive compounds. D. stramonium used traditionally to treat asthma, epilepsy, rheumatoid arthritis, Chavhan et al., (2018). Treatment of injuries, wounds, bleeding and management of pains, Njoroge, (2012). Its seeds are used as purgative, in cough, fever and asthma, Aquib et al., (2013). The seeds are smoked due to its narcotic action, Khan et al., (2013), used to make somebody unconscious Rahmatullah et al.. (2009), also used for baldness, Khan et al., (2008) Applied to scalp for falling hairs and as antidandruff et al., (2009). Shah et al., (2006). Paste of leaves is topically applied for skin diseases, Rahmatullah et al., (2010). Pharmacologically it has anti-asthmatic activity and leaves are used in asthma treatment, Savithramma et al., (2007). It has anticholinergic activity as reported by Taha et al., (1984). The alkaloids found in D.stramonium , are organic esters used clinically as anticholinergic agents. The anticholinergic syndrome results from the inhibition of central and peripheral muscarinic neurotransmission, Taha et al., (1984). As acaricidal, repellent and oviposition deterrent properties, Kurnal et al., (2009) and antimicrobial activity against gram positive bacteria in a dose dependent manner (Sharma et al., 2010, Fereshteh et al., 2004). Toxicity to environment and body chemistry, Ikpeazu and Igwe (2023). Acute toxicity test (LD₅₀) of *D.stramonium* using up and down method was 3185.25 mg/kg b.w. Khudhair and Abed, (2015)

Laboratory reports on excitability studies in animal models

Medically excitability refers to a state of being alert and hyperactive. An animal can be said to hyperactive if it shows the following; Alert and awake (absence of sleep), a state of panic (ie. increased pacing), easily aggrieved, raised blood pressure, an increased feeling of energy, may tend to have an increased libido and may have a tendency to get into a fight with other herd mates Ziad *et al.*, (2019). Excitability at cellular level describes the ease with which cells respond to stimulus with a regenerative action potential depending on the passive and active properties of the cell membrane, Ziad *et al.*, (2019). Adenkola and Oluremi (2014) reported excitability scores of rabbits fed graded level of *Hibiscus sabdariffa* calyx (HSC), showed increasing excitability scores as the inclusion levels were increased.

MATERIALS AND METHODS

Plant Material

Fresh leaves of *Datura stramonium* were collected at Ubakala in Umuahia, Abia State, Nigeria. The plant was identified using Google Plant Identifier and confirmed by the Forestry Department of Michael Okpara University of Agriculture Umudike, Nigeria.

Preparation of Extract

Cold masceration technique was used in this study as described by Igwe *et al* (2020). The leaves were dried under room temperature for 10 days and were grounded using manual grinder model (Corona-Landers C 1A SA). The grounded leaves were soaked in ethanol for 48 hours and was filtered using filter paper. The extract was

concentrated using hot air at 30 0 C and the dried extract weighed and kept in labelled sterile specimen bottle for the experiment.

Experimental Design

Sixteen (16) adult male albino rats were used for the study. The rats were acclimatized for one week and fed *ad libitum* with standard pelleted rat feed. The Wistar rats were randomly distributed into 4 groups; (1-4). Rats in group 2, 3 and 4 were given 100, 200 and 400mg/kg graded doses of the extract respectively, via oral gavage, while group 1 received distilled water at 5ml/kg body weight, and served as the normal control. The treatment lasted for fourteen (14) days.

Measurement of Excitability Scores in rats

Excitability scores were recorded daily immediately, five (5) minutes after administration. They were measured as described by Adenkola and Oluremi (2014)), while weighing them. A score of one to four was allocated to each rat by a single observer; a higher score representing a greater level of excitability. A score of one was allocated to a rat that was calm, and made little movement during the handling. Two was allocated to a rat that occasionally shook itself in an attempt to escape from the weighing pan, while three was assigned to a rat that continuously attempted to free itself. A score of four was given to a rat that struggled violently the entire weighing period.

Excitability Scores in rats		
-	Scores	Degree of excitability
	1	No struggling, calm, and make little movement during the handling/weighing
	2	No struggling, occasionally shake itself in an attempt to escape from the weighing pan
	3	No struggling, continuously attempt to free itself from the weighing pan
	4	Struggling violently throughout the entire weighing period.
Source : Adenkola and Oluremi (2014)		
$MCV (\mu m^3)$	=	$PCV \times 10$

Number of erythrocytes per μ L blood $\times 10^{-6}$

MCH (μ g) = Haemoglobin (g/dL) × 10 Number of erythrocytes per μ L blood × 10⁻⁶

$$MCHC (g/dL) = \frac{Haemoglobin (g/dL) \times 100}{PCV (mL / dL)}$$

Serum Clinical Biochemistry Determination Procedures

The serum ALT activity was determined by the Reitman-Frankel colorimetric method, Reitman and Frankel, (1957). The serum AST activity was determined by the Reitman-Frankel colorimetric method, Reitman and Frankel, (1957). The serum ALP activity was determined by phenolphthalein monophosphate method (Klein *et al.*, 1960; Babson *et al.*, 1966). The serum total protein levels were assayed by the direct Biuret method, (Lubran, 1978; Johnson, 2008). The serum total bilirubin levels were determined by Jendrassik and Grof method (Doumas *et al.*, 1973; Higgins *et al.*, 2008). The serum total cholesterol was determined by enzymatic colorimetric method, Allain *et al.*, (1974).

STATISTICS

The data will be analyzed using statistical package of social sciences (SPSS) version 23. Data will be expressed as Mean \pm Standard Error of mean. The data will be subject to One-way analysis of varience (ANOVA). The different doses will be compared and

seperated using posthoc analysis (Duncan test) to check mean that is significant. The statistical confidence will be placed at 95% (p<0.05)



RESULTS



Figure 2 shows excitability effect of D. stramonium on Wistar rats.

Excitability of animals depends on their temperament, and temperament is a trait that seems to be stable over time. The excitability score in the experimental rats treated with the graded doses of the extract were increased as the doses were increased in a dose dependent manner. On a scale of 4, the 100, 200 and 400 mg/kg graded doses of the extract recorded mean scores

of 2.50 ± 0.28 , 3.25 ± 0.25 and 3.50 ± 0.29 , respectively, compared to 1.25 ± 0.25 excitability mean score of the control rats. Similarly, the number of times the individual rat from each experimental group attempted to jump out from the weighing trough were higher in the extract groups as the doses were increased and compared to control.



Figure 3 shows the effect of *D. stramonium* extract on the erythrogram of Wistar albino rats.

Values presented as mean \pm S.E.M (Standard Error of Mean) at (p<0.05).

Hb (g/dl) 16.00±0.64, 15.37±0.13, 15.65±0.86 compared to control 15.15±0.02;

PCV (%) 47.25±1.79, 44.00±0.70, 45.00±2.27, compared to control 43.75±0.47;

RBC ($\times 10^{12}$) 7.15±0.25, 7.02±0.06, 7.57±0.44, compared to control 7.01±0.01;

MCV (**fl**) 66.08±1.84, 62.69±1.22, 59.56±1.68 compared to control 62.34±0.67;

MCH (pg) 22.35±0.16, 21.90±0.10, 20.78±1.22, compared to control 21.58±0.03; MCHC (g/dl), 35.89±0.93, 34.96±0.57, 34.81±1.07 compared to control 34.63±0.31.

The result shows no significant difference between the treated and control groups at (p<0.05) on the erythrogram.



Figure 4: graph shows the effect of D stramonium extract on leukogram of Wistar albino rats.

Values are presented as mean \pm S.E.M (Standard Error of Mean) significant difference at (p<0.05).

TWBC ($\times 10^{9}$ /L), 9.62±01.25^a, 8.80±0.14^b, 8.13±0.04^c compared to control 7.23±0.02^d; Lymphocytes (%) 61.25±0.47^a, 61.25±0.25^a, 56.75±2.28^b compared to control 57.25±0.25^b;

Neutrophils (%) 34.75 ± 0.94^{a} , 34.25 ± 0.25^{a} , 32.25 ± 0.25^{b} compared to control 32.75 ± 0.62^{b} ;

Eosinophils (%) 1.25 ± 0.25^{b} , 2.25 ± 0.25^{a} , 2.75 ± 0.25^{a} compared to control 3.00 ± 0.40^{a} ; **Monocytes** (%) 5.00 ± 0.40^{a} 4.75 ± 0.47^{a} 3.80 ± 0.28^{b} compared to control 3.50 ± 0.28^{ab} ; **Basophils** (%) 0.00 ± 0.00 0.00 ± 0.00 0.00 ± 0.00 compared to control 0.00 ± 0.00

The result showed no significant difference between the treated and control groups at (p<0.05) on the leukogram. There was mild eosinopaenia and monocytosis.



Figure 5: graph shows the effect of *D. stramonium* extract on serum biochemical parameter of Wistar albino rats.

Creatinine (mg/dl): 0.85±0.01^b, 0.96±0.05^a, 0.87±0.01^{ab}

There were no significant changes in the levels of serum

total protein (TP), Aspartate Transaminase (AST),

Phosphatase (ALP) evaluated. All values were within

normal reference range across the treatment groups

compared to the control. Similarly, the kidney enzymes;

urea, creatinine and the serum bilirubin were also within

normal reference range compared to control indicating

(ALT)

and

Alkaline

compared to control 0.93±0.01^{ab}.

liver and kidney safety.

Alanine

Aminotransferase

Values are presented as mean \pm S.E.M (Standard Error of Mean) significant difference at (p<0.05).

TP (g/dl): 6.06 ± 0.28^{b} , 7.08 ± 0.35^{a} , 6.77 ± 0.19^{ab} compared to control 6.40 ± 0.03^{ab} ;

AST (**IU/L**): 62.30 ± 1.03^{b} , 60.94 ± 0.15^{b} , 61.76 ± 0.26^{b} compared to control 64.78 ± 0.26^{a} ;

ALT (IU/L): 36.11 ± 0.59^{b} , 36.54 ± 0.70^{b} , 36.54 ± 0.70^{b} compared to control 39.02 ± 0.27^{a} ;

ALP (**IU/L**): $66.98\pm0.56^{\circ}$, $69.02\pm0.84^{\circ}$, $65.33\pm0.31^{\circ}$ compared to control $71.56\pm0.15^{\circ}$;

Bilirubin (mg/dl): 0.31 ± 0.10^{b} , 0.48 ± 0.02^{ab} , 0.46 ± 0.01^{ab} compared to control 0.55 ± 0.00^{a} ;

Urea (mg/dl): 18.01 ± 0.49^{b} , 19.89 ± 0.21^{a} , 18.67 ± 0.29^{b} compared to control 20.06 ± 0.05^{a} ;



Figure 6: Effect of D. stramonium extract on lipid profile of Wistar albino rats.

Values are presented as mean \pm S.E.M (Standard Error of Mean) significant level at (p<0.05).

Total cholesterol (mg/dL): $98.03\pm5.75^{\text{b}}$, $79.67\pm0.24^{\text{c}}$, $108.40\pm1.44^{\text{a}}$ compared to control $103.52\pm0.50^{\text{ab}}$;

 71.55 ± 1.09^{b} , Triglyceride (mg/dL): $64.00\pm0.45^{\circ}$, 76.80 ± 1.54^{a} compared to control 77.74 ± 0.28^{a} ; HDLC (mg/dL): 43.05 ± 0.38^{b} , 45.95±0.24^a, 42.61±0.55^b compared to control 42.98±0.26^b; LDLC (mg/dL): 40.67 ± 5.86^{a} , 20.91 ± 0.17^{b} , 50.43 ± 1.68^{a} compared to control 44.98±0.50^a; VLDLC (mg/dL): 14.31±0.21^b, 15.36 ± 0.30^{a} compared $12.80 \pm 0.09^{\circ}$, to control 15.54±0.05^a. Triglyceride (TG) level and Very Low Density Lipoprotein Cholesterol (VLDLC) were reduced compared to the control. The High Density Lipoprotein Cholesterol (HDLC) was mildly elevated within normal reference range. These indicate safety and could prevent cardiac associated diseases like arthereosclerosis.

DISCUSSION

Most of the natural herbs used in traditional medicine are believed to be safe, compared to synthetic drugs, even when there are no toxicological records or scientific evidence to this believe.

Excitability of animals depends on their temperament and temperament, and such animal seems to be stable over time. The excitability score and the number of times each experimental rat attempted to escape from the weighing trough in the group treated with the graded doses of the extract were increased (p<0.05) as the dose of the extract were increased in a dose dependent manner, higher than the control rats. It has been reported that the primary biologically active substances in D. stramonium are the alkaloids atropine and scopolamine (Otuokere et al., 2020). D. stramonium has been documented to contain vitamin C (Wang et al., 2000; Essas et al., 2006) and other antioxidant principles (Tee et al., 2002; Ologundudu et al., 2009), and has been shown to increase excitability scores in animals, Avo et al., (2006) possibly because it plays a significant role in the synthesis of vitaminergic neurotransmitters in the brain. This could possibly explain the higher excitability score observed in rats administered with the extract especially, with higher doses of D. stramonium. This

may suggest the ability of *D. stramonium* leaf extract to activate the nervous system in those treated rats more than the control group. The excitability effect of the leaf extract as observed in this study could also could be due to the phytochemical compound; 2-Phenanthrenol, 1,2,3,4,4a,9,10,10a-octahydro-7-methoxy-1,1,4a-trimethyl- as reported by Otuokere *et al.* (2020), which is 5-alpha-reductase-inhibitor, and acetyl-CoA-carboxylase-inhibitor known to elicit excitability in animals.

Haematological and serum biochemical indices are used in monitoring feed, drug toxicity and health status of Ovawove and Ogunkunle, (2004). No animals. significant change in erythrogram values as recorded in this study (Fig 3), might be attributed to the non-toxic effect of Datura on the haematological system Bouzidi et al. (2011). According to Fatoba et al. (2013) the determination of total leucocyte and differential count are important markers of immune function. In this study, the total leukocyte counts of the D. stramonium extract treated groups significantly (p<0.05) increased relative to the control (Fig 4) but within normal reference range. D. stramonium caused mild leukocytosis in this research. This result is consistent with the findings of Fatoba et al. (2013), who stated that rabbits treated with aqueous seed extract of D. stramonium recorded higher white blood cell counts than the control. The result of the TWBC appears to validate the indigenous claim that the leaves of this plant can boost the body immunity, and assist the body system to fight against mixed infections (Iwuji and Herbert, 2012; Soetan et al., 2013; Isaac et al., 2013). The lymphocyte counts of low and medium doses were observed to be significantly (p<0.05) higher when compared with the highest dose (Fig 4). This implies that extract at low dose was enough to stimulate very high lymphocytes production and has capacity to boost both antibody-dependent and cell-mediated immune responses since lymphocytes play major role in immune responses. This observation strongly supports the findings of Fatoba et al. (2013) who reported increase in lymphocyte counts with decreasing extract dosage. The neutrophil counts also increased significantly (p<0.05) in low and medium doses compared with the control and the highest dose groups; suggesting that the extract at lower doses can stimulate cell mediated elimination of bacterial pathogens because major phagocytes mobilized during bacterial invasion are neutrophils. As observed in the lymphocytes count, the effect had an inverse dose dependence effect, meaning that lower doses of the extract appeared to stimulate more synthesis of neutrophils than the highest dose. This finding strongly affirm the study done by Ogunmoyole et al. (2019), who reported that D. stramonium extract at higher dose of 400 mg/kg generally caused a significant decrease in neutrophils count. Monocyte counts significantly (p<0.05) followed the same trend as leucocytes and lymphocytes compared to all control groups. The eosinophil is specialized in producing peroxidase enzymes and proteins that are toxic to invading elements

and is seen to increase in the presence of parasites, toxin or allerge (Faghani *et al.*, 2014). The eosinophils count decreased significantly (p<0.05) in the 100 mg/kg dose relative to 200, 400 mg/kg doses and the normal control respectively. No change in Basophil count.

AST and ALT elevation is seen in condition of hepatocyte damage, in inflammatory condition of liver, hepatotoxicity by toxicants, trauma and some plant extracts, Reitman and Frankel, (1957). Liver ALP elevation is seen in hepatocyte and biliary epithelial damage. In kidney, elevated creatinine occurs in pathological condition of decrease in glomerular filtration rate which could be pre-renal, renal or post renal. Bouzidi et al. (2011). An obvious sign of hepatic injury is leakage of cellular enzyme into plasma Udem et al., (2009). When the liver cell membrane is damaged, a variety of enzymes normally located in the cytosol are released into blood stream accounting for their raised levels in the serum (Kumar et al., 2004; Batool et al., 2017). The elevation is a consequence of hepatic injury, resulting in the leakage of enzymes that are normally localized within the hepatocytes. Similarly, kidneys as the principal organ for the excretion of xenobiotics and their metabolites are particularly prone to their toxic effects. Estimation of serum levels of these enzymes are considered as important indicators of the functional integrity of hepatocellular membranes, Pari and Amali, (2005). According to Adeneye et al. (2006) elevation in these enzymes is related to hepatic and heart disorders. Mild reduction in levels of both the liver and the kidney enzymes in treated groups could indicate that D. stramonium leaves possesses hepato-protective, nephronprotective and cardio-protective properties. Ogunmoyole et al. (2019) reported that D. stramonium extract at 200mg/kg dose, generally caused a significant decrease in ALT, AST, ALP and MDA in selected tissue homogenates while these parameters increased significantly in the serum relative to the control group. Abbas (2013) and Başaran et al. (2018), in their separate studies reported low level of serum AST and ALP relative to the control animals and concluded that extract of *D. stramonium* are hepatoprotective at 200mg/kg. The decrease in ALP with the reference range showed that the D. stramonium extract does not cause harmful effect, Clementine and Tar, (2010). However, the result of this study did not agree with the work done by Jaroslaw et al. (2009) and Udem et al. (2009), who in separate studies reported significant increase in the serum ALP by D. stramonium extract leading to biliary obstruction and heart failure.

Cholesterol is synthesized in the liver, and it is important for cell wall structure, cell immunity, steroid hormones production by the adrenal glands and gonads, membrane strength, nerve protection and formation of bile acids, which accounts for about 80 %. Cholesterol is also necessary for formation of antibodies and enzymes and is also used to evaluate risk for atherosclerosis (Soetan *et al.*, 2013). Cholesterol is among the prominent lipids responsible for membrane integrity (Bouzidi et al., 2011). Total cholesterol in this study was decreased in the serum of experimental animals treated with the 200 mg/k dose of the extract relative to the control. The lowering of cholesterol level in the sera of the experimental rats administered with the 200 mg/kg dose might suggest that the ingredients contained in the leaf extract at the medium dose was capable of inhibiting the activities of hepatic lipogenic and cholesterogenic enzymes, such as malic enzyme, fatty acid synthase, glucose 6-phosphate dehydrogenase and HMG-CoA reductase (Vega et al., 2003) which are all required for cholesterol synthesis. This cholesterol lowering action of the crude extract might be attributed to the presence of a bioactive phyto-constituents, i.e. β-sitosterol (Berkov et al., 2006: Swathi *et al.*, 2012), and N.N'-Bis(salicylidene)-3,3'-bis(aminopropyl)aminocobalt(II) contained in the leaves as identified by Otuokere et al., (2020), which is an Antitopoisomerase-II, Casein-Kinase-II-Inhibitor, Topoisomerase-II-Inhibitor known to decrease cholesterol synthesis in the liver. This finding partially agreed with the studies of Tariq et al. (1989) and Gharaibeh et al. (1988), but completely agreed with recent studies of Rasekh et al. (2001); Couladis et al. (2003) and Ogunmoyole et al. (2019), who in their separate studies reported that the crude extract of D. stramonium leaves at moderate doses has a significant cholesterol lowering action.

Triglycerides are form of fat and a major source of energy for the body and are made up of three fatty acids attached to a glycerol molecule, hence the term triglycerides (Purves et al., 2003). Triglyceride is synthesized in the liver, and the result obtained in this study demonstrated the ability of D. stramonium to influence liver metabolism towards decreasing the synthesis of lipids (hypolipidemic potential) in the 100 and 200 mg/kg doses. The low levels of serum triglyceride may be due to a number of factors such as the decreased availability of fatty acids for esterification (Bopanna et al., 2017), increased catabolism of LDL, enhancing of tissues lipases, activation of acetyl-CoA carboxylase (McCarty, 2021) and production of triglycerides precursors such acetyl-CoA and glycerol phosphate (Campillo et al., 2014). The result obtained in this study shows D. stramonium to have hypolipidemic potential. HDL-cholesterol is otherwise called 'good cholesterol' which returns cholesterol to the liver where it is converted into bile and subsequently removed from the body, Nwanjo, (2005). It is important to note that the serum level of HDL-cholesterol and LDL were within the normal range at both lowest and highest doses used in this study, whereas, a noticeable increase and decrease in HDL-cholesterol and LDL, respectively, were observed in the sera of the rats treated with the 200 mg/kg dose suggesting hepato- and cardio-protection of D. stramonium crude extract at moderate dose. This observation could explain why D. stramonium leaf extract had no alteration on the lipid profile, hence, do not compromise the membrane integrity (its stability and

fluidity) and their functions. The Very Low Density Lipoprotein Cholesterol (VLDLc) serum levels at 100 and 200 mg/kg doses were significantly higher than the normal value compared with the serum level obtained in the highest dose. This also gave support to the hepatoand cardio-protective ability of the crude extract. The outcome of this result strongly agreed with that of Ghasi *et al.* (2000); Boumba *et al.* (2005); Nandave *et al.* (2009) and Başaran *et al.* (2018).

CONCLUSION

D.stramonium had no significant effect on the evaluated blood parameters (Hb, PCV and RBC) of male Wistar rats used, but however, improved the leucocytes counts and most differentials (lymphocytes, neutrophils and monocytes) at lower doses much better than the high dose, implying an improved index of immune function. The decreases in the kidney and liver enzymes suggested hepato-protective, nephron-protective and cardioprotective properties of the leaf extract. The study also revealed a lowering effect of bad cholesterol (TC), while increasing the good cholesterol (HDL-c) at lower doses of the extract, implying that the extract even at the low dose of 100 mg/kg, may have inhibited the activities of hepatic lipogenic and cholesterogenic enzymes, required for bad cholesterol synthesis. Therefore, apart from the excitatory effect which occurred in a dose dependent manner, it can be concluded that the extract of Datura stramonium exhibited desired pharmacological and biochemical activities at lower doses, than at higher doses.

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REFERENCES

- 1. Abbas, D. A., (2013). Analgesiac, anti-Inflammatory and antidiarrhoeal effects of *Datura stramonium* hydroalcoholic leaves extract in mice, International Journal of Recent Research and Applied Studies, 14(1): 22–29.
- 2. Adenkola, A. Y. and Oluremi, O. I. A. (2014). Erythrocyte Osmotic Fragility and Excitability Score in Rabbit fed *Hibiscus sabdariffa* in Graded Level. Nigerian Journal of Physiological Society, 29: 113-117.
- 3. Allain CC, Poon LS, Chan CS, Richmond W, Fu PC (1974) enzymatic determination of total Cholesterol. *Clinical Chemistry*, 20(4): 470-475.
- Ayo, J. O., Minka, N. S. and Mamman, M. (2006). Excitability scores of goats administered ascorbic acid and transported during hot-dry conditions. *Journal of Veterinary Science*, 7: 127 - 131.
- 5. Babson AL, Greeley S.J, Coleman CM, Philips GE (1966) Phenolphthalein monophoshpate as a substrate for serum alkaline phosphate. *Clinical Chemistry*, 12: 482-490.

- Başaran, S., Dündar, G. and Ogun, M. N. (2018). Datura stramonium poisoning case with Wernicke aphasia-like symptoms: Case report. Kocaeli Medicine Journal, 7(1): 83–86.
- Batool, R., Khan, M. R. and Majid, M. (2017). *Euphorbia dracunculoidesL.* abrogates carbon tetrachloride induced liver and DNA damage in rats. *BMC Complementary and Alternative Medicine*, 17(1): 223.
- Bopanna, K. H., Kannan, J., Gadgil, S., Balaraman, E. R. and Rathore, S. P. (2017). *Indian Journal of Pharmacology*, 29: 162-167.
- Boumba, A., Mitselou, A., and Vougiouklakis, T. (2005). Fatal poisoning from ingestion of *Datura* stramoniumseeds. Veterinary and Human Toxicology, 46: 81–82.
- Bouzidi, A., Mahdeb, N. and Kara, N. (2011). Toxicity studies of alkaloids of seeds of *Datura* stramonium and synthesis alkaloids in male rats. *Journal of Medicinal Plants Research*, 5(15): 3421-3431.
- 11. Campillo, J. C., Torres, M. D., Dominguez, E., Romero, A. and Perez, C. (2014). *Diabetologia*, 37(1): 213.
- Chavhan, S. A., Kadam, S. D., Shinde, S. A. and Sapkal, P. N. (2018). Pharmacognostic review on Datura. International Journal of Pharmacognosy and Chinese Medicine, 2(4): 145–9.
- 13. Clementine, Y. F. and Tar, C. (2010). Liver function tests (LFTs). *Lab. Insights*, 19(1): 1–3.
- 14. Das S, Kumar P, Basu SP. (2012) Review article on phytoconstituents and therapeutic potentials of *Datura stramonium* linn. *J Drug Del Therap*; 2(3): 4–7.
- 15. Doumas BT and Peters T Jr. (1997). Serum and urine albumin: a progress report on their measurement and clinical significance. *Clinica Chimica Acta*, 258(1): 3 20.
- Doumas BT, Perry BW, Sasse EA, Straumfjord Jr. J.V. (1973) Standardization in bilirubin assays: evaluation of selected methods and stability of bilirubin solutions. Clinical Chemistry, 19: 984–993.
- Essas, M. M., Subramanian, P., Suthakar, G., Manivasagam, T., Dakshayani, K. B., Sivaperumai, R., Subash, S. and Vinothini, G. (2006). Influence of *Hibiscus sabdariffa (Gongura)* on the levels of circiulatory lipid peroxidation producers and liver marker enzymes in experimental hyperammonemia. *Journal of Applied Biomedicine*, 49(1): 53-5.
- Fatoba, T, A., Adeloye, A. A. and Soladoye, A. O. (2013). Effect of *Datura stramonium* seed extracts on haematological parameters of West African Dwarf (WAD) bucks. *European Journal of Experimental Biology*, 3(4): 1-6.
- 19. Fereshteh Eftekhar, Morteza Yousefzadi, V Tafakori (2004). Antimicrobial activity of Datura innoxia and *Datura stramonium*. Fitoterapia, 76(1): 118-20.
- Freye, E. (2010). Pharmacology and Abuse of Cocaine, Amphetamines, Ecstasy and Related Designer Drugs: A comprehensive review on their

mode of action, treatment of abuse and intoxication. In Freye, E. Toxicity of *Datura stramonium*. Springer Netherlands, 217–218.

- Gidado, A., Zainab, A. A., Hadiza, M. U., Serah, D. P., Anas, H. Y. and Milala, M. A. (2007). Toxicity studies of ethanol extract of the leaves of *Datura stramonium* in rats. *African Journal of Biotechnology*, 6(8): 1012-1015.
- 22. Glatstein, Miguel; Alabdulrazzaq, Fatoumah; Scolnik, Dennis (2016). "Belladonna Alkaloid Intoxication". *American Journal of Therapeutics*, 23(1): e74–e77.
- 23. Grieve, Maud (1971). A Modern Herbal: The Medicinal, Culinary, Cosmetic and Economic Properties, Cultivation and Folk-lore of Herbs, Grasses, Fungi, Shrubs, & Trees with All Their Modern Scientific Uses, Volume 2. Dover Publications. p. 804.
- 24. Haegi I, 1976. A taxonomic account of Datura L. Solanaceae in Australia with a note on Brugmansia Pers. Australian Journal of Botany, 24: 415-435.
- Hadkins ES, Bye R, Brandenburg WA, Jarvis CE, (1997). Typification of Linnaean Datura names (Solanaceae). Botanical Journal of the Linnean Society, 125(4): 295-308.
- Halpern, J. H. (2004). Hallucinogens and dissociative agent naturally growing in the United States. *Journal of Pharmacology Therapy*, 102: 131-138.
- 27. Higgins T, Beutler E & Doumas BT (2008) Bilirubin. Analytical Methodology – Serum bilirubin. In: Burtis, CA, Ashwood ER & Bruns DE (Eds.), Tietz Fundamentals of Clinical Chemistry. 6th ed. Saunders Elsevier, Missouri, pp. 524 - 525. Ihedioha J.I. and Chineme C. N. (2005) Fundamentals of Systemic Veterinary Pathology (Vol 2) Great AP Express Publishers Ltd, Nsukka, Nigeria, 206-207.
- 29. Ibiam, O. F. A., Kalu, E. N., Kanayochukwu, L. U. and Akpo, S. O. (2017). Proximate and Phytochemical Analysis of the Fruits and Leaves of *Datura Stramonium* L, and the Effect of Fungi Associated with Foliar Blight on Them. *Austin Journal of Biotechnology & Bioengineering*, 4(1): 1-5.
- 30. Igwe, K.K., Ikpeazu O.V., Ezeja M.I., (2020). Evaluation of the Stem Back Extract *Picralima nitida* for Antinociceptive property. *International Journal of Biochemistry Research and Review*, 2020; 29(8): 108-113.
- Igwe K.K., Nwakundu O.N., Ijioma S.N., Madubuike A.J., Achi N.K., (2016). Screening for Secondary metabolites in *Huru crepitans* bark ethanol extract using GCMS analysis: A preliminary study approach. *Journal of Science and Technology Advances*, 1(2): 64-71.
- 32. Ijeh I.I., Igwe K.K., Ejike C.E.C.C, (2009) Effect of administration of aqueous extracts of *Vernonia amygdalina*. Del. Leaves to Guinea Pig dams on milk production and contraction of the mammary

gland and uterus. *African Journal of Traditional, Complementary and Alternative Medicines,* 439-440.

- Ikpeazu O.V., Otuekere I.E., Igwe K.K., (2017). Preliminary studies on the secondary metabolites of *Buchholzia coriacea* (Wonderful Kola) seed ethanol extract by GCMS analysis. *Intern J Res Eng. Appl*, 7(3): 17-26.
- Ikpeazu O.V., and Igwe K.K., (2023). The Biochemistry of Environmental Pollution. World Journal of Pharmacological and Life Science, 9(7): 1-20.
- Isaac, L. J., Abah, G., Akpan, B. and Ekaette, I. U. (2013). Haematological properties of different breeds and sexes of rabbits. *Proceedings of the 18th Annual Conference of Animal Science Association of Nigeria*, 24-27.
- 36. Iwuji, T. C. and Herbert, U. (2012). Haematological and serum biochemical characteristics of rabbit bucks fed diets containing garcimiola kola seed meal. *Proceedings of 37th Annual Conference of Nigerian Society for Animal Production*, 87-89.
- Jaroslaw, S., Armand, M., Gizowska, M., Marcinek, M., Sasim, E. and Szafran, E. (2009). Ceramicin-Polymer versus Polymer-In-Ceramic polymeric electrolytes—*A novel approach. Journal of Power Sources*, 194: 66–72.
- Johnston, J. K. and Morris, D. D. (1996). Alterations in blood proteins. In B. P. Smith (Ed.), International Animal Medicine (2nd ed.). USA: Mosby Publishers.
- Johnson AM (2008) Amino acids and proteins. In: Burtis CA, Ashwood ER & Bruns DE (Eds.), *Tietz Fundamentals of Clinical Chemistry*. 6th ed. Saunders Elsevier, Missouri, 206–316.
- 40. Kabir, M., Akpa, G. N., Nwagu, B. I., Adeyinka, I. A. and Bello, U. I. (2011). Sexual dimorphism, breed and age characteristics of rabbits in Zaria, Nigeria. *Proceedings of the 16th Annual Conference of Animal Science Association of Nigeria*, 133-137.
- Kailemia, M. J., Ruhaak, L. R., Lebrilla, C. B. and Amster, I. J. (2014). Oligosaccharide analysis by mass spectrometry: A review of recent developments. *Analytical Chemistry*, 86(1): 196–212.
- 42. Khan, B. R. and Majid, M. (2017). Euphorbia dracunculoides L. abrogates carbon tetrachloride induced liver and DNA damage in rats. BMC Complementary and Alternative Medicine, 17(1): 223.
- 43. Khan J, Khan R, Qureshi R. A. (2013). Ethnobotanical Study of Commonly Used Weeds of District Bannu, Khyber Pakhtunkhwa (Pakistan). *Journal of Medicinal Plants Studies;* 1(2): 1-6.
- 44. Khan SW, Khatoon S. Pak J Bot (2008). Ethnobotanical studies on some useful herbs of Haramosh and Bugrote valleys in Gilgit, 40(1): 43-58.
- 45. Khudhair A.M and Abed AL Ani (2015) The protective role of *Datura stramonium* leaves ethanolic extract against acute carbaryl toxicity in

rats. Int J of Biomedical and Adv Research, 6: 400–405.

- 46. Kirtikar JD, Basu BD, Lalit Mohan Basu, Allahabad. (1994) Indian Medicinal Plants, 1229–1231.
- 47. Klein, B., P. A., Babson, A. L. (1960 Northern areas of Pakistan;), Rapid method for the quantitative determination of serum alkaline phosphatase. *Clinical Chemistry*, 6: 269 275.
- Kurnal NA, Yalcin SCC. (2009). Acaricidal, repellent and oviposition deterrent activities of *Datura stramonium* L. against adult Tetranychus urticae (Koch). *J Pest Sci*; 14: 54-57.
- 49. Lubran M M (1978) The measurement of serum proteins by Biuret method. Annals of *Clinical Laboratory Science*, 8(2): 106-110.
- 50. McCarty, M. F. (2021). *Medical Hypotheses*, 56: 314-317.
- 51. Njoroge GN. (2012). Traditional Medicinal Plants in Two Urban Areas in Kenya (Thika and Nairobi): Diversity of traded species and conservation concerns. *Ethnobotany Research & Applications*, 9: 329-338.
- 52. Nwanjo, H. U. (2005). Efficacy of aqueous leaf extract of *Vernonia amygdalina* on plasma lipoprotein and oxidative status in diabetic rat models. *Nigerian Journal of Physiological Sciences*, 20: 39–42.
- Ogunmoyole, T., Adeyeye, R. I., Olatilu, B. O., Akande, A. O. and Agunbiade, O. J. (2019). Multiple organ toxicity of *Datura stramonium* seed extracts. *Toxicology Reports*, 6: 983–989.
- 54. Okunlola, D. O., Olorunisomo, A. O., Aderinola, A. O., Agboola, A. S. and Omole, O. G. (2012). Haematology and serum quality of red Sokoto goats fed Baobab (*Adansoniadigitata*) fruit meal as supplement to guinea grass (*Panicum maximum*). Proceedings of the 17th Annual Conference of Animal Science Association of Nigeria, 427-433.
- 55. Ologundudu, A., Ologundudu, A. O., Olabode, I. A. and Obi, F. O. (2009). Effect of *Hibiscus sabdariffa* anthocyanins on 2, 4-dinitrophenylhydrazine-induced hematotoxicity in rabbits. *African Journal of Biochemistry Research*, 3(4): 140 144.
- Otuokere, I. E., Igwe K. K. and Ikpeazu. O. V. (2020). GC–MS analysis of phytochemicals present in ethanol extract of *Datura stramonium* leaves. *International Research Journal of Modernization in Engineering, Technology and Science*, 2(7): 869–876.
- 57. Oyawoye, B. M. and Ogunkunle, H. N. (2004). Biochemical and haematological reference values in normal experimental animals. New York: Masson, 212-218.
- 58. Pari, L. and Amali, D. R. (2005). Protective role of tetrahydrocurcumin (THC) an active principle of turmeric on chloroquine induced hepatotoxicity in rats. *Journal of Pharmacy & Pharmaceutical Sciences*, 8(1): 115–123.

- 59. Parsons WT, Cuthbertson EG, (1992). Noxious Weeds of Australia. Melbourne, Australia: *Inkata Press*, 692 pp.
- Purves, W. K., Sadava, D., Orians, G. H. and Heller, H. C. (2003). *Life: The science of Biology* (7th edition). *Sinauer Associates and W. H. Freeman*. Pp. 954.
- 61. Rahmatullah M, Islam R, Kabir Z, Rashid H, Jahan R, Begum R. (2010). Folk Medicinal Practices in Vasu Bihar Village, Bogra District, Bangladesh. *American-Eurasian Journal of Sustainable Agriculture;* 4(1): 86-93.
- Rahmatullah M, Das AK, Mollik AH, Jahan R, Khan M, Rahman T. (2009). An Ethnomedicinal survey of Dhamrai sub-district in Dhaka district, Bangladesh. *American1Eurasian Journal of Sustainable Agriculture*, 3(4): 881-888.
- Ray F. Evert, Susan E. Eichhorn. Raven Biology of plants (2013). W. H. Freeman and company publisherss 8th Edition Chapter 2, page 30.
- 64. Reitman S., Frankel S. (1957) A colorimetric method for determination of serum glutamic oxaloacetic and glutamic pyruvic transaminases. *Amercan Journal of Clinical Pathology*, 28: 56-62.
- 65. Savithramma N, Sulochana C, Rao KN. (2007). Ethnobotanical survey of plants used to treat asthma in Andhra Pradesh, *India J Ethnopharmacol*, 113(1): 54-61.
- 66. Sharma MC, Sharma S. (2010). Phytochemical, preliminary pharmacognostical and antimicrobial evaluation of combined crude aqueous extract. *Int J Microbiol Res.*, 1(3): 166-170.
- 67. Soetan, K. O., Akinrinde, A. S. and Ajibade, T. O. (2013). Preliminary studies on the haematological parameters of cockerels fed raw and processed guinea corn (*Sorghum bicolor*). *Proceedings of 38th Annual Conference of Nigerian Society for Animal Production*, 49-52.
- 68. Taha SA, Mahdi AW. (1984). *Datura* intoxication in Riyadh. Trans R Soc Trop Med Hgy; 78: 134-135.
- 69. Tee, P. L., Yusof, S. and Mohammed, S. (2002). Antioxidative properties of roselle (*Hibiscus* sabdariffa L.) in linoleic acid model system. *Nutrition and Food Science*, 32(1): 17-20.
- Udem, S. C., O. Obidoa, O. and Asuzu, I. U. (2009). Acute and chronic toxicity studies of *Erythrina* senegalensis stem bark extract in mice. *Complementary Clinical and Pathology*, 19: 275–282.
- 71. Vega, G. L., Weiner, M. F. and Lipton, A. M. (2003). Archive of Neurology, 60: 510-515.
- Vijendra N, Kumar K. P. (2010). Traditional knowledge on ethno1medicinal uses prevailing in tribal pockets of Chhindwara and Betul Districts, Madhya Pradesh, *India. African Journal of Pharmacy and Pharmacology*, 4(9): 662-670.
- 73. Wang, C. J., Wang, J. M., Lin, W. L., Chu, C. Y., Chou, F. P. and Tseng, T. H. (2000). Protective effect of Hibiscus anthocyanins against tert-butyl

hydroperoxide-induced hepatic toxicity in rats. *Food Chemistry and Toxicology*, 38: 411- 416.

- 74. Whitby L.G., Percy-RobbI.W., Smith A.F., (1984) 3rd Edition. Lecture notes on clinical chemistry, *Blackwell Scientific Publications*.
- 75. Ziad F. Issa MD, Douglas P. Zipes MD. (2019). Clinical Arrhythnology and Electrophysiology (Third Edition).