

## LARVICIDAL EFFECT OF TRIDAX PROCUMBENS AND AZADIRACHTA INDICA PLANT EXTRACT AGAINST AEDES AEGYPTI AND CULEX QUINQUEFASCIATUS

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### ABSTRACT

*Aedes aegypti* is a vector for transmitting several tropical fevers which include dengue virus and the virus spread to other disease including mosquito salivary gland. Whereas *Culex quinquefasciatus* found in eutropic water of artificial containers and in open pond, diectuss train of sewage. It is important to develop the non-toxic and inexpensive method to conclude the mosquito larvae using medicinal important plant extract such as *Azadirachta indica* and *Tridax procumbens*. The larvae were categorized based on their prey size such as 4<sup>th</sup> instar larvae of both species. *Tridaax procumbens* revealed larval growth incubation were as *Azadirachta india* extracts as bio-pesticides and as natural insecticides proved to be an efficient and inexpensive and non-toxic method for control of mosquito. The percentage of prey killed of *culex quinquefasciatus* at 5% concentration revealed 82% at 10mg/ml and At different concentration of the *Azadirachta indica* leaf observed for the period of 24 hours exposed to the prey density of 25 showed an increased percentage of prey killed of 90%. The larvae mortality rate at 10 mg/ml of leaf extract found exhibit at maximum activity against *Aedes agypti*. The results demonstrate and excellent activity which contributes to rare opportunity of natural control. Similary, *Tridaax procumbens* showed increased the higher larval (*culex quinquefasciatus*) mortality was found to 62% at the concentration of 10mg/ml and *Aedes agypti* percentage of prey killed was found to be 74% at the concentration of 10mg/ml.

**KEYWORDS:** *Culex quinquefasciatus*, *Aedes aegypti*, *Tridax procumbens* and *Azadirachta indica*.

### INTRODUCTION

Mosquitoes play a major role in the transmission of dengue fever, yellow fever, malaria, filariasis. Mosquito are one of the most medically significant vectors, and they transmit parasites and pathogens, which continue to have a devastating impact on human beings and other animals (Elumalai *et al.*, 2013 a, b). Some mosquito species like *Culex* and *Aedes* are the vectors for the pathogens of various disease and contribute significantly to poverty and social debility in tropical countries (Jiang *et al.*, 2009).

*Culex quinquefasciatus* is a peridomestic mosquito seldom found far from human residence or activity, and readily feeds on avian, mammalian or human hosts. The larvae are typically found in the eutrophic water of artificial containers or man-made impoundments including open ponds, ditches and drains containing human or animal sewage. As such, *Culex quinquefasciatus* was uniquely adapted to the environs of historical sailing ships outfitted for long voyages where

polluted water and livestock were common. Since adult mosquitoes can fly short distances to shore (Subra, 1981; LaPointe, 2008) and immature forms could be carried ashore in water casks taken to be refilled (Hardy, 1960), it is likely that this mosquito was spread worldwide by commercial sailing vessels involved in the Atlantic slave trade, Old China trade and American whale oil industry between the 17 and 19<sup>th</sup> centuries (Lounibos, 2002).

Dengue virus is primarily transmitted by *Aedes* mosquitoes, particularly *A. aegypti*. *Aedes aegypti* is a vector for transmitting several tropical fever and only the female bites for blood which she need to mature her eggs (Hahn *et al.*, 2001). They typically bite during the early morning and in the evening, but they may bite and thus spread infection at any time of day WHO (2009). An infection can be acquired via a single bite. A female mosquito that takes a blood meal from a person infected with dengue fever, during the initial 2- to 10-day febrile period, becomes itself infected with the

virus in the cells lining its gut **Georgiev, Vassil (2009)**.

About 8–10 days later, the virus spreads to other tissues including the mosquito's salivary glands and is subsequently released into its saliva. The virus seems to have no detrimental effect on the mosquito, which remains infected for life. *Aedes aegypti* is particularly involved, as it prefers to lay its eggs in artificial water containers, to live in close proximity to humans, and to feed on people rather than other vertebrates. **Gubler (2010)**.

#### ***Azadirachtaindica* (Neem)**

Every part of *Azadirachta indica* (Neem) has been advocated to possess medicinal properties. **Pruthi (1937)** first proved scientifically the insecticidal effect of neem. Naturally occurring bio pesticides could be an alternative to chemical pesticides. (**Abdelouaheb *et al.*, 2009**). *Azadirachta indica* is a natural insecticide and is nonhazardous to man and other mammals (**Oudegans, 1991**). Therefore simple nonhazardous and inexpensive methods of extraction should be developed to enable practical use of neem (**Feuerhake, 1984**). The aims and objectives of the present work is to develop a simple inexpensive and nontoxic method for the control of mosquito larvae by *Azadirachta indica* (neem), which can be applied easily by the ordinary man without use of costly spraying equipment.

#### ***Tridax procumbens***

*Tridaxprocumbens* is commonly used in Indian traditional medicine as anticoagulant, antifungal and insect repellent, in bronchial catarrh, diarrhoea and dysentery (**Ali, *et al.*, 2001**). The entire plant is used for the treatment of malaria, leishmaniasis, vaginitis, dysentery and gastrointestinal disorders (**Caceres *et al.*, 1994**).

*Tridaxprocumbens* showed growth inhibitory and juvenile hormone mimicing activity to the treated larvae of *Culex quinquefasciatus*. It has been extensively used in Indian traditional medicine for wound healing, as anticoagulant, antifungal and insect repellent, in diarrhea and dysentery. (**Ali *et al.*, 2001**).

### **MATERIALS AND METHODS**

#### **Collection of prey (*AedesAegypti* and *Culex quinquefasciatus*)**

Larvae of *Aedes aegypti* were taken as the prey item throughout the investigation. *Aedes aegypti* and *Culex quinquefasciatus* egg craft were collected from ICMR, Chinnachokkikulam, Madurai, India. There were brought to the laboratory and culture to a plastic bucket containing clean water. The bucket was covered with the mosquito mesh. Larvae were categorized based on their prey size as large and small class the covered with nylon mesh. They were fed with the dog biscuits and yeast powder in the ration of 3:1 as food.

#### **Collection of Plant**

Fresh leaves of *Tridax procumbens* and *Azadirachta indica* were collected from Government Arts college, Melur, Madurai District, Tamil Nadu.

#### **Preparation of the Leaves extract**

The leaves of the *Tridax procumbens* and *Azadirachta indica* were shade dried 18°C ground and sieved to get fine powdered from which the extracts were prepared. Acetone extracts of the plant was obtained by taking 25g dried leaf powder with 250 ml of acetone and kept for 24hrs with periodic shaking.

The leaves were dried to a constant weight at in an enclosed air conditioned research laboratory. The dried leaves were blended to powder to increase the surface area for extraction.

Soxhelt method of extraction was carried out. In a conventional soxhlet system, the powdered leaf material is placed in a thimble-holder and filled with condensed fresh solvent from a distillation flask. When the liquid reaches the overflow level, a siphon aspirates the solution of the thimble-holder and unloads it back into the distillation flask, carrying extract solutes into the bulk liquid. In the solvent passes back into the plant solid bed. The operation is repeated until complete extraction is achieved (**Wang and Weller, 2006**).

#### **Phytochemical Analysis**

Phytochemical test were carried out on the aqueous extract of the powdered leaves from soxhlet using standard procedures.

#### **Larvicidal Bioassay**

The larvicidal activity was assessed using Acetone-leaf extract of *Tridaxprocumbens* and *Azadirachtaindica* by the procedure of **WHO (1996)** with some modification and as per the method of **Rahuman *et al.*, (2000)**, For the stock (50 mg/ml) preparation 5ml of desired plant extract was taken and 95 ml of dechlorinated tap water. For the bioassay test 25 larvae/concentration were used for all the experiment. For mortality studies, 25 larvae each of 4<sup>th</sup> instar larvae introduced in different concentration (2, 4, 6, 8 and 10 mg/ml) of leaf extract were added. Percentage of mortality was observed in acetone leaf extract of *Tridaxprocumbens* and *Azadirachtaindica* against *Aedesaegypti* and *Culex quinquefasciatus*. The number of dead mosquito larvae was counted after 24 hrs of exposure, and the percentage mortality was reported from five replicates. The percentage of mortality was calculated by (No of larvae dead / Total no. larvae)\*100. Corrected mortality was calculated by **Abbott (1925)**.

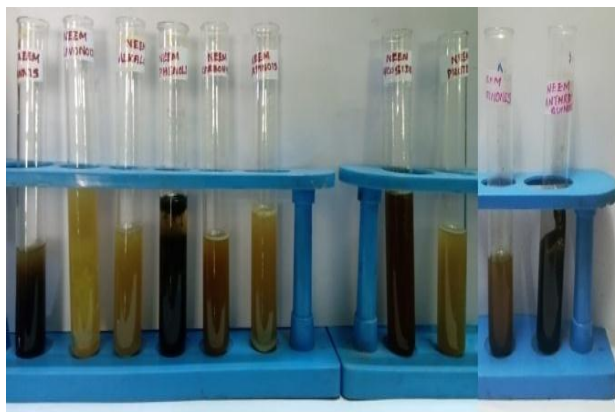


Fig. 1: Phytochemical Analysis of *Azadirachta indica* Extract.



Fig. 2: Phytochemical Analysis of *Tridax procumbens* Extract.

### Statistical analysis

From the results Statistical analysis such as Mean, Median, Mode, Standard Deviation and Percentage was performed using from MS Excel 2007. The larvicidal mortality were subjected to probit analysis (Chandra *et al.*, 2014) for compute lethal concentration of 50% (LC<sub>50</sub>) values were calculated by using the Regression Equation for hasty determination of lethal concentration.

## RESULTS

### Larvicidal activity of *Tridax procumbens* and *Azadirachta indica* leaf extract against *Aedes aegypti* and *Culex quinquefasciatus*

The percentage mortality and LC<sub>50</sub> rates of the fourth instar larvae of *Culex quinquefasciatus* and *Aedes Aegypti* exposed to the acetone extracts of *Azadirachta indica* presented in **Table 2**.

The percentage of prey killed of the adult mortality of *Culex quinquefasciatus* at 5% concentration of leaf extract in a prey density of 25 showed an increased percentage of prey killed with 82% at 10mg/ml. The result of the study showed that plant based components of *Azadirachta indica* leaf extract demonstrated an excellent activity which contributes to an increased opportunity for the natural control.

At different concentration of the *Azadirachta indica* leaf observed for the period of 24 hours exposed to the prey density of 25 showed an increased percentage of prey killed of 90%. The larvae mortality rate at 10 mg/ml of leaf extract found exhibit at maximum activity against *Aedes aegypti*.

Results on the larvicidal activity of the medicinally important plant *Tridax procumbens* as plant extract have the potential to be used as eco-friendly approach to control the mosquito. The effect of larvicidal activity of different prey density of *Aedes aegypti* treated with the concentration of the *Tridax procumbens* leaf extract were depicted in the **Table 3**.

The higher larval mortality was found to be 62% at the prey density of 25 was recorded at the concentration of 10mg/ml. Among the different prey density, the larvicidal efficiency of the leaf extract *Tridax procumbens* at a lower prey density of 25 exhibited better high activity of 62%.

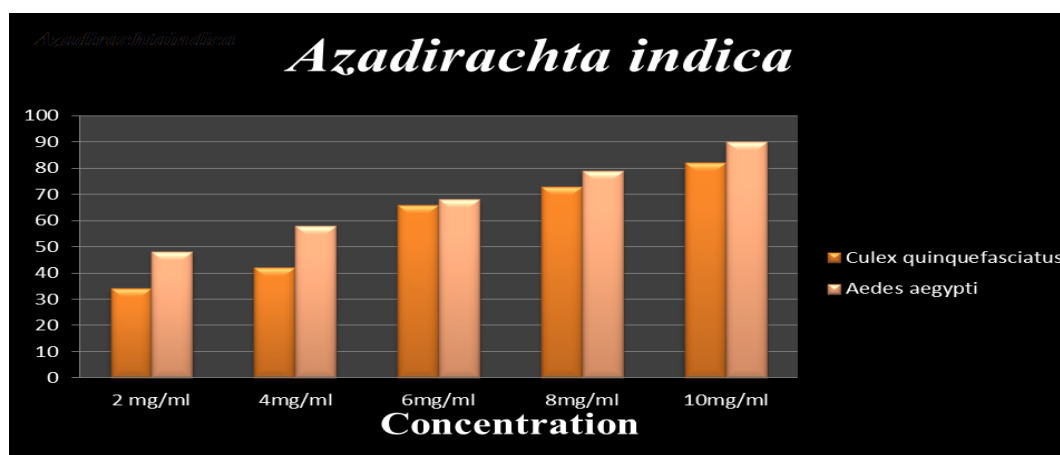
The larvicidal activity of *Aedes aegypti* have been tested with plant extract *Tridax procumbens*. It is presumed the extract of *Tridax Procumbens* containing essential oil which source as effective repellent activity against larvae of *Aedes aegypti* at the prey density of 25. The percentage of prey killed was found to be 74% at the concentration of 10mg/ml.

Table 1: Phytochemical Analysis of Plant Extracts.

Phytoconstituents	Aqueous Extract	
	<i>Azadirachta indica</i>	<i>Tridax procumbens</i>
Tannins	+	+
Flavonoids	+	+
Alkaloid	-	-
Phenols	+	+
Protein	-	-
Carbohydrate	+	-
Terpinoids	-	-
Glycosides	-	-
Quinones	+	+
Anthro-quinones	-	-

Table 2: Larvicidal activity of *Azadirachta indica* leaves against *Aedes aegypti* and *Culex quinquefasciatus*.

plant	Mosquito	Prey Density	Concentration (mg/ml)	% of Mortality	LC <sub>50</sub>
<i>Azadirachta indica</i>	<i>Culex quinquefasciatus</i>	25	2	34	5.79
			4	42	
			6	66	
			8	73	
			10	82	
	<i>Aedes aegypti</i>	25	2	48	2.59
			4	58	
			6	68	
			8	79	
			10	90	

Figure 4: Larvicidal Activity of *Aedes aegypti* and *Culex quinquefasciatus* in *Azadirachta indica* Extract.Table 3: Larvicidal activity of *Tridax Procumbens* leaves against *Aedes aegypti* and *Culex quinquefasciatus*.

plant	Mosquito	Prey Density	Concentration (mg/ml)	% of Mortality	LC <sub>50</sub>
<i>Tridax Procumbens</i>	<i>Culex quinquefasciatus</i>	25	2	26	6.31
			4	38	
			6	48	
			8	55	
			10	62	
	<i>Aedes aegypti</i>	25	2	38	3.63
			4	50	
			6	60	
			8	69	
			10	74	

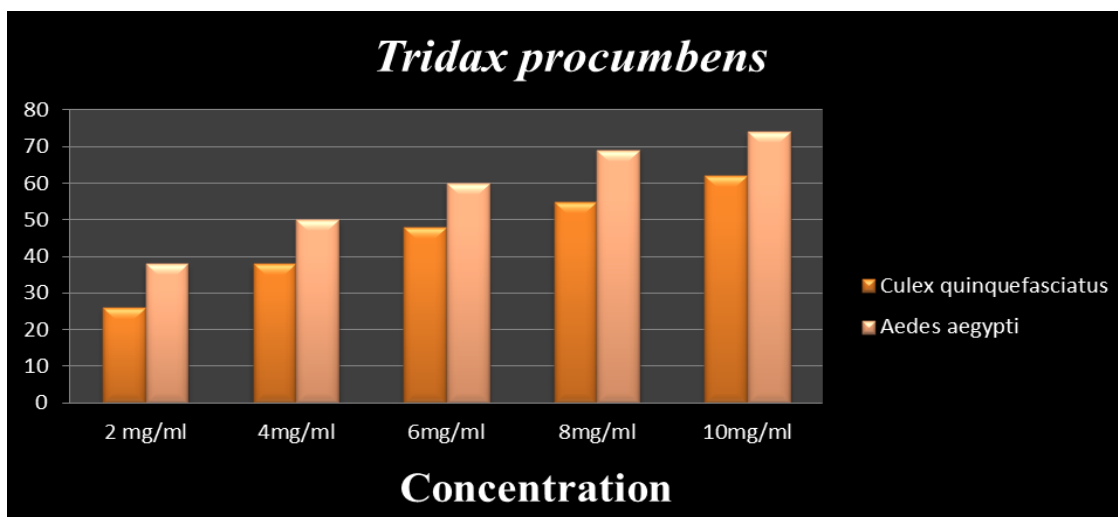


Figure 5: Larvicidal Activity of *Aedes aegypti* and *Culex quinquefasciatus* in *Tridax procumbens* Extract.

## DISCUSSION

Larvicidal activity of *Azadirachta indica* against various species of mosquitoes has been observed by various researchers (Wandscheer, 2004; Chavan, 1984; Virendra *et al.*, 2009; Aliero *et al.*, 2003; Vatandoost and Vaziri, 2004; Abdelouaheb *et al.*, 2009; Senthil *et al.*, 2006). The most important of these bioactive constituents of plant are alkaloids, tannins, flavonoids and phenolic compounds. Presently many scientist and organizations are search of traditional remedies as alternative medicines. It has been estimated that about 25% of all prescribed medicines today are substances derived from plants (Puri, 1999).

The extracts produced some abnormalities in larvae. Larval pupal intermediates were observed. Partially emerged adults showed crumpled legs and entangled in pupation. All these abnormalities were also reported by Naqvi (1987). Several studies have focused on natural products for controlling *Aedes* mosquitoes as insecticides and larvicides, but with varied results. [14,15,16,17,18,1]

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The phytoconstituents of *Tridax procumbens* responsible for effect on cellular and hormonal functions in mice

were attempted to explore. The *in vitro* (phagocytosis) and *in vivo* (haemagglutination and delayed hypersensitivity) were used to study the effect of extract and fraction on the cellular and hormonal immunity. The results obtained indicate the ability of flavonoidal and saponin fraction of *Tridax procumbens* to modulate both cell mediated and the hormonal components of the immune system and explored the phytoconstituents responsible for immunomodulatory potential of *Tridax procumbens*. (Agarwal *et al.*, 2010).

The leaves essential oils of *T. procumbens* are more effective repellents activity at 6 per cent concentration against *An. Stephensi* (Rajkumar and Jebanesan, 2007).

Leaf acetone extract of *Tridax procumbens* against the larvae of *An. subpictus* (LC<sub>50</sub>=93.80, 39.98, and 51.57 mg/l; LC<sub>90</sub>=524.90, 145.70, and 226.56 mg/l) respectively. Ethyl acetate extract of *Tridax procumbens* have the potential to be used as an ecofriendly approach for the control of the *An. subpictus*, and *Cx. tritaeniorhynchus*. (Kamaraj *et al.*, 2011).

## CONCLUSION

In this study, the acetone extracts of *Tridax procumbens* and *Azadirachta indica* leaves showed potential larvicidal activities against *Aedes aegypti* and *Culex quinquefasciatus*. The biological activity of the experimental aqueous *Tridax procumbens* plant extracts may be recognized to the presence of various compounds, including Tannins, Flavonoids, Phenol and Quinones. *Azadirachta indica* plant extracts attributed to the present of various compounds, including Tannins, Flavonoids, phenols, Carbohydrate and Quinones, which may be jointly or independently contribute to produce larvicidal activity against *Aedes aegypti* and *Culex quinquefasciatus*. The acetone extract of *Azadirachta indica* leaves showed good larvicidal activity against larvae of *Aedes aegypti* (LC<sub>50</sub>=2.59 mg/ml) *Culex quinquefasciatus* (LC<sub>50</sub>=5.79mg/ml at 24 hrs). The acetone

extract of *Tridax procumbens* also showed potential larvicidal activity against 4<sup>th</sup> instar larvae of *Aedes aegypti* (LC<sub>50</sub>=6.31 mg/ml) *Culex quinquefasciatus* (LC<sub>50</sub>=3.63 mg/ml at 24 hrs). The results demonstrate an excellent activity which contributes to a rare opportunity of natural control.

## REFERENCE

1. Feuerhake, K.J. 1984. Effectiveness and selectivity of technical solvents for the extraction of neem seed components with insecticidal activity. in Schmutterer and Ascher, (see above under conference reports), 1984; 103-114.
2. Wang, L., C.L. Weller, Recent advances in extraction of nutraceuticals from plants. Trends Food Sci. Technol., 2006; 17: 300-312.
3. Ramaiah KD, Das PK, Michael E, Guyatt H The economic burden of lymphatic filariasis in India. Parasitol Today, 2000; 16(6): 251-253.
4. Ramaiah KD, Das PK, Michael E, Guyatt H The economic burden of lymphatic filariasis in India. Parasitol Today, 2000; 16(6): 251-253.
5. Ramaiah KD, Das PK, Michael E, Guyatt H The economic burden of lymphatic filariasis in India. Parasitol Today, 2000; 16(6): 251-253.
6. St. Georgiev, Vassil (2009). *National Institute of Allergy and Infectious Diseases, NIH* (1 ed.). Totowa, N.J.: Humana. p. 268. ISBN 978-1-60327-297-1. Archived from the original on, 1 May 2016.
7. Ali M, Ravinder E, Ramachandran R. A new flavonoid from the aerial parts of *Tridax procumbens*. Fitoterapia, 2001; 72: 313-5.
8. Oudegans, J.A.N.H., Agro-pesticides. United Nation Economic and Social Commission for Asia and the Pacific. Bangkok, 1991; 31.
9. Caceres A, Cano O, Samayoa B, Aguilar L. Plants used in Guatemala for the treatment of gastrointestinal disorders. I. Screening of 84 plants against enterobacteria. J Ethnopharmacol, 1994; 30: 55-73.
10. Subra, R., Ecological studies on *Culex pipiens fatigans* Wiedemann, 1828 (Diptera, Culicidae) in an urban zone of the West-African Sudan savanna. Length of life and dispersal of adults marked with fluorescent dusts. Cahiers ORSTOM. Serie Entomol. Med. Parasit, 1972; 10: 3-36.
11. Lapointe DA, Dispersal of *Culex quinquefasciatus* (Diptera: Culicidae) in a Hawaiian rain forest. Journal of Medical Entomology, 2008; 45(4): 600-609. <http://www.ingentaconnect.com/content/esa/jme/2008/00000045/00000004/art00003;jsessionid=xzqbc93gnv.alice>.
12. Hardy DE, Diptera: Nematocera - Brachycera (Except Dolichopidae). In: Insects of Hawaii Volume 10 [ed. by Zimmerman, E. C.]. Honolulu, Hawaii: University of Hawaii Press, 1960; 1-24.
13. Lounibos LP, Invasions by insect vectors of human disease. Annual Review of Entomology, 2002; 47: 233-266.
14. Subra R, Biology and control of *Culex pipiens quinquefasciatus* Say, 1823 (Diptera, Culicidae) with special reference to Africa. Insect Science and its Application, 1981; 1(4): 319-338.
15. Gould EA, Solomon T (February). "Pathogenic aviviruses". Lancet., 2008; 371(9611): 500-9. doi: 10.1016/S0140-6736(08)60238-X. PMID 18262042.
16. Reddy PJ, Krishna D, Murthy Us, Jamil K. A microcomputer FORTRAN program for rapid determination of lethal concentration of biocides in mosquito control, ComputApplBiosci, 1992; 8(3): 209-213.
17. Mohineesh Chandra, Jaya Raj, Tirath Das Dogra, Avinash Chander Rajvanshi, Anupama Raina, Determination of Median Lethal Dose of Triazophos with DMSO in wistar rats. Asian J Pharm Clin Res, 2014; 7(4): 66.
18. Abbott WA. A method for computing the effectiveness of an insecticide. J Econ Entomol, 1925; 18: 265-267.
19. Jiang, J., Oberdorster, G., Biswas, P., Characterization of size, surface charge, and agglomeration state of nanoparticle dispersions for toxicological studies. J. Nano. Res., 2009; 11: 77-89.
20. Hahn CS, French OG, Foley P, Martin EN and Taylor RP Bispecific monoclonal antibodies mediate binding of dengue virus to erythrocytes in a monkey model of passive viremia. J. Immunol., 2001; 66(2): 1057-1065.
21. Elumalai, D., Kaleena, P.K., Fathima, M., Muttapan, M., Evaluation of Biological activity of *Hyptis suaveolens* (L) Poit and *Leucas aspera* (Wild) against *Culex quinquefasciatus*. Int. J. Biosci. Res., 2013; 2: 1-6.
22. Elumalai, D., Kaleena, P.K., fathima, Mujeera, Nareshkumar, C., Phytochemical screening and larvicidal activity of *Tridax procumbens* (L) against *Anopheles stephensi* (Liston), *Aedes aegypti* (L) and *Culex quinquefasciatus* (say). Int. J. Biosci. Res., 2013b; 2: 1-14.
23. Gubler DJ. Dengue viruses. In: Mahy BW, Regenmortel MH, editors. Desk encyclopedia of human and medical virology. Boston (MA): Academic Press, 2010; 372-381.
24. Pruthi, H.S., Report of the Imperial entomologist. Sci. Rep. Agric. Res. Inst., New Delhi, 1937; 1935-1936.
25. Lfgkhlhk, ahuman, A.A., Gopalakrishnan, G., Ghose, B.S., Arumugam, S. & Himalayan, B. Effect of Feronialimonia on mosquito larvae. Fitoterapia, 2000; 71: 55.
26. Ahuman, A.A., Gopalakrishnan, G., Ghose, B.S., Arumugam, S. & Himalayan, B. Effect of Feronialimonia on mosquito larvae. Fitoterapia, 2000; 71: 55.
27. Rahuman, A.A., Gopalakrishnan, G., Ghose, B.S., Arumugam, S. & Himalayan, B. Effect of Feronialimonia on mosquito larvae. Fitoterapia, 2000; 71: 553-555.

28. WHO, 2009. Country profile of Environmental Burden of Disease: Somalia. Geneva, Switzerland: World Health Organization World Health Organization. Informal consultation on the evaluating and testing of insecticides. CTD/WHO PES/IC/96.1. Geneva, Switzerland: WHO, 1996; 69.
29. Wandscheer, C.B., Duque, J.E., da Silva, M.A.N., Fukuyama, Y., Wohlke, J.L., Adelman, J., & Fontana, J.D., Larvicidal action of ethanolic extracts from fruit endocarps of *Melia azedarach* and *Azadirachta indica* against the dengue mosquito *Aedes aegypti*. *Toxicon*, 2004; 44(8): 829–835.
30. Aliero, B.L., Larvicidal effects of aqueous extracts of *Azadirachta indica* (neem) on the Larvae of *Anopheles* mosquito. *AFRJ BIOTECHNOL.*, 2003; 2(9): 325–327.
31. Chavan, S.R., Chemistry of alkanes separated from leaves of *Azadirachta indica* and their larvicidal/insecticidal activity against mosquitoes. *Schriftenr. GTZ.*, 1984; 161: 59-65.
32. Vatandoost, H. & Vaziri, V.M., Larvicidal activity of a neem tree extract (Neemarin) against mosquito larvae in the Islamic Republic of Iran. *EMHJ*. 10, No., 2004; 4/5: 573–581.
33. Virendra, K.D., Akhilesh, C.P., Kamaraju, R., Ashish, G., Trilochen, S., & Aditya, P.D., Larvicidal activity of neem oil (*Azadirachta Indica*) formulation against mosquitoes. *Malaria J.*, 2009; 8: 124.
34. Abdelouaheb, A., Nassima, R. & Noureddine, S., Larvicidal activity of a neem tree extract (Azadirachtin) against mosquito larvae in the Republic of Algeria. *Jordan J. Biol. Sci.*, 2009; 2(1): 15–22.
35. Senthil, N.S., Savitha, G., George, D.K., Narmadha, A., Suganya, L. & Chung, P.G., Efficacy of *Melia azadirachta* extract on the malarial vector *Anopheles stephensi liston* (Diptera: Culicidae). *Bioresource Technol.*, 2006; 97: 1316-1323.
36. Naqvi, S.N.H., Biological evaluation of fresh neem extracts and some neem components with reference to abnormalities and esterase activity in insects. *Proc. 3rd Int. Neem conf.* (Nairobi 1986), 1987; 315-330.
37. Rajan M and Savarimuthu I. A novel herbal formulation against dengue vector mosquitoes *Aedes aegypti* and *Aedes albopictus*. *Parasitology Research*, 2012; 110(5): 1801-1813.
38. Sarita K, Naim W, Radhika W. Bioefficacy of *Mentha piperita* essential oil against dengue fever mosquito *Aedes aegypti* L Asian Pacific Journal of Tropical Biomedicine, 2011; 85-88.
39. Raveen, R. Dhayanidhi, P. Dhinamala1, K. Arivoli, S. and Samuel Tennyson. Larvicidal activity of *pedilanthus tithymaloides* (L.) Poit (Euphorbiaceae) leaf against the dengue vector *aedes aegypti* (L.) (diptera:culicidae). *International Journal of Environmental Biology*, 2012; 2(2): 36-40.
40. Govindarajan M and P Karuppannan. Mosquito larvicidal and ovicidal properties of *Eclipta alba* (L.) Hassk (Asteraceae) against chikungunya vector, *Aedes aegypti* (Linn.) (Diptera: Culicidae). *Asian Pac J Trop Biomed*, 2011; 49(1): 24-28.
41. Govindarajan M. Larvicidal efficacy of *Ficus benghalensis* L. plant leaf extracts against *Culex quinquefasciatus* Say, *Aedes aegypti* L. and *Anopheles stephensi* L. (Diptera: Culicidae) *European Review for Medical and Pharmacological Sciences*, 2010; 14: 107-111.
42. Radhika W, Naim W and Sarita K. Larvicidal potential of commercially available pine (*Pinus longifolia*) and cinnamon (*Cinnamomum zeylanicum*) oils against dengue fever mosquito, *Aedes aegypti* L. (Diptera; Culicidae) *Fi Acta Entomologica Sinka*, 2011; 54(7): 793 -799.
43. Rajkumar S. Jebanesan A. Repellent activity of selected plant essential oils against the malarial fever mosquito *Anopheles stephensi*. *Trop Biomed*, 2007; 2: 71-5.
44. Kamaraj, C. Bagavan, A. Elango, G. Abdur Zahir, A. Rajakumar, G. Marimuthu, S. Santhoshkumar, T and Abdul Rahuman, A., *Indian J Med Res.*, 2011; 134: 101-106.
45. Puri HS. *Neem: The divine tree; Azadirachta indica*. Amsterdam: Harwood Academic Publishers, 1999; 1-3.
46. Agarwal S, Khadese S, Talele G Bioactive immunomodulatory fraction from *Tridax procumbens*. *Science Alert*, 2010; 3: 120-127.