World Journal of Pharmaceutical and Life Sciences <u>WJPLS</u>

www.wjpls.org

SJIF Impact Factor: 4.223

ISOLATION AND CHARACTERIZATION OF SOME FLAVONES FROM ARUM CYRENAICUM (ARACEAE)

*¹Dr. Abdel Karim M., ^{3,4}Khaled A. Abdelshafeek, ²Fatima A. Saada and ²Salma M. M. Attafa

¹Sudan University of Science and Technology, Faculty of Science.
²Sirte University, Faculty of Science, Chemistry Department, Sirte, Libya.
³National Research Centre, Department of Phytochemistry, Dokki, Giza, Egypt.
⁴AlBaha University, Faculty of Science, Chemistry Department, AlBaha, KSA.

*Corresponding Author: Dr. Abdel Karim M.

Sudan University of Science and Technology, Faculty of Science.

Article Received on 13/12/2017

Article Revised on 03/01/2018

Article Accepted on 24/01/2018

ABSTRACT

Information on *Arum cyrenaicum*, which is a common herb in Aljabal Al-Akhdar (Libya), is very scarce. Hence it was planned to investigate the flavonoids of this species which is used in traditional medicine against some human disorders. Phytochemical screening of the whole plant revealed the presence of flavonoids, alkaloids, terpenes, carbohydrates and sterols.Fractionation of the butanol fraction over a polyamide column gave three flavones-compounds I,II and III.The structures of these flavonoids were elucidated via a combination of spectral techniques(UV,¹HNMR and MS).

KEYWORDS: Arum cyrenaicum, Flavonoids, Isolation.

INTRODUCTION

The genus *Arum* which comprises 28 species is native to Europe, Asia and north Africa.^[1,2] *Arum* have been used traditionally for centuries.^[3,4] This genus is divided into two sub-genera *Arum* which contains all the species except *Arum pictum* which belongs to the sub-genera Gymnomesium. The toxicity of *Arum* species is well known and most traditional uses concentrate on the anticancer potential of these species.

Kochmarov et.al.^[5] reported that Arum balansanum and Arum detruncatum are used externally for haemorroids. In Indian system of medicine Arum is used against boils,^[6] while in Jordon it is used as anticancer.^[7] The plant is used in Palestine for prostate disorders.^[8] Arum flowers are used for wounds and haemorroids. Arum *italicum* is applied topically for warts.^[9,10] This species is used in Itally for CNS disorders and rheumatic pains. It also used against eczema, canacer is and haemorroids.[11,12]

Arum cyrenaicum is a perennial herb in the sub-family Aroideae of the family Araceae. Th plant reaches 13-27 cm in height. Information on this plant which is a common herb in Aljabal Al-Akhdar (Libya), is very scarce. Hence it was planned to investigate the flavonoids of this species which is used in traditional medicine against some human disorders.

MATERIALS AND METHODS

Materials

Plant material

Arum cyrinaicum was collected from Aljabal AlAkhder region- Libya during the flowering stage. The plant was kindly identified and authenticated by Dr. Naser Elshekhi, lecturer of taxonomy at Botany Dept, Faculty of Science, Binghazi University. A voucher speciemin was deposited at the herbarium of faculty of science, Sirt University The aerial parts of the plant were air dried and powdered.

Instruments

Instruments used in flavonoids measurements include:

- i- UV viewing lamp at the long wavelength.
- ii- UV- Vis spectrophotometer 2401Shimadzu.
- iii- UVIKON 931 double beam UV- vis. spectrophotometer. All measurements in region of 200-500 nm.
- iv- Bruker NMR spectrophotometer operating at 300 MHz for ¹HNMR in DMSO- d_6 .
- v- Mass spectrometer Finnigan Mat SSQ 7000.

Methods

Phytochemical screening

Powdered whole plant of *Arum cyrenaicum* was screened for major secondary metabolites according to the method described by Harborne.^[13]

Isolation of flavonoidal constituents

Whole plant of *Arum cyrenaicum* (500g) was macerated with aqueous alcohol till exhaustion .Removal of the solvent under reduced pressure gave a crude product. The crude product was dissolved in worm water and partitioned with n-butanol. Paper chromatography of the butanol fraction using Whatmann 3mm irrigated with 20% acetic acid gave the best separation of the flavonoids. A Polyamide column chromatography of the butanol fraction gave three flavonoids: compound I (R_f 0.79), compound II (R_f 0.89) and compound III (R_f 0.58-solvent 20% acetic acid). The polyamide column is summarized in Table (1)

| Table 1: Column | chromatography | of butanol | fraction A | cvrenaicum |
|-----------------|----------------|------------|-------------|-------------|
| Table 1. Column | cm omatography | of Dutanoi | n action A. | cyrenaicam. |

| Solvent | Fractions (25ml) | R _f | Colour NH ₃ | in UV AlCl ₃ | Isolated compound |
|--------------------------|---------------------|----------------|---------------------------|----------------------------|----------------------|
| Methanol: Water 30:70 | 10-13 | 0.79 | YG | GY | Compd. I |
| Methanol: Water 90:10 | 25-28 | 0.89 | Br. | Br. | Compd. II |
| Methanol: Water 60:40 | 20-24 | 0.58 | GY | Y | Compd. III |

Paper chromatography: 20% acetic acid; whatmann No. 3mm. Y.=Yellow

Br.=Brown G.Y.=Greenish-Yellow

RESULTS AND DISCUSSION

The whole plant of *Arum cyrenaicum* was screened for major secondary metabolites and the results are summarized in Table (2).

Table 2: Results phytochemical screeningA.cyrenaicum.

| Constituents | Results |
|-----------------------------------|---------|
| Flavonoids | + |
| Carbohydrates and / or glycosides | + |
| Sterols and / or triterpenoids | + |
| Tannins | - |
| Alkaloids | + |
| Saponins | - |

Identification of compound I

The UV absorption spectra of compound I (Fig. 1 and Table 3) showed λ_{max} 274, 335nm. Such absorption is characteristic of flavones.^[14,15] It also revealed a bathochromic shift(15nm) in band-I (Fig.2) with sodium methoxide, indicating^[14] the presence of a free OH function at C- 4[°]. The aluminium chloride spectrum(Fig.3) showed a bathochromic shift(49nm) in band -I relative to methanol spectrum suggesting^[14] the presence of a free OH group at C- 5. The AlCl₃/ HCl spectrum showed no hypsochomic shift in band- I relative to AlCl₃ spectrum which lends evidence for the absence of a catechol system (Fig.4). The absence of *ortho*-dihydroxy systems is further confirmed through the boric acid spectrum (Fig.5). The sodium acetate spectrum (Fig.6) did not show a bathochromic shift in band- II diagnostic of a 7-OH function.

Table 3: UV data of compound I.

| Addition to MeOH | $\lambda_{\max}(nm)$ |
|--|----------------------|
| None | 274, 335 |
| NaOMe | 281, 305,350 |
| AlCl ₃ | 281, 305, 350, 384 |
| HCl | 282, 303, 343 |
| NaOAc | 282, 396 |
| NaOAc / H ₃ BO ₃ | 275, 339, 347 |

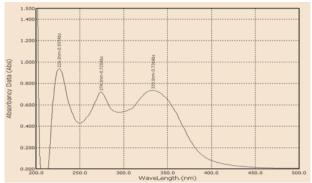


Fig. 1: UV spectrum of compound I.

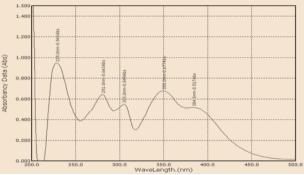


Fig. 2: Sodium methoxide spectrum of compound I.

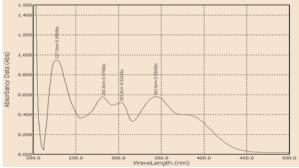


Fig. 3: Aluminium chloride spectrum of compound I.

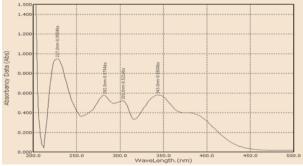


Fig. 4: Aluminium chloride/HCl spectrum of compound I.

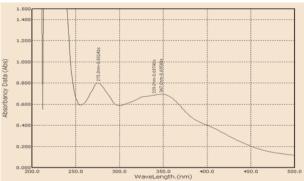
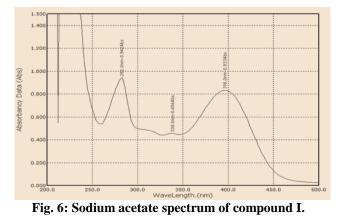
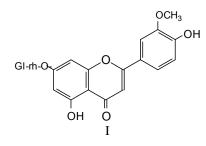


Fig. 5: Boric acid spectrum of compound I.



The ¹H- NMR spectrum (DMSO-d₆) (Fig.7) showed signals at δ (ppm): 1.22(rhaminose Me) ; 300-3.70(rhamnoglucosyl moiety), 3.95 (MeO) ; 4.85(sugar anomeric proton) ; 6.77(C₆-H) ; 6.93(C₈ –H) ; 7.94,8.08(Ar. protons). The mass spectrum (Fig. 8) gave m/z607 (for M⁺-H).

The above cumulative data is consistent with the published data for chrysoeriol-7-rhamnoglucoside:



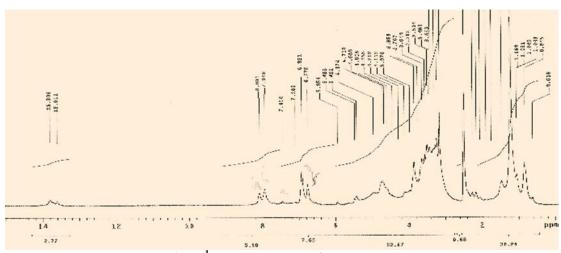
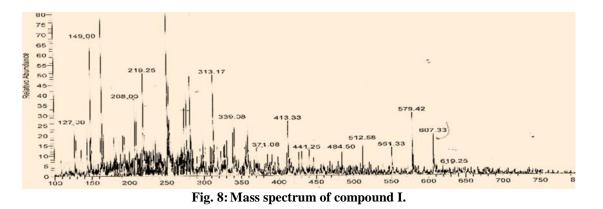


Fig. 7: ¹HNMR spectrum of compound I.



Identification of compound II

The UV spectrum of compound II (Fig. 9 and Table 4) showed a pattern characteristic of flavones at λ_{max} 273, 332nm. Addition of the UV shift reagent sodium methoxide revealed a bathochromic shift(Fig. 10) in band-I (71nm) with increased intensity suggesting the presence of a free OH group at C- 4^{\circ}. The AlCl₃ spectrum(Fig.11) showed a bathochromic shift in band - I (71nm) relative to methanol spectrum.This shift suggests a B-ring catechol system as evidenced by the degeneration of AlCl₃ / HCl spectrum (Fig. 12). The sodium acetate spectrum(Fig.13) showed a bathochromic shift (8nm) in band - II diagnostic of free 7- OH function.

Table 4: UV absorption spectra of compound II.

| Addition to MeOH | $\lambda_{max}(nm)$ |
|-------------------|---------------------|
| None | 273, 332 |
| NaOMe | 283, 336, 403 |
| AlCl ₃ | 283, 336, 403 |
| HCl | 281, 339, 341 |
| NaOAc | 281, 339 |

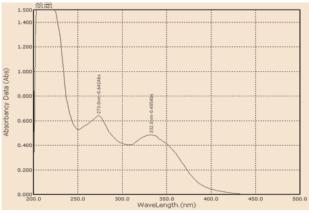


Fig. 9: UV spectrum of compound II.

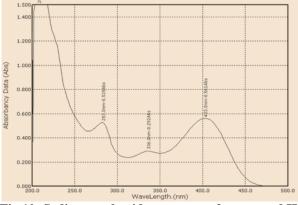


Fig.10: Sodium methoxide spectrum of compound II.

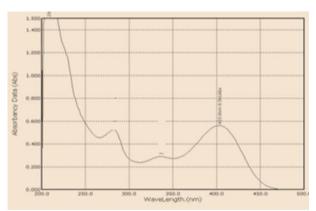


Fig. 11: Aluminium chloride spectrum of compound II.

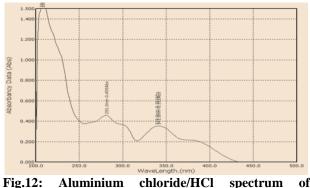


Fig.12: Aluminium chloride/HCl spectrum of compound II.

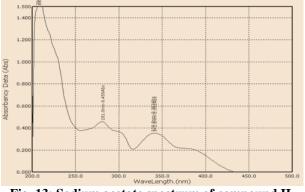
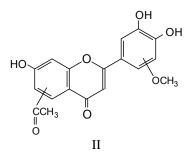


Fig. 13: Sodium acetate spectrum of compound II.

The ¹H- NMR spectrum (DMSO-d₆) (Fig.14) showed signals at δ (ppm) : 1.85(assigned for acetyl function) ; 3.71(MeO) ; 6.90(C₆ -H) ; 6.92(C₈ -H) ; 7.93(Ar.

protons). The mass spectrum (Fig.15) gave $m/z341\ for\ M^+$ - H.

On the basis of the above cumulative data, the following partial structure was proposed for compound II:



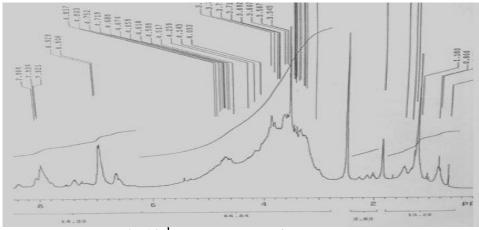


Fig. 14: ¹HNMR spectrum of compound II.

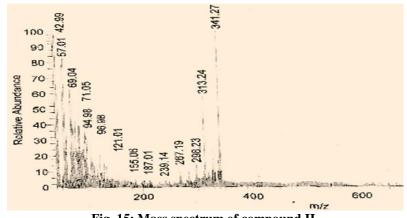


Fig. 15: Mass spectrum of compound II.

Identification of compound III

In the UV compound IV absorbs at λ_{max} 275, 341 nm (Fig.16 and Table 6). Such absorption is characteristic of flavones. Addition of sodium methoxide caused a 62nm bathochromic shift (Fig.17) in band-I with increased intensity, indicating a 4⁻OH function. The boric acid spectrum (Fig.18) suggested a B-ring catechol system .The sodium acetate spectrum (Fig.19) which is diagnostic of 7-hydroxylation showed a bathochromic

shift (8nm) in band - II indicating the presence of a C_7 – OH function.

| Table 5: The UV | absorption spectra | of compound. |
|-----------------|--------------------|--------------|
|-----------------|--------------------|--------------|

| Addition to MeOH | $\lambda_{\max}(nm)$ |
|--|----------------------|
| None | 275, 341 |
| NaOMe | 283, 336, 340, 403 |
| NaOAc | 283, 395 |
| NaOAc / H ₃ BO ₃ | 281, 353 |

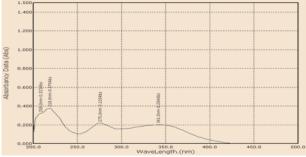


Fig. 16: UV spectrum of comound III.

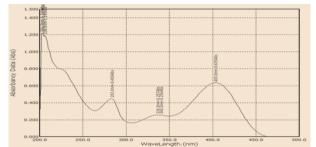


Fig. 17: Sodium methoxide spectrum of compound III.

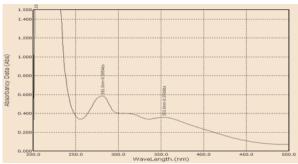


Fig. 18: Boric acid spectrum of compound III.

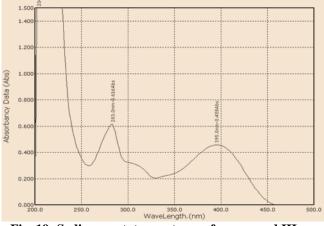
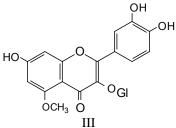


Fig. 19: Sodium acetate spectrum of compound III.

The ¹H- NMR spectrum of compound III (DMSO-d₆) (Fig.20) showed signals at δ (ppm): 3.87 (MeO); 4.40-5.00(glucose moiety) ; 6.50(C₆-H) ; 6.90(C₈ –H) ; 7.85, 8.04(Ar. protons). The mass spectrum(Fig.21) gave m/z479 for M⁺ - 2H (glycoside) and m/z317 for M⁺¹ - 2H(aglycone).

On the basis of the above spectral data, the following tentative structure was proposed for compound III:



However, a future 2D NMR experiments (${}^{1}H{}^{-1}$ H COSY NMR HSQC, HMBC) may lend evidence for the citation of the methoxyl function at C₅ and the location of the glycosidic linkage at C₃.

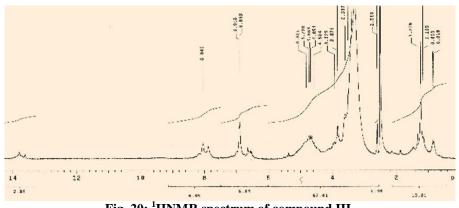
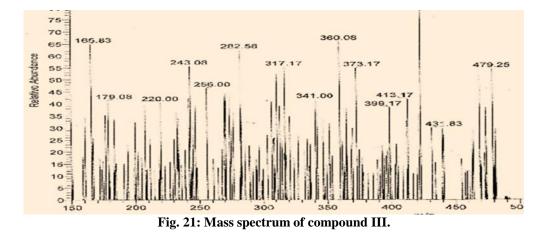


Fig. 20: ¹HNMR spectrum of compound III.



REFERENCES

- 1. Boyce, P. "The *Genus Arum*", The Royal Botanic Gardens, Kew, London, 1993; 197.
- Mayo, S., Bogner J. and P. C. Boyce, *The Genera of Araceae*, Royal Botanical Gardens, Kew., 1997; 370.
- Koach, J. Araceae, *In* D. N. Feinburn (ed.), Flora of Palaestina-Israel N. Feinburn (ed.), Flora Palaestina. Israel Academy of Science and Humanities, Jerusalem, 1986; 330-340.
- Fridlender, A. 1999. Identification des Arum de la flore frans, aise. *j. Bot. Soc. bot. France*, 1999; 11: 29-43.
- Kochmarov, V., Kozuharova, E., Naychov, Z., Momekov, G., Mincheva, I., *Int. J. Pharm. Chem. Biol. Sci.*, 2015; 5: 394.
- Sajem, A. L., Gosai, K., J. Ethnobiol. Ethnomed., 2006; 2: 33.
- Afifi-Yazar, F. U., Kasabri, V., Abu-Dahab, R., *Planta Med.*, 2011; 77: 1203.
- 8. Polat, R., Cakilcioglu, U., Satil, F., *J. Ethnopharmacol.*, 2013; 148: 951.
- 9. Pieroni, A., Quave, C. L., Santoro, R. F., J. *Ethnopharmacol*, 2004; 95: 373.
- Montesano, V., Negro, D., Sarli, G., De Lisi, A., Gaetano Laghetti, G., Hammer, K., J. Ethnobiol. Ethnomed., 2012; 8: 15.
- 11. Leporatti, M. L., Ghedira, K., J. Ethnobiol. Ethnomed., 2009; 5: 31.
- 12. Genc, G. E., Ozhatay, N., Turk. J. Pharm. Sci., 2006; 3: 73.
- 13. Harborne, J. B., "Phytochemical Methods", Chapman and Hall, London, 1986.
- 14. Jard, L., In: "The Chemistry of Flavonoid Compounds", Pergamon Press, New York, 1962.
- 15. Geissman, T.A. "Chemistry of Flavonoid Compounds", 1962, Pergamon Press, Oxford.