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TOXICITY EVALUATION OF NANOCONSTRUCT OF CHITOSAN AND TRIPOLYPHOSPHATE (TPP)

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1. INTRODUCTIN

The prefix "nano" has found in last decade an ever-increasing application to different fields of the knowledge.^[1] Nanoscience, nanotechnology, nanomaterials or nanochemistry are only a few of the new nano-containing terms that occur frequently in scientific reports, in popular books as well as in newspapers and that have become familiar to a wide public, even of non-experts.^[2] The prefix comes from the ancient Greek *vavoç* through the Latin *nanus* meaning literally *dwarf* and, by extension, *very small*. Within the convention of International System of Units (SI) it is used to indicate a reduction factor of 109 times.^[3] So, the nanosized world is typically measured in nanometers (1nm corresponding to 10^{-9} m) and it encompasses systems whose size is above molecular dimensions and below macroscopic ones (generally > 1 nm and < 100 nm).^[4]

Nanoparticles (NPs) represent a specific type of matter (from about 1 to 100 nm in size).^[5] They are intermediate in size between bulk materials and atomic/molecular structures, and possess unique physical and chemical properties.^[6] These distinctive properties, related to a high surface area to volume ratio and/or quantum effects. have spawned notable interest from engineers, biologists, chemists, and physicists. In the past decade, there has been an exponential growth in the synthesis of NPs, commonly termed as engineered nanoparticles (ENPs), due to their extensive use in emerging technologies and in consumer products such as electronic devices and other products used for personal care, biomedicine, agriculture, water/soil treatment, and renewable energy.^[7-9] An array of ENPs have been manufactured which include mainly metals, non-metals, metal oxides, , and polymers nanocomposites.^[10,11] as well as various

Nanoparticle drug delivery systems are nanometeric carriers used to deliver drugs or biomolecules.^[12] Generally, nanometeric carriers also comprise sub-micro particles with size below 100 nm and with various morphologies, including nanospheres, nanocapsules, nanomicelles, nanoliposomes, and nanodrugs, etc. Nanoparticle drug delivery systems have outstanding advantages:^[13-16]

 They can pass through the smallest capillary vessels because of their ultra-tiny volume and avoid rapid clearance by phagocytes so that their duration in blood stream is greatly prolonged;

- (2) They can penetrate cells and tissue gap to arrive at target organs such as liver, spleen, lung, spinal cord and lymph;
- (3) They could show controlled release properties due to the biodegradability, pH, ion and/or temperature sensibility of materials;
- (4) They can improve the utility of drugs and reduce toxic side effects; etc.

Chitosan is a non-toxic (In case of low amount non-toxic but in high amount in show toxic) biodegradable polycationic polymer with low immunogenicity.^[17-20] Chitosan is a linear polysaccharide composed of randomly distributed-(1-4)-linked d-glucosamine and Nacetyl-d-glucosamine.^[21-23] Due to the advantageous biological properties of chitosan, such as relative nontoxicity, biocompatibility, biodegradability, cationic properties, bio-adhesive characteristics and permeabilityenhancing properties, chitosan-based particles have been extensively studied for delivery of anti-cancer agents, therapeutic proteins, genes, antigens.^[24,25]

Chitosan (CS), the N-deacetylation form of chitin mostly found in the exoskeleton of crustacean, insects, and fungi, is a natural polysaccharide.^[26-28] CS is not only non-toxic and biodegradable with low immunogenicity, but also possesses a high density of positive charge in an acid solution attributed to the glucosamine group on its backbone.^[29,30] Because of these beneficial characteristics, increasing attention has been drawn to the applications of CS-based micro and nanoparticles in the pharmaceutical and nutraceutical field.^[31,32] Ionic gelation is the most studied and widely used method for fabricating CS nanoparticles, in which cationic CS and multivalent polyanions interact to form CS nanoparticles under simple and mild conditions. Among various polyanions, Tripolyphosphate (TPP) is the most investigated due to its quick gelling capability and non-toxic property.^[33-36]

As drug delivery system, nanoparticles can entrap drugs or biomolecules into their interior structures and/or absorb drugs or biomolecules onto their exterior surfaces. Presently, nanoparticles have been widely used to deliver drugs, polypeptides, proteins, vaccines, nucleic acids, genes and so on.^[37,38] Over the years, nanoparticle drug delivery systems have shown huge potential in biological, medical and pharmaceutical applications Chitosan has attracted considerable interest because of their unique combination of properties, such as biocompatibility, biodegradability, metal complexation and antibacterial activity.^[39] Therefore, chitosan has a variety of current and potential applications in various fields, for example, biotechnology, pharmaceutics, wastewater treatment.^[40]

The antibacterial activity of chitosan has been widely explored. A number of chitosan derivates with different modifications have been prepared to improve its antibacterial activity.^[41] The metal-chelating property of chitosan has been mainly used in wastewater treatment. Recently, different metal chitosan complexes have been prepared to improve its antimicrobial activity.^[42-44]

Toxicology traditionally addresses adverse poisoning effects of chemicals to humans, animals and the environment.^[45,46] Historically, toxicology is often associated with Paracelsus and the concept of dose and dose response. He is attributed with having coined the phrase "the dose makes the poison", implying a linear relationship. However toxicological dose responses can be complex and decidedly non-linear especially in the low and high dose range.^[47,54]

2. MATERIALS AND METHODS

2.1 Materials

Low molecular weight (LMW) water-soluble chitosan, Sodium Tripolyphosphate (TPP), glacial acetic acid, sodium hydroxide and all other chemicals were analytical grade, ultrapure water was used throughout this study.^[39]

2.2 Animal used

Male Wistar rat of weight 180-250 gm were taken. Experimental protocols used in the experiment were approved by the Institutional Animal Ethic Committee (IAEC reg. no. CIP/IAEC/2015-16/063) of Columbia Institute of Pharmacy Tekari, Raipur (C.G.). The animals were housed in polycarbonate cages in a room with a 12 h day-night cycle, temperature of 22 ± 2 °C, and humidity of 45-64%. During the whole experimental period,

animals were fed with balanced commercial pellet diet (Ashirwad Industries, Mohali, India) and water *ad libitum* and normal control, Test Control, Reference group each containing six animals for In-vivo Nanoconstruct Toxicity activity.^[56,57]

2.3 Preparation of Chitosan/TPP Nanoconstruct 2.3.1 Ionic gelation method

Chitosan was dissolved in an aqueous solution of acetic acid to form a 0.5 mg/mL chitosan solution. The concentration of acetic acid was 0.4 times (0.2 mg/mL) that of chitosan. The chitosan solution was stirred overnight at room temperature using a magnet stirrer. The pH of the resulting solution was around 3.6 and this was adjusted to 4.7-4.8 using 20 wt% aqueous sodium hydroxide solution. The chitosan solution was then passed through a syringe filter (pore size 0.45 µm, Millipore, USA) to remove residues of insoluble particles. TPP was dissolved in ultrapure water at a concentration of 0.5 mg/mL and also passed through a syringe filter (pore size 0.22 µm, Millipore, USA). To prepare chitosan nanoparticles, a magnetic stirrer was placed in a chest freezer, in which the ambient temperature was controlled at 2-4°C, temperature fluctuations and flow of cold air should be avoided as much as possible. Ten millilitres of chitosan solution in a 25 mL round-bottom flask was preheated in a water bath at 60°C For 10 min, the flask was then placed on the magnetic stirrer stirring at 700 rpm, 3.0 ml of 2-4°C TPP solution was quickly added to the chitosan solution with a plastic Pasteur pipette. The reaction was carried out for 10 min and the resulting Suspension was subjected to further analysis.[39]

2.4 Characterization and morphology of Chitosan/TPP Nanoconstruct

2.4.1 Transmission electron microscope (TEM)

The size of the nanoparticles was examined using a high resolution Transmission Electron Microscope (TEM) machine (JOEL 2100F).

2.4.2 Particle size and zeta potential

The sizes and zeta potential of the CNP were measured with a Malvern Zetasizer Nano ZS (Malvern Instruments Ltd., Malvern, U.K.). The particle size distribution of the nanoparticles is reported as a polydispersity index (PDI). All measurements were performed in triplicates. 3 ml of sample was taken in a cuvette and was analyzed at 25 _C with at an angle of 90.

2.5 In-Vivo experimental method

2.5.1 Acute oral toxicity studies

The animals used in the toxicity studies were approved by the Institute Animal Ethics Committee (IAEC), India. Wistar male rats weighing 180-250 gm were used. The acute toxicity study was carried out on male Wistar rats by administering Nanoconstruct orally at one of the levels (0.5, 1.0, 1.5, 2.0 and 2.5 ml/kg) once only. Mortality was not found up to 2.5 ml/kg dose. So, as per OECD guideline 423 which was half of the maximum dose was considered for therapeutic exploration.^[56]

2.5.2 Repeated dose 28 days oral toxicity studies:

The animals used in the toxicity studies were approved by the Institute Animal Ethics Committee (IAEC), India. Wistar male rats weighing 180-250 gm were used. The acute toxicity study was carried out on male Wistar rats by administering Nanoconstruct orally at repeated 28 days of the levels (0.5, 1.0, 1.5, 2.0 and 2.5 ml/kg) single dose daily only. So, as per OECD guideline 407.^[57]

2.5.2.1 Haematology

Blood was collected on the initial day, after 7, 14, 21 and 28 days by Retro orbital plexus from the overnight fasted animals. Investigation of whole blood for following was done: Red blood cells (RBCs), White blood cells (WBCs), Haemoglobin (Hb), Platelet count, Haematocrit (HTC), Mean Corpuscular Volume (MCV), Mean Corpuscular Haemoglobin (MCH), Mean Corpuscular Haemoglobin Concentration (MCHC), RDW-SD, RDW-CV, PDW, MPV, P-LCR, PCT, Neutrophil (N), Lymphocytes (L), Monocytes (M), Eosinophil's (E), Basophils (B).^[57]

2.5.2.2 Clinical Biochemistry

Blood was collected on the initial day, after 7, 14, 21 and 28 days from retro orbital plexus of overnight fasting rats and different parameters in blood like determination in serum or plasma should include: Sodium, potassium, glucose, total cholesterol, urea, blood urea nitrogen, creatinine, total protein and albumin, Alkaline Phosphatase (ALP), Alanine aminotransferase, Aspartate aminotransferase, Gamma glutamyl transpeptidase, Sorbitol dehydrogenase, Bile acids.^[57]

2.5.2.3 Histopathology

All animals were sacrificed and organs Liver, Kidney were fixed immediately in 10% formalin for routine Histopathological examination. The tissues were embedded in paraffin and then sectioned, stained with Haematoxylin and Eosin and were examined under light microscope.^[57]

2.5.2.4 Statistical analysis

All studies were performed in triplicate and the values were expressed in mean \pm SD. The data was analysed by one way analysis of variance (ANOVA) Graph Pad Prism Instat Software (version 6.00, Graph Pad Software), using one way ANOVA followed by student test. ANOVA was done to show that the work done is statistically significant a value of P < 0.05 was considered to be statistically insignificant and significant.

3. RESULTS AND DISCUSSION

3.1 Morphological characterization

3.1.1 Transmission Electron Microscopy (TEM)

TEM is an important parameter for the surface morphology characterization of Nanoconstruct. The surface morphology of the prepared Nanoconstruct was characterized by Transmission Electron Microscope (TEM) studies. The results of TEM pictures reveal that Nanoconstruct were almost spherical in shape. TEM images of the formulation are shown in fig. 3.1.

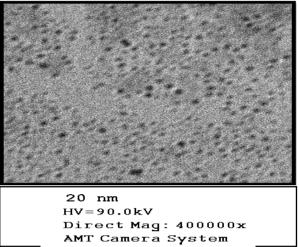
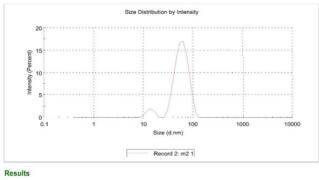


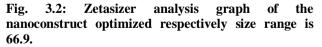
Fig. 3.1: Transmission Electron Microscope (TEM) of the Nanoconstruct Optimized Respectively Size, Range – 20 nm taken at 400000 X Magnification.

3.2 Zetasizer

The Zeta potential of formulations were analysed by using Malvern Zetasizer. Zeta potential is the electric potential of a particle in a formulation. This parameter is useful for the assessment of physical stability of the colloidal dispersion. Zeta potential and particle size of Nanoconstruct were found in Zetasizer and shown in fig.3.2.







3.3 Acute oral toxicity studies

There was no mortality or morbidity observed in animals through the 3-days period following single oral administration at all selected dose levels of the Nanoconstructs.

3.4 Repeated dose 28 days oral toxicity study

3.4.1 Body weight (gm)

 Table 3.1: Effect of Nanoconstruct on Body weight in Albino Wistar Rats.

Control	0.5 ml/kg	1.0 ml/kg	1.5 ml/kg	2.0 ml/kg	2.5 ml/kg
170.21±1.03	171.97±0.71	173.53±1.05	173.50±0.84	172.62±0.87	171.72±0.43
180.66 ± 0.52	181.86±0.48	183.83 ± 0.80	183.09±0.60	182.61±0.75	181.23±0.45
190.83±0.58	192.36±0.52	194.87±0.50	192.47±0.63	193.29±0.82	191.01±0.46
200.83±0.45	201.27±0.33	205.37±0.83	202.16±0.72	201.76±0.55	200.12±1.41
212.25±0.73	211.92±0.70	214.10±0.79	211.90±0.49	209.23±0.34	206.58±1.35
	170.21±1.03 180.66±0.52 190.83±0.58 200.83±0.45	170.21±1.03171.97±0.71180.66±0.52181.86±0.48190.83±0.58192.36±0.52200.83±0.45201.27±0.33	170.21±1.03171.97±0.71173.53±1.05180.66±0.52181.86±0.48183.83±0.80190.83±0.58192.36±0.52194.87±0.50200.83±0.45201.27±0.33205.37±0.83	170.21±1.03171.97±0.71173.53±1.05173.50±0.84180.66±0.52181.86±0.48183.83±0.80183.09±0.60190.83±0.58192.36±0.52194.87±0.50192.47±0.63200.83±0.45201.27±0.33205.37±0.83202.16±0.72	170.21±1.03171.97±0.71173.53±1.05173.50±0.84172.62±0.87180.66±0.52181.86±0.48183.83±0.80183.09±0.60182.61±0.75190.83±0.58192.36±0.52194.87±0.50192.47±0.63193.29±0.82200.83±0.45201.27±0.33205.37±0.83202.16±0.72201.76±0.55

Value are Mean±SEM. (n=6).

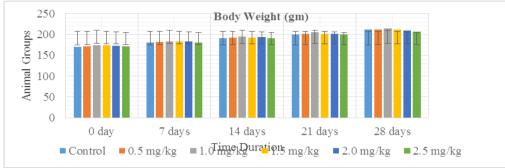


Fig. 3.3: Animals mean body weight during pre-dosing, Number of animals per group=6. All value are reported as Mean±SEM.

3.4.2 Food consumption (gm)

Table 3.2: Effect of Nanoconstruct on Food consumption in Albino Wistar Rats.

Duration	Control	0.5 ml/kg	1.0 ml/kg	1.5 ml/kg	2.0 ml/kg	2.5 ml/kg
0 Day	83.16±0.28	83.33±0.30	82.17±0.28	83.50±0.39	82.50±0.45	83.17±0.36
7 Days	83.50±0.20	84.00±0.40	85.50±0.39	84.67±0.56	84.67±0.38	85.50±0.39
14 Days	87.66±0.38	87.16±0.28	88.00±0.33	87.83±0.43	87.67±0.45	87.83±0.43
21 Days	95.16±0.28	95.67±0.30	94.00±0.33	93.50±0.39	94.33±0.45	94.17±0.28
28 Days	106.16±0.43	106.00±0.33	105.83±0.43	104.33±0.65	106.00±0.33	105.83±0.49

Value are Mean±SEM. (n=6).

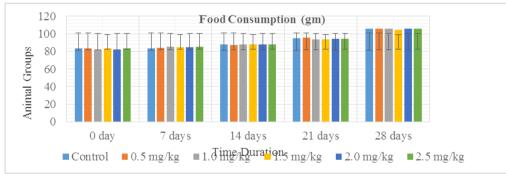


Fig. 3.4: Animals mean food consumption during dosing, Number of animals per group=6. All value are reported as Mean±SEM.

3.4.3 Water consumption (ml)

Table 3.3: Effect of Nanoconstruct on Water consumption in Albino Wistar Rats.

Duration	Control	0.5 ml/kg	1.0 ml/kg	1.5 ml/kg	2.0 ml/kg	2.5 ml/kg
0 Day	80.17±0.28	81.17±0.28	80.67±0.30	81.00±0.33	81.83±0.43	81.83±0.43
7 Days	82.00±0.33	82.00±0.52	82.33±0.38	83.50±0.39	84.67±0.30	82.500.54
14 Days	89.83±0.64	90.17±0.49	90.17±0.54	88.50±0.31	89.17±0.64	89.67±0.50
21 Days	94.00±0.78	94.83±0.43	93.67±0.45	92.50±0.56	93.67±0.93	93.33±0.76
28 Days	106.33±0.30	105.67±0.73	105.67±0.73	105.67±0.38	105.67±0.83	106.00±0.47

Value are Mean±SEM. (n=6).

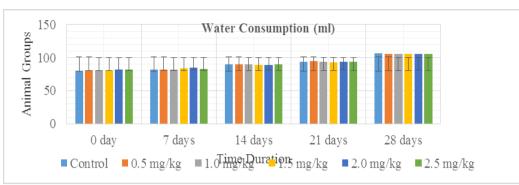


Fig. 3.5: Animals mean water consumption during dosing, Number of animals per group=6. All value are reported as Mean±SEM.

3.4.4 Organ weight (gm)

Table 3.4: Effect of Nanoconstruct on Organ weight in Albino Wistar Rats.

Animal Organ	Control	0.5 ml/kg	1.0 ml/kg	1.5 ml/kg	2.0 ml/kg	2.5 ml/kg
Liver	5.195±0.013	5.175±0.013	5.171±0.013	5.181±0.007	5.230±0.005	5.242±0.002
Kidney	0.573±0.003	0.482 ± 0.004	0.516 ± 0.001	0.542±0.004	0.544 ± 0.003	0.551±0.002
Heart	0.853 ± 0.001	0.835 ± 0.002	0.815 ± 0.002	0.850 ± 0.008	0.849 ± 0.002	0.858 ± 0.002
Pancreas	0.244±0.004	0.241±0.004	0.221±0.002	0.234±0.001	0.238±0.002	0.245±0.004

Value are Mean±SEM (n=6).

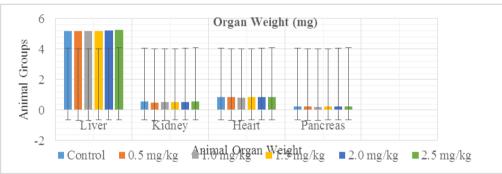


Fig. 3.6: Animals mean Organ weight after during dosing, Number of animals per group=6. All value are reported as Mean±SEM.

3.4.5 Haematological parameters

3.4.5.1 Comparison of First day Haematological parameters (Complete Blood Count, Liver Function Test, Lipid Profile) test reports with Mean±SEM.

Table 3.5: Effect of Nanoconstruct on Haematological parameters (Complete Blood Count) in Albino Wistar Rats.

Parameters	Control	0.5 ml/kg	1.0 ml/kg	1.5 ml/kg	2.0 ml/kg	2.5 ml/kg				
Complete Blood Count										
Haemoglobin	15.32±0.142	15.53±0.030	13.53±0.030	14.11±0.028	14.55±0.031	15.63±0.038				
WBC Count $(10^3/uL)$	10.13±0.123	10.16±0.030	9.13±0.019	10.35±0.031	10.13±0.019	10.46±0.019				
Neutrophils	41.16±0.597	42.33±0.384	41.5±0.311	44.16±0.495	43.33±0.192	43.33±0.962				
Lymphocytes	51.66±0.384	50.66±0.384	48.16±0.495	42.16±0.495	41.33±0.192	41.66±0.962				
Eosinophil	5.83±0.152	5.66±0.192	5.66±0.192	4.5±0.311	5.66±0.192	5.16±0.152				
Monocytes	1±0	1.33±0.192	2±0.235	1.33±0.192	4.16±0.152	6.16±0.495				
Basophils	0.1±0	0.2±0	0.13±0.019	0.13±0.019	0.13±0.019	0.16±0.019				
RBC Count (10 ⁶ /uL)	6.10±0.012	6.11±0.002	6.14±0.034	6.14±0.009	6.16±0.003	6.25±0.101				
Platelet Count	2.9±0	3.00±0.003	2.91±0.008	2.81±0.001	2.91±0.001	2.62±0.004				
Mean Platelet Value	10.5±0	9.21±0.003	9.71±0.036	9.78±0.059	10.86±0.031	8.92±0.007				
Packed Cell Volume	42.96±0.161	42.25±0.065	40.15±0.031	42.13±0.019	40.18±0.059	42.13±0.019				
Mean Corp. Vol.	70.52±0.072	70.15±0.003	68.14±0.003	69.17±0.006	79.18±0.006	80.14±0.002				
Mean Corpuscular	24.56±0.013	24.33±0.007	26.34±0.004	26.73±0.005	25.72±0.007	23.23±0.009				
Mean Corp. Hb Con.	34.65±0.087	32.66±0.069	31.73±0.030	33.78±0.036	32.2±0.074	33.13±0.019				

Red Cells Destribution Width	14.5±0.108	12.6±0.047	11.63±0.038	13.65±0.031	19.18±0.049	19.43±0.019				
Biochemistry (Liver Function Test)										
Bilirubin-Total	0.88±0.003	0.67±0.005	0.65±0.002	0.61±0.001	0.75±0.001	0.77±0.005				
Bilirubin-Direct	0.20±0.003	1.1±0.003	0.21±0.007	0.24±0.003	0.14±0.001	0.16±0.003				
Bilirubin-Indirect	0.67±0.010	2.89±0.003	0.47 ± 0.004	0.46±0.003	0.41±0.001	0.45±0.001				
Total Protein	6.6±0.047	6.85±0.020	6.56±0.045	5.45±0.031	6.65±0.031	6.73±0.096				
Albumin	3.31±0.047	3.21±0.028	3.13±0.030	3.61±0.049	3.83±0.038	3.43±0.038				
Globulin	2.81±0.039	2.63±0.038	2.21±0.036	2.48±0.036	3.13±0.019	3.25±0.031				
A/G Ratio	1.17 ± 0.005	1.12±0.003	1.32±0.004	1.11±0.003	1.22±0.003	1.27±0.005				
SGOT	80.28±0.042	82.41±0.059	83.53±0.019	78.5±0.023	79.36±0.038	84.33±0.019				
SGPT	16.81±0.028	17.68±0.059	18.83±0.019	19.13±0.019	20.13±0.019	22.43±0.019				
Alkaline Phosphatase	43.26±0.054	44.16±0.030	40.15±0.031	45.13±0.019	48.86±0.019	58.8±0.057				
		Biochemistr	y (Lipid Profil	e)						
Blood Glucose (Random)	82.86±0.215	87.18±0.028	81.2±0.023	89.55±0.031	89.56±0.019	95.36±0.038				
Serum CREATININE	0.71±0.023	0.78 ± 0.028	0.76±0.019	0.75 ± 0.031	0.76 ± 0.030	0.63±0.038				
S. Cholesterol	124.5±0.865	136.1±0.028	126.2±0.023	116.6±0.038	136.6±0.040	126.4±0.019				
S. Triglycerides	70.0541.16±	81.5±0.031	79.13±0.090	84.05±0.031	89.16±0.019	89.13±0.019				
HDL Cholesterol	19.23±0.005	18.24±0.003	19.23±0.005	17.13±0.003	19.13±0.009	19.21±0.003				
LDL Cholesterol	62.74±0.006	61.73±0.006	58.74±0.004	68.75±0.002	66.72±0.002	76.74±0.005				
VLDL Cholesterol	14.28±0.052	15.23±0.038	16.16±0.019	17.2±0.023	17.36±0.038	19.33±0.019				
LDLC/HDLC Ratio	2.44±0.005	2.43±0.010	2.44±0.003	2.46±0.004	2.41±0.001	2.12±0.007				
TC/HDLC Ratio	3.67±0.004	3.56±0.003	3.34±0.005	3.31±0.002	3.43±0.005	3.85±0.012				
S. Cholesterol/HDLC Ratio	5±0	5±0	5±0	5±0	5±0	5±0				

Value are Mean±SEM. (n=6).

3.4.5.2 Comparison of After 7 day Haematological parameters (Complete Blood Count, Liver Function Test, Lipid Profile) test reports with Mean±SEM.

Table 3.6: Effect of Nanoconstruct on Haematological parameters (Complete Blood Count) in All	oino Wistar
Rats.	

Parameters	Control	0.5 ml/kg	1.0 ml/kg	1.5 ml/kg	2.0 ml/kg	2.5 ml/kg				
Complete Blood Count										
Haemoglobin	15.65±0.085	15.08±0.036	15.5±0.031	16.13±0.019	15.06±0.038	16.51±0.028				
WBC Count (10 ³ /uL)	10.11±0.064	9.41±0.043	9.48±0.059	9.73±0.045	9.16±0.019	9.58±0.026				
Neutrophils	43.5±0.515	45.16±0.435	41.66±0.192	45.5±0.311	42±0.471	41.33±0.384				
Lymphocytes	50.5±0.392	54.33±0.384	51.5±0.311	51±0.745	51.33±0.192	54.33±0.192				
Eosinophil	6±0	5.33±0.192	5±0.233	5.16±0.152	5.16±0.152	5.66±0.192				
Monocytes	1±0	2.33±0.192	2±0.235	2±0.235	1.83±0.152	1.66±0.192				
Basophils	0.2±0	0.11±0.015	0.13±0.019	0.13±0.019	0.23±0.050	0.16±0.019				
RBC Count (10 ⁶ /uL)	6.12 ± 0.002	6.37±0.004	6.36±0.006	6.32 ± 0.004	6.37±0.005	6.42±0.010				
Platelet Count	2.91±0.005	3.84 ± 0.002	3.82±0.006	3.74±0.001	3.84±0.001	3.84±0.005				
Mean Platelet Value	10.5±0	8.24±0.003	9.24±0.001	9.25±0.001	9.21±0.001	9.23±0.009				
Packed Cell Volume	43.13±0.019	39.45±0.031	39.55±0.031	38.55±0.031	39.48±0.015	39.38±0.054				
Mean Corp. Vol.	70.61±0.003	63.10±0.003	68.14±0.003	68.10±0.004	68.15±0.001	68.12±0.010				
Mean Corpuscular	24.51±0.004	24.74±0.003	25.75±0.003	25.74±0.001	25.79±0.003	25.62±0.010				
Mean Corp. Hb Con.	38.75±0.031	42.42±0.005	42.42±0.002	42.46±0.001	42.41±0.009	42.41±0.009				
Red Cells Destribution Width	14.18±0.025	14.43±0.019	14.43±0.019	15.4±0.023	14.23±0.078	14.43±0.019				
	B	Biochemistry (I	Liver Function							
Bilirubin-Total	0.81±0.003	0.74 ± 0.003	0.73 ± 0.001	0.74 ± 0.001	0.62 ± 0.001	0.79±0.001				
Bilirubin-Direct	0.21 ± 0.004	0.23 ± 0.003	0.21±0.007	0.25 ± 0.002	0.24 ± 0.004	0.23±0.009				
Bilirubin-Indirect	0.68 ± 0.005	0.56 ± 0.007	0.57 ± 0.001	0.57 ± 0.004	0.46 ± 0.003	0.52 ± 0.007				
Total Protein	6.63±0.035	6.15±0.031	6.2±0.062	6.13±0.019	6.11±0.015	6.56±0.019				
Albumin	3.45 ± 0.054	3.21±0.036	3.2±0.023	3.25±0.031	3.21±0.015	3.16±0.019				
Globulin	2.35±0.055	3.15±0.031	3.23±0.065	3.13±0.019	3.15±0.031	3.63±0.038				
A/G Ratio	1.16±0.003	1.06 ± 0.003	1.07 ± 0.005	1.03±0.006	1.03±0.006	1.07±0.005				

SGOT	81.31±0.055	152.2±0.023	152.2±0.059	152.4±0.065	152.5±0.019	132.4±0.061				
SGPT	16.76±0.038	31.45±0.031	31.23±0.080	31.13±0.019	31.11±0.015	31.2±0.057				
Alkaline Phosphatase	43.16±0.038	101.35±0.03	101.3±0.051	101.8±0.019	101.8±0.045	102.2±0.020				
	Biochemistry (Lipid Profile)									
Blood Glucose	81.26±0.054	80.33+0.019	80.23+0.050	82.15+0.020	80.16+0.096	82.36±0.038				
(Random)	81.20±0.034	80.33±0.019	80.23±0.030	82.13±0.020	80.10±0.090	82.30±0.038				
Serum CREATININE	0.7±0	0.58 ± 0.015	0.55 ± 0.031	0.56±0.019	0.54 ± 0.152	0.56±0.019				
S. Cholesterol	126±0.101	50.15±0.318	50.15±0.031	50.13±0.019	50.13±0.019	50.36±0.038				
S. Triglycerides	71.06±0.037	104.8±0.047	104.6±0.038	104.8±0.049	104.8±0.076	104.8 ± 0.038				
HDL Cholesterol	19.13±0.003	11.36±0.006	11.37±0.003	11.37±0.004	11.36±0.009	11.36±0.006				
LDL Cholesterol	62.55±0.002	24.20±0.004	24.20±0.003	24.25±0.001	24.21±0.009	24.21±0.009				
VLDL Cholesterol	14.23±0.052	23.24±0.001	23.24±0.002	23.21±0.003	23.24±0.001	23.54±0.001				
LDLC/HDLC Ratio	2.45±0.004	2.41±0.003	2.42±0.002	2.40±0.003	2.41±0.001	2.45±0.002				
TC/HDLC Ratio	3.68±0.003	9.54±0.005	9.5±0.007	9.51±0.003	9.74±0.001	9.53±0.005				
S. Cholesterol/HDLC Ratio	5±0	5±0	5±0	5±0	5±0	5±0				
Line and Manual CEM (()									

Value are Mean±SEM. (n=6).

3.4.5.3 Comparison of After 14 days Haematological parameters (Complete Blood Count, Liver Function Test, Lipid Profile) test reports with Mean±SEM.

Table 3.7: Effect of Nanoconstruct on Ha	Iaematological parameters (Complete	e Blood Count) in Albino Wistar
Rats.		

Parameters	Control	0.5 ml/kg	1.0 ml/kg	1.5 ml/kg	2.0 ml/kg	2.5 ml/kg
		Complet	e Blood Count			
Haemoglobin	15.41±0.065	15.05±0.031	15.1±0.074	15.05±0.031	13.6±0.038	15.25±0.155
WBC Count (10 ³ /uL)	10.28±0.154	7.46±0.019	7.43±0.019	7.2±0.023	7.38±0.036	7.53±0.019
Neutrophils	44.66±0.385	40.33±0.192	42±0.235	41.5±0.204	42.16±0.597	43±0.577
Lymphocytes	51.16±0.282	51.33±0.192	51.66±0.451	51.83±0.597	51.83±0.597	50.66±0.192
Eosinophil	5.66±0.190	4.66±0.192	5±0.235	4.5±0.311	5.33±0.192	5.33±0.192
Monocytes	1.66±0.190	2.16±0.152	2±0.235	1.66±0.192	1.83 ± 0.152	1.66±0.192
Basophils	0.11±0.015	0.13±0.019	0.13±0.019	0.21±0.015	0.16±0.038	0.13±0.019
RBC Count (10 ⁶ /uL)	6.41±0.006	6.37±0.005	6.35±0.004	6.16±0.005	6.37±0.005	6.37±0.005
Platelet Count	2.13±0.004	.3.84±0.005	3.43±0.005	3.42±0.006	3.84±0.003	3.84±0.001
Mean Platelet Value	10.23±0.038	9.21±0.003	9.23±0.008	9.14±0.001	9.23±0.009	9.23±0.009
Packed Cell Volume	41.33±0.038	39.45±0.031	39.5±0.023	39.25±0.031	39.23±0.050	39.46±0.019
Mean Corp. Vol.	73.22±0.006	63.15±0.001	68.14±0.002	68.36±0.001	68.11±0.009	68.12±0.010
Mean Corpuscular	21.43±0.006	25.74±0.003	25.63±0.045	25.74±0.001	25.41±0.003	25.72±0.010
Mean Corp. Hb Con.	35.14±0.005	43.70±0.002	42.73±0.006	42.71±0.001	42.23±0.008	42.41±0.009
Red Cells Destribution Width	14.36±0.045	15.38±0.036	14.31±0.068	14.43±0.019	14.43±0.019	14.6±0.147
2 courouron () fun	1	Biochemistry (Liver Function	Test)		
Bilirubin-Total	0.81±0.001	0.75±0.003	0.73±0.003	0.70±0.003	0.71±0.001	0.75±0.017
Bilirubin-Direct	0.25±0.003	0.26±0.003	0.26±0.004	0.21±0.001	0.26±0.001	0.24±0.002
Bilirubin-Indirect	0.72±0.006	0.55±0.006	0.56±0.004	0.56±0.003	0.55±0.010	0.56±0.003
Total Protein	6.55±0.039	6.46±0.019	6.38±0.008	6.46±0.050	6.46±0.019	6.15±0.020
Albumin	3.15±0.020	3.2±0.023	3.26±0.045	3.13±0.019	3.15±0.031	3.16±0.019
Globulin	2.23±0.019	3.13±0.019	3.13±0.019	3.28±0.068	3.2±0.023	3.15±0.020
A/G Ratio	155±0.003	1.07 ± 0.004	1.07±0.003	1.06±0.003	1.07 ± 0.005	1.07±0.001
SGOT	83.40±0.028	159.4±0.028	142.4±0.068	150.4±0.045	150.5±0.019	152.3±0.061
SGPT	12.21±0.028	31.15±0.031	31.28±0.111	31.13±0.019	31.13±0.019	31.21±0.068
Alkaline Phosphatase	38.78±0.028	102.6±0.031	101.6±0.049	100.1±0.019	103.7±0.038	101.3±0.057
		Biochemist	ry (Lipid Profi	le)		
Blood Glucose (Random)	82.18±0.028	81.2±0.023	81.31±0.068	79.28±0.049	80.33±0.019	80.3±0.057
Serum CREATININE	0.56±0.019	0.55±0.031	0.56±0.019	0.51±0.059	0.55±0.020	0.55±0.020
S. Cholesterol	140.15±0.02	50.4±0.023	50.45±0.051	50.35±0.031	50.33±0.038	50.13±0.019
S. Triglycerides	52.06±0.030	104.8±0.031	104.4±0.031	104.9±0.031	104.9±0.019	104.8±0.076
HDL Cholesterol	21.20±0.003	11.37±0.003	$10.0051.27 \pm$	11.31±0.003	11.97±0.001	11.38±0.006

LDL Cholesterol	65.52±0.007	24.18±0.002	24.17±0.003	24.17±0.003	24.17±0.004	24.20±0.002
VLDL Cholesterol	25.21±0.028	23.24±0.001	23.24±0.009	23.23±0.008	23.14±0.006	23.24±0.002
LDLC/HDLC Ratio	2.44 ± 0.003	2.41 ± 0.002	2.42 ± 0.004	2.42 ± 0.005	2.42 ± 0.011	2.43±0.006
TC/HDLC Ratio	3.57 ± 0.002	9.55±0.001	9.52 ± 0.006	9.52±0.009	9.51±0.003	9.51±0.001
S. Cholesterol/HDLC Ratio	5±0	5±0	5±0	5±0	5±0	5±0

Value are Mean±SEM. (n=6).

3.4.5.4 Comparison of After 21 days Haematological parameters (Complete Blood Count, Liver Function Test, Lipid Profile) test reports with Mean±SEM.

Table 3.8: Effect of Nanoconstruct or	Haematological	parameters (Complete	Blood	Count) in A	Albino V	Wistar
Rats.	_					

Parameters	Control	0.5 ml/kg	1.0 ml/kg	1.5 ml/kg	2.0 ml/kg	2.5 ml/kg	
1 di dilletti 5	Control		e Blood Count	1.0 III/Kg	2.0 III/Kg	2.0 m/ kg	
Haemoglobin	13.58±0.028	15.05±0.031	15.1±0.062	15.06±0.038	15.1±0.040	16.16±0.096	
WBC Count (10 ³ /uL)	10.51±0.028	5.5±0.023	5.6±0.023	5.36±0.050	5.35±0.045	5.66±0.019	
Neutrophils	44.5±0.311	42±0.235	41.83±0.366	41.83±0.597	42.33±0.509	47±0.577	
Lymphocytes	51.16±0.280	51.5±0.204	53.83±0.683	51.33±0.192	51.16±0.366	51±0.235	
Eosinophil	5.66±0.192	5±0.235	6.5±0.311	4.83±0.152	4.66±0.502	4.83±0.152	
Monocytes	2±0.235	1.66±0.192	2±0.235	1.66±0.192	1.66±0.192	1.83±0.152	
Basophils	0.15±0.020	0.13±0.019	0.18±0.028	0.13±0.019	0.16±0.038	0.83±0.152	
RBC Count (10 ⁶ /uL)	6.41±0.003	36.37±0.003	6.46±0.012	6.37±0.001	6.77±0.001	6.37±0.005	
Platelet Count	2.14±0.001	3.83±0.003	3.84±0.009	3.84±0.001	3.84±0.001	3.84±0.001	
Mean Platelet Value	10.21±0.028	9.25±0.002	9.25±0.002	9.24±0.001	9.22±0.009	9.25±0.001	
Packed Cell Volume	41.4±0.047	39.48±0.036	31.53±0.019	39.4±0.057	39.25±0.087	39.4±0.057	
Mean Corp. Vol.	73.41±0.002	68.15±0.001	62.15±0.003	68.14±0.001	68.14±0.008	69.21±0.052	
Mean Corpuscular	21.54±0.003	25.74±0.003	25.73±0.008	25.73±0.007	25.64±0.011	22.73±0.009	
Mean Corp. Hb Con.	35.15±0.003	42.71±0.007	40.71±0.008	42.31±0.009	42.73±0.009	42.70±0.001	
Red Cells Destribution Width	14.21±0.028	14.45±0.031	14.43±0.019	14.51±0.015	14.43±0.019	15.43±0.019	
	В	liochemistry (I	Liver Function	Test)			
Bilirubin-Total	0.84±0.001	0.73±0.004	0.75±0.003	0.72±0.001	0.73±0.003	0.73±0.003	
Bilirubin-Direct	0.25±0.002	0.26±0.003	0.27±0.002	0.26±0.003	0.26±0.003	0.25±0.010	
Bilirubin-Indirect	0.71±0.003	0.56 ± 0.001	0.56±0.003	0.55 ± 0.007	0.58 ± 0.006	0.56±0.003	
Total Protein	6.50±0.019	6.36±0.065	6.41±0.036	6.38±0.068	6.51±0.083	6.4±0.057	
Albumin	3.25±0.028	3.16±0.019	3.26±0.038	3.21±0.015	3.18±0.015	3.2±0.023	
Globulin	2.13±0.019	3.2±0.047	3.1±0.023	3.11±0.015	3.11±0.015	3.13±0.019	
A/G Ratio	1.55±0.003	1.07 ± 0.004	1.06 ± 0.005	1.08 ± 0.002	1.07±0.003	1.07 ± 0.005	
SGOT	81.41±0.028	151.3±0.065	105.4±0.057	142.5±0.019	153.4±0.049	132.5±0.019	
SGPT	12.16±0.019	31.15±0.031	31.23±0.076	31.13±0.019	31.16±0.030	31.13±0.019	
Alkaline Phosphatase	31.78±0.028	100.5 ± 0.050	107.5±0.069	108.7 ± 0.031	102.6±0.038	101.6±0.040	
Biochemistry (Lipid Profile)							
Blood Glucose (Random)	80.15±0.020	80.2±0.023	80.25±0.031	82.18±0.015	82.4±0.091	80.3±0.057	
Serum CREATININE	0.53±0.019	0.56±0.019	0.51±0.059	0.58±0.015	0.55±0.031	0.56±0.019	
S. Cholesterol	140.15±0.02	50.36±0.050	52.31±0.049	50.45±0.031	51.43±0.019	50.43±0.019	
S. Triglycerides	52.06±0.030	104.8±0.031	114.8±0.049	104.9±0.019	104.9±0.019	104.8±0.076	
HDL Cholesterol	21.21±0.005	11.37±0.005	11.37±0.005	11.37±0.005	11.37±0.003	13.36±0.006	
LDL Cholesterol	65.55±0.003	24.17±0.003	24.15±0.011	24.23±0.003	23.17±0.005	24.14±0.015	
VLDL Cholesterol	25.25±0.020	23.24±0.001	23.23±0.007	23.23±0.003	23.23±0.001	23.24±0.001	
LDLC/HDLC Ratio	2.40±0.001	2.41±0.001	2.41±0.003	2.41±0.002	2.42 ± 0.008	2.31±0.002	
TC/HDLC Ratio	3.57±0.004	9.56±0.001	9.42±0.002	9.52±0.007	9.57±0.011	9.53±0.006	
S. Cholesterol/HDLC Ratio	5±0	5±0	5±0	5±0	5±0	5±0	

Value are Mean±SEM. (n=6).

3.4.5.5 Comparison of After 28 days Haematological parameters (Complete Blood Count, Liver Function Test, Lipid Profile) test reports with Mean±SEM.

Table 3.9: Effect of Nanoconstruct on Haematological	parameters (Complete Blood Count) in Albino Wistar
Rats.	

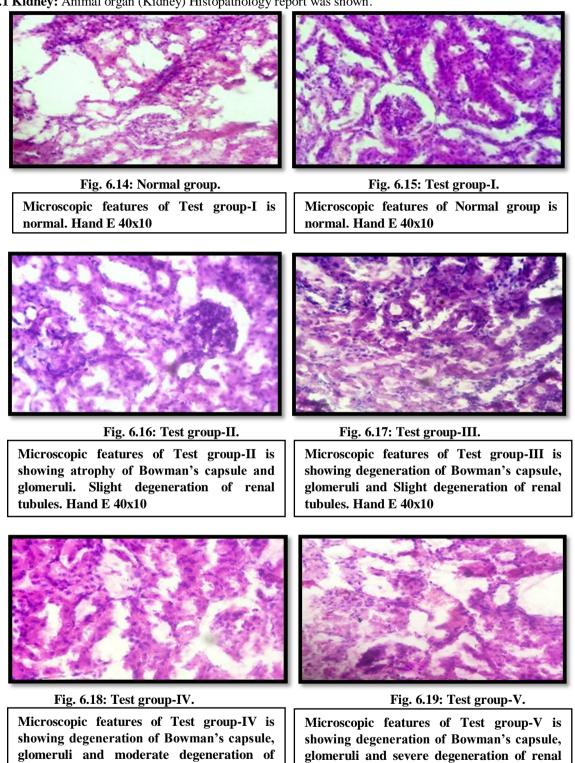
Parameters	Control	0.5 ml/kg	1.0 ml/kg	1.5 ml/kg	2.0 ml/kg	2.5 ml/kg	
			Blood Count			8	
Haemoglobin	12.26±0.140	15.05±0.031	14.5±0.031	16.48±0.106	16.06±0.038	16.16±0.096	
WBC Count (10 ³ /uL)	10.28±0.151	3.61±0.049	3.25±0.031	3.33±0.038	3.7±0.357	3.56±0.019	
Neutrophils	41.5±0.390	42±0.235	38.8±0.311	38.66±0.192	44±0.235	40.33±0.192	
Lymphocytes	51.83±0.280	51.5±0.311	51.5±0.311	54.33±0.192	53±0.577	54.33±0.192	
Eosinophil	5.33±0.304	5±0.235	4.16±0.597	4.83±0.152	5.16±0.152	5.16±0.152	
Monocytes	2±0.235	2±0.235	2.66±0.192	1.66±0.608	1.33±0.192	1.33±0.192	
Basophils	0.13±0.019	0.2±0.023	0.13±0.019	0.13±0.030	0.23±0.019	0.2±0.023	
RBC Count (10 ⁶ /uL)	6.49±0.057	6.36±0.006	6.17±0.003	5.26±0.009	6.17±0.005	6.27±0.005	
Platelet Count	2.12±0.007	3.94±0.003	3.66±0.003	3.54±0.001	3.42±0.007	3.64±0.003	
Mean Platelet Value	10.46±0.030	10.25±0.002	9.04±0.001	10.04±0.003	10.24±0.005	10.21±0.096	
Packed Cell Volume	41.16±0.030	39.46±0.019	37.13±0.019	39.13±0.019	39.23±0.076	39.23±0.076	
Mean Corp. Vol.	73.42±0.003	62.15±0.001	60.25±0.004	61.25±0.002	62.23±0.009	62.25±0.001	
Mean Corpuscular	21.49±0.032	25.75±0.002	24.46±0.010	24.47±0.004	23.46±0.006	25.47 ± 0.005	
Mean Corp. Hb Con.	35.17±0.008	40.70±0.003	42.92±0.005	40.80±0.003	41.92±0.001	40.93±0.005	
Red Cells Destribution Width	14.46±0.030	13.4±0.023	12.56±0.019	13.6±0.023	14.55±0.020	13.56±0.019	
	В	liochemistry (I	Liver Function	Test)			
Bilirubin-Total	0.85±0.003	0.71±0.001	0.74±0.003	0.70±0.003	0.72 ± 0.001	0.73±0.003	
Bilirubin-Direct	0.21±0.003	0.23±0.003	0.25 ± 0.001	0.11 ± 0.007	0.24 ± 0.005	0.21±0.004	
Bilirubin-Indirect	0.74±0.003	0.57 ± 0.002	0.50 ± 0.003	0.51 ± 0.006	0.55 ± 0.001	0.53±0.003	
Total Protein	6.53±0.030	6.4 ± 0.062	6.41±0.049	6.41±0.149	6.56±0.019	6.56±0.019	
Albumin	3.36±0.038	3.16±0.019	3.15±0.031	3.35±0.031	3.33 ± 0.038	3.43±0.019	
Globulin	2.21±0.054	3.15±0.031	3.16±0.030	3.3±0.057	3.15 ± 0.020	3.23±0.019	
A/G Ratio	1.14 ± 0.004	1.07 ± 0.003	1.05 ± 0.003	1.05 ± 0.001	1.05 ± 0.001	1.05 ± 0.001	
SGOT	81.16±0.030	132.4 ± 0.031	139.4±0.068	127.4±0.076	157.6±0.031	137.4 ± 0.080	
SGPT	12.46±0.030	31.18±0.059	33.2±0.047	30.16±0.038	30.13±0.019	30.13±0.019	
Alkaline Phosphatase	31.16±0.030	104.6±0.049	102.6±0.038	103.6±0.090	102.7±0.019	101.6±0.038	
Biochemistry (Lipid Profile)							
Blood Glucose (Random)	81.35±0.020	81.2±0.023	80.23±0.019	83.4±0.115	81.5±0.115	83.35±0.093	
Serum CREATININE	0.51±0.043	0.56 ± 0.019	0.46 ± 0.019	0.58 ± 0.015	0.56 ± 0.019	0.56±0.019	
S. Cholesterol	130.16±0.03	56.45±0.031	57.41±0.015	68.33±0.038	59.33±0.038	58.43±0.019	
S. Triglycerides	72.06±0.030	104.2±0.023	101.2±0.023	106.2±0.023	106.2±0.019	106.3±0.057	
HDL Cholesterol	21.22±0.004	11.37±0.003	11.63±0.003	12.67±0.005	11.68 ± 0.001	11.67 ± 0.004	
LDL Cholesterol	64.55±0.002	24.16±0.006	25.43±0.007	25.47±0.003	25.07±0.005	25.45±0.013	
VLDL Cholesterol	25.22±0.028	23.23±0.004	21.22±0.014	24.25±0.012	21.22±0.007	21.24±0.001	
LDLC/HDLC Ratio	2.43±0.004	2.47 ± 0.004	2.11±0.004	2.13±0.004	2.37 ± 0.005	2.18±0.003	
TC/HDLC Ratio	3.54±0.003	9.58±0.004	9.33±0.019	9.01±0.002	10.24±0.089	9.09±0.001	
S. Cholesterol/HDLC Ratio	5±0	5±0	5±0	5±0	5±0	5±0	

Value are Mean±SEM. (n=6).

tubules along with Hemorrhage. Hand E

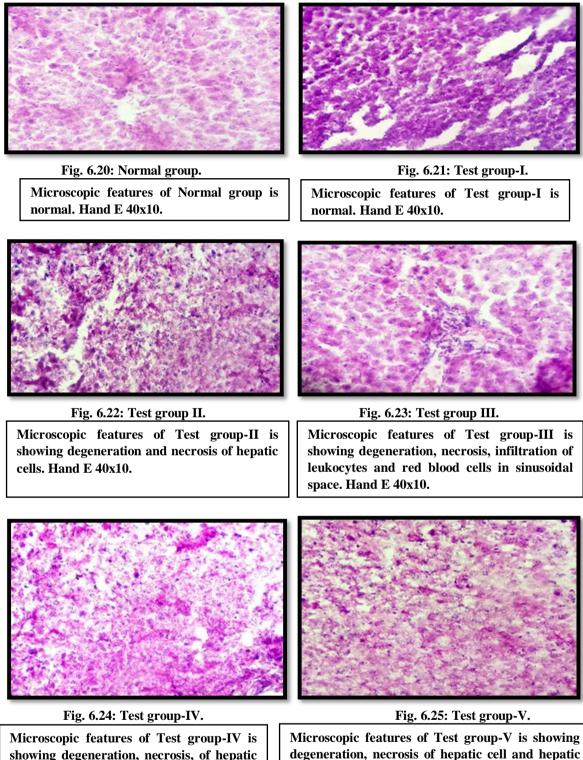
3.4.5 Histopathology:

3.4.6.1 Kidney: Animal organ (Kidney) Histopathology report was shown.



renal tubules. Hand E 40x10

3.4.6.2 Liver: Animal organ (Liver) Histopathology report was shown.



showing degeneration, necrosis, of hepatic cell and hepatic nuclei. Fatty degeneration are also observed. Hand E 40x10.

4. CONCLUSION

In conclusion, the developed formulations Nanoconstructs were found to be safer in less amount and mortality activity shown in more amount shown in Haematological and Histopathological reports. Toxicity

nuclei. Nucleus become disintegrated and hydropic degeneration are also observed. Hand E 40x10.

of a formulation is a primary concern for its future applications and the developed formulations may prove their capabilities *In-vivo* due to toxicity. A part from toxicity the formulation must be stable enough to withstand various environmental conditions to give its highest performance whenever used during its self-life. The prepared formulations were stable enough in normal storage conditions e.g. at room temperature, assuring its easy storage and transportation, if so, in future.

The given Nanoconstruct was found safe in less amount but more amount in mortality shown, toxicity studies in Albino Wistar Rats. The toxicity data of Nanoconstructs may be useful for further development in pre-clinical and clinical studies.

5. REFERENCES

- Hameed A.S. Sahul, Vimal S., Majeed S. Abdul, Taju G., Nambi K.S.N., Raj N. Sundar, Madan N., Farook M.A., Rajkumar T., Gopinath D., Chitosan tripolyphosphate (CS/TPP) nanoparticles: Preparation, characterization and application for gene delivery in shrimp, Acta Tropica, 2013; 128: 486– 493.
- Ziyong Zhang, Zonghua Liu, Yanpeng Jiao, Yifei Wang, Changren Zhou, Polysaccharides-based nanoparticles as drug delivery systems, Advanced Drug Delivery Reviews, 2008; 60: 1650–1662.
- C. Vyvyan Howard, Andreas Elsaesser, Toxicology of nanoparticles, Advanced Drug Delivery Reviews, 2012; 64: 129–137.
- Yanyun Zhao, Hongcai Zhang, Preparation, characterization and evaluation of tea polyphenole Zn complex loaded b-chitosan nanoparticles, Food Hydrocolloids, 2015; 48: 260e273.
- Lucimara Gaziola de la Torrea, Caroline Casagrande Sipoli, Nathalia Santana, Andrea Arruda Martins Shimojo, Adriano Azzoni, Scalable production of highly concentrated chitosan/TPP nanoparticles indifferent pHs and evaluation of the in vitro transfection efficiency, Biochemical Engineering Journal, 2015; 94: 65–73.
- Christian Remesy, Claudine Manach, Christine Morand, Christian Demigne, Odile Texier, Françoise Regerat, Bioavailability of rutin and quercetin in rats, FEBS Letters, 1997; 409; 12-16.
- Bismita Nayak, Debasis Nayak, Aliva Prity Minz, Sarbani Ashe, Pradipta Ranjan Rauta, Manisha Kumari, Pankaj chopra, Synergistic combination of antioxidants, silver nanoparticles and chitosan in a nanoparticle based formulation: Characterization and cytotoxic effect on MCF-7 breast cancer cell lines, Journal of Colloid and Interface Science, 2016.
- Alessandro F. Martins, Daiane M. de Oliveira, Antonio G.B. Pereira, Adley F. Rubira, Edvani C. Muniz, Chitosan/TPP microparticles obtained by microemulsion method applied in controlled release of heparin, International Journal of Biological Macromolecules, 2012; 51: 1127–1133.
- Aws Alshamsan, Mohd Abul Kalam, Abdul Arif Khan, Shahanavaj Khan, Abdulaziz Almalik, Optimizing Indomethacin-Loaded Chitosan Nanoparticle Size, Encapsulation, and Release Using Box-Behnken Experimental Design,

International Journal of Biological Macromolecules, 2016.

- Peter X. Ma, Yaobin Wu, Ling Wang, Baolin Guo, Yongpin Shao, Electroactive biodegradable polyurethane significantly enhanced Schwann cells myelin gene expression and neurotrophin secretion for peripheral nerve tissue engineering, Biomaterials, 2016; 87: 18e31.
- 11. Jie Dou, Changlin Zhou, Lingman Ma, Yanrong Wang, Mengxiao Wang, Yuwei Tian, Wei Kang, Hanhan Liu, Hui Wang, Effective antimicrobial activity of Cbf-14, derived from a cathelin-like domain, against penicillin-resistant bacteria, Biomaterials, 2016; 87: 32e45.
- 12. Guangjin Pan, Shicheng Wei, Ping Zhou, Fujian Wu, Tiancheng Zhou, Xiujuan Cai, Siqi Zhang, Xiaohong Zhang, Qiuhong Li, Yongliang Li, Yunfei Zheng, Mengke Wang, Feng Lan, Duanqing Pei, Simple and versatile synthetic polydopamine-based surface supports reprogramming of human somatic cells and long-term self-renewal of human pluripotent stem cells under defined conditions, Biomaterials, 2016; 87: 1e17.
- Zi-Rong Xu, Wen-Li Du, Shan-Shan Niu, Ying-Lei Xu, Cheng-Li Fan, Antibacterial activity of chitosan tripolyphosphate nanoparticles loaded with various metal ions, Carbohydrate Polymers, 2009; 75: 385– 389.
- R. Jayakumar, K.P. Chennazhi, R.A.A. Muzzarelli, H. Tamura, S.V. Nair, N. Selvamurugan, Chitosan conjugated DNA nanoparticles in gene therapy, Carbohydrate Polymers, 2010; 79: 1–8.
- 15. Qin Wang, Yangchao Luo, Boce Zhang, Wen-Hsing Cheng, Preparation, characterization and evaluation of selenite-loaded chitosan/TPP nanoparticles with or without zein coating, Carbohydrate Polymers, 2010; 82: 942–951.
- Safaa K.H. Khalil, Gina S. El-Feky, Sally T. El-Banna, Wafaa A. Khalil, Preparation and evaluation of warfarin-_-cyclodextrin loaded chitosan nanoparticles for transdermal delivery, Carbohydrate Polymers, 2012; 90: 1244–1253.
- Mosaad A. Abdel-Wahhab, Abdulhadi Aljawish, Aziza A. El-Nekeety, Sekena H. Abdel-Aiezm, Heba A.M. Abdel-Kader, Bertrand H. Rihn, Olivier Joubert, Chitosan nanoparticles and quercetin modulate gene expression and prevent the genotoxicity of aflatoxinB1in rat liver, Toxicology Reports, 2015; 2: 737–747.
- Luiz H.C. Mattoso, Marcia R. de Moura, Fauze A. Aouada, Roberto J. Avena-Bustillos, Tara H. McHugh, John M. Krochta, Improved barrier and mechanical properties of novel hydroxypropyl methylcellulose edible films with chitosan/tripolyphosphate nanoparticles, Journal of Food Engineering, 2009; 92: 448–453.
- 19. Quan Gan, Tao Wang, Colette Cochrane, Paul McCarron, Modulation of surface charge, particle size and morphological properties of chitosan–TPP

nanoparticles intended for gene delivery, Colloids and Surfaces B: Biointerfaces, 2005; 44: 65–73.

- 20. Brabu Balusamy, Yamuna Gowri Kandhasamy, Anitha Senthamizhan, Gopalakrishnan Chandrasekaran, Murugan Siva Subramanian, Kumaravel Tirukalikundram S, Characterization and bacterial toxicity of lanthanum oxide bulk and nanoparticles, *Journal of Rare Earths*, Dec. 2012; 30(12): 1298.
- 21. Y. Martin Lo, Meng Li, Jun-Jie Yin, Wayne G. Wamer, Mechanistic characterization of titanium dioxide nanoparticle-induced toxicity using electron spin Resonance, journal of food and drug analysis, 2014; 22: 76e85.
- 22. Nguyen T.K. Thanh, Lara Yildirimer, Marilena Loizidou, Alexander M. Seifalian, Toxicological considerations of clinically applicable Nanoparticles, Nano Today, 2011; 6: 585-607.
- Wing-Hin Lee, Ching-Yee Loo, Daniela Traini, Paul M. Young, Inhalation of nanoparticle-based drug for lung cancer treatment: Advantages and challenges, asian journal of pharmaceuticals ciences, 2015; 10; 481–489.
- 24. Chun-Ho Kim, Eunsun Lee, Sang Jun Park, Jae Ho Lee, Min Sup Kim, Preparation of chitosan–TPP nanoparticles and their physical and biological properties, asian journal of pharmaceuticals ciences 2015.
- 25. J. Guan, P. Cheng, S.J. Huang, J.M. Wu, Z.H. Li, X.D. You, L.M. Hao, Y. Guo, R.X.Li, H. Zhang, Optimized Preparation of Levofloxacin-loaded Chitosan Nanoparticles by Ionotropic Gelation, Physics Procedia, 2011; 22: 163 – 169.
- Asgar Ali, Ameeduzzafar, Javed Ali, Development and validation of UPLC/ESI-Q-TOF-MS for carteolol in aqueous humour: Stability, stress degradation and application in pharmacokinetics of nanoformulation, Arabian Journal of Chemistry, 2013.
- 27. Tapan Kumar Girin, Amrita Thakur, Amit Alexander, Ajazuddin, Hemant Badwaik, Dulal Krishna Tripathi, Modified chitosan hydrogels as drug delivery and tissue engineering systems: present status and applications, Acta Pharmaceutica Sinica B, 2012; 2(5): 439–449.
- Xiaojie Cheng, Xiguang Chen, Chao Feng, Zhiguo Wang, Changqing Jiang, Ming Kong, Xuan Zhou, Yang Li, Chitosan/o-carboxymethyl chitosan nanoparticles for efficient and safe oral anticancer drug delivery: In vitro and in vivo evaluation, International Journal of Pharmaceutics, 2013; 457: 158–167.
- Jingkun Jiang, Gu nter Oberdo rster, Pratim Biswas, Characterization of size, surface charge, and agglomeration state of nanoparticle dispersions for toxicological studies, J Nanopart Res, 2009; 11: 77– 89.
- Gaurav K. Jain, Nilu Jain, Shadab A. Pathan, Sohail Akhter, Sushma Talegaonkar, Prakash Chander, Roop K. Khar, Farhan J. Ahmad, Ultra high-

pressure liquid chromatographic assay of moxifloxacin in rabbit aqueous humor after topical instillation of moxifloxacin nanoparticles, Journal of Pharmaceutical and Biomedical Analysis, 2010; 52: 110–113.

- 31. Jun-min Zheng, Yan Pan, Ying-jian Li, Hui-ying Zhao, Hui Xu, Gang Wei, Jin-song Hao, Fu-de Cui, Bioadhesive polysaccharide in protein delivery system: chitosan nanoparticles improve the intestinal absorption of insulin in vivo, International Journal of Pharmaceutics, 2002; 249: 139-147.
- 32. J. Guan, P. Cheng, S.J. Huang, J.M. Wu, Z.H. Li, X.D. You, L.M. Hao, Y. Guo, R.X. Li, H. Zhang, Optimized Preparation of Levofloxacin-loaded Chitosan Nanoparticles by Ionotropic Gelation, Physics Procedia, 2011; 22: 163 – 169.
- 33. Dongzhi Hou, Zuyong Feng, Ruyi Gui, Sheng Hu, Yi Huang, Qineng Ping, Preparation and Characterization of Novel Drug-Inserted-Montmorillonite Chitosan Carriers for Ocular Drug Delivery, Advances in Nanoparticles, 2015; 4: 70-84.
- 34. Aleksandra B. Djurisic, Yu Hang Leung, Alan M. C. Ng, Xiao Ying Xu, Patrick K. H. Lee, Natalie Degge, and R. S. S. Wu, Toxicity of Metal Oxide Nanoparticles: Mechanisms, Characterization, and Avoiding Experimental Artefacts, Wiley-VCH Verlag GmbH and Co. KGaA, Weinheim, small, 2015; 11(1): 26–44.
- 35. Mark Hermann Rümmeli, Rafael Gregorio Mendes, Britta Koch, Alicja Bachmatiuk, Ahmed Aboud El-Gendy, Yulia Krupskaya, Armin Springer, Rüdiger Klingeler, Oliver Schmidt, Bernd Büchner, Samuel Sanchez, Synthesis and toxicity characterization of carbon coated iron oxide nanoparticles with highly defined size distributions, Biochimica et Biophysica Acta, 2014; 1840: 160–169.
- 36. Santosh Kumar Kar, Feroz Akhtar, M. Moshahid Alam Rizvi, Oral delivery of curcumin bound to chitosan nanoparticles cured Plasmodium yoelii infected mice, Biotechnology Advances, 2012; 30: 310–320.
- 37. Hong Ni, Wen Fan, Wei Yan, Zushun Xu, Formation mechanism of monodisperse, low molecular weight chitosan nanoparticles by ionic gelation technique, Colloids and Surfaces B: Biointerfaces, 2012; 90: 21– 27.
- 38. Ilaiyaraja Nallamuthu, Aishwarya Devi, Farhath Khanum, Chlorogenic acid loaded chitosan nanoparticles with sustained release property, retained antioxidant activity and enhanced bioavailability, asian journal of pharmaceutical sciences, 2015; 10: 203e211.
- Schlede E., Mischke U., Roll R. and Kayser D. A National Validation Study of the Acute-Toxic-Class Method – An Alternative to the LD50 Test. Arch. Toxicol, 1992; 66: 455-470.
- 40. Virender K. Sharma, Jan Filip, Radek Zboril and Rajender S. Varma, Natural inorganic nanoparticles

- formation, fate, and toxicity in the environment, *Chem. Soc. Rev.*, 2015, 44: 8410-8423,

- Maria Jose Alonso, Kevin A. Janes, Marie P. Fresneau, Ana Marazuela, Angels Fabra, Chitosan nanoparticles as delivery systems for doxorubicin, Journal of Controlled Release, 2001; 73: 255–267.
- 42. Dimitrios Bikiaris, Sofia Papadimitriou, Konstantinos Avgoustakis, Evangelos Karavas, Manolis Georgarakis, Chitosan nanoparticles loaded with dorzolamide and pramipexole, Carbohydrate Polymers, 2008; 73: 44–54.
- 43. L. Harivardhan Reddy, Jose L. Arias, Julien Nicolas, and Patrick Couvreur, Magnetic Nanoparticles: Design and Characterization, Toxicity and Biocompatibility, Pharmaceutical and Biomedical Applications, American Chemical Society, 2012.
- 44. Erem Bilensoy, Can Sarisozena, Gunes, Esendagli, A. Lale Do gan, Yesim Aktas, Murat Sen, N. Aydın Mungan, Intravesical cationic nanoparticles of chitosan and polycaprolactone for the delivery of Mitomycin C to bladder tumors, International Journal of Pharmaceutics, 2009; 371:170–176.
- 45. Lipnick R L, Cotruvo, J A, Hill R N, Bruce R D, Stitzel K A, Walker A P, Chu I; Goddard M, Segal L, Springer J A and Myers R C Comparison of the Up-and Down, Conventional LD50, and Fixed Dose Acute Toxicity Procedures. Fd. Chem. Toxicol, 1995; 33: 223-231.
- 46. Ryan C. Gott, Yangchao Luo, Qin Wang, William O. Lamp, Development of a biopolymer nanoparticle-based method of oral toxicity testing in aquatic invertebrates, EcotoxicologyandEnvironmentalSafety, 2014; 104: 226–230.
- Sanghoon Ko, Sae-Yeol-Rim Paik, Jong-Seok Kim, Sung Jae Shin, Characterization, Quantification, and Determination of the Toxicity of Iron Oxide Nanoparticles to the Bone Marrow Cells, *Int. J. Mol. Sci*, 2015; 16.
- 48. Mary Ann Foglio, Leila Servat-Medina, Alvaro Gonzalez-Gómez, Felisa Reyes-Ortega, Ilza Maria Oliveira Sousa, Nubia de Cassia Almeida Queiroz, Patricia Maria Wiziack Zago, Michelle Pedrosa Jorge, Karin Maia Monteiro, Joao Ernesto de Carvalho, Julio Chitosan-San Roman, tripolyphosphate nanoparticles as Arrabidaea chica standardized extract carrier: synthesis. characterization, biocompatibility, and antiulcerogenic activity, International Journal of Nanomedicine, 2015; 10: 3897-3909.
- Wim Jiskoot, Maryam Amidi, Stefan G. Romeijn, Gerrit Borchard, Hans E. Junginger, Wim E. Hennink, Preparation and characterization of protein-loaded N-trimethyl chitosan nanoparticles as nasal delivery system Journal of Controlled Release, 2006; 111: 107 – 116.
- 50. Siddhartha Shrivastava, Tanmay Bera, Arnab Roy, Gajendra Singh, P Ramachandrarao and Debabrata Dash, Characterization of enhanced antibacterial

effects of novel silver Nanoparticles, Nanotechnology, 2007; 18: 225103 (9pp).

- 51. Qin Wang, Yangchao Luo, Boce Zhang, Wen-Hsing Cheng, Preparation, characterization and evaluation of selenite-loaded chitosan/TPP nanoparticles with or without zein coating, Carbohydrate Polymers, 2010; 82: 942–951.
- 52. Diener W., and Schlede E. Acute Toxicity Class Methods: Alternatives to LD/LC50 Tests. ALTEX, 1999; 16: 129-134.
- 53. Mosaad A. Abdel-Wahhab, Ezzeldeen S. El-Denshary, Abdulhadi Aljawish, Aziza A. El-Nekeety, Nabila S. Hassan, Raghda H. Saleh, Bertrand H. Rihn, Possible Synergistic Effect and Antioxidant Properties of Chitosan Nanoparticles and Quercetin against Carbon Tetrachloride-Induce Hepatotoxicity in Rats, Soft Nanoscience Letters, 2015; 5: 36-51.
- 54. David B. Warheit, Toxicological Highlight, How Meaningful are the Results of Nanotoxicity Studies in the Absence of Adequate Material Characterization? Toxicological Sciences, 2008; 101(2): 183–185.
- 55. OECD. (2006). Report of the Validation of the Updated Test Guideline 407: Repeat Dose 28-day Oral Toxicity Study in Laboratory Rats. Series on Testing and Assessment No 59, ENV/JM/MONO, 2006; 26.
- 56. OECD. (2002). Detailed Review Paper on the Appraisal of Test Methods for Sex Hormone Disrupting Chemicals. Series on Testing and Assessment No 21, ENV/JM/MONO, 2002; 8.
- 57. OECD Guidance Document on the Recognition, Assessment and Use of Clinical Signs as Humane Endpoints for Experimental Animals Used in Safety Evaluation Environmental Health and Safety Monograph Series on Testing and Assessment, 2000; 19.