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EVALUATION AND VALIDATION OF A UPLC METHOD FOR THE STABILITY INDICATING ASSAY OF MIDOSTAURIN IN IN CAPSULE DOSAGE FORM USING UPLC-MS/MSⁿ

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ABSTRACT

This proposal provides a comprehensive review of the optimization and approval of the LCMS technology for evaluating midostaurin, a chemical that potentially be a tyrosine kinase inhibitor. Analysts had to choose a permanent and movable stage, as well as develop standard stock and diluent arrangements, in order to find the best circumstances for division. The study indicated that a BDS C18 provided the best determination and affectability when combined with slope programming, using a portable stage of methanol and acetonitrile (80:20 v/v). By analyzing the method's precision, linearity, adaptability, and exactness, the panel also looked at the method's validity. Considerations of accuracy showed respectable recoveries, with a brutality level of 99.28%. There were factually irrelevant ponders of correctness that proved the system and approach were accurate.

KEYWORDS: Midostaurin, generation of degradation products and evaluation of the analytical method.

INTRODUCTION

Ensuring the quality, safety, and effectiveness of pharmaceutical products relies heavily on the development and validation of analytical techniques in the field of pharmaceutical analysis. Rigid and exact procedures are required for the quantitative analysis of APIs in bulk dosage forms in order to fulfill regulatory criteria. A technique for the bulk dosage form estimation of midostaurin using Ultra-Performance Liquid Chromatography (UPLC) is the subject of this study's assessment and validation.

The tyrosine kinase inhibitor midostaurin has recently been the subject of much interest due to its possible therapeutic use, mainly in the management of certain types of blood cancers. For the sake of both therapeutic efficacy and regulatory compliance, it is essential to accurately determine the midostaurin content in bulk medicinal preparations.

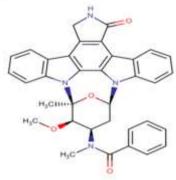
The benefits of ultra-performance liquid chromatography (UPLC), such as high resolution, fast analysis, and enhanced sensitivity, have led to its growing popularity in pharmaceutical analysis. To guarantee accuracy, dependability, and repeatability of findings, a thorough validation procedure is required for the effective deployment of UPLC techniques.

In order to estimate midostaurin in bulk dosage forms, this project will first develop a reliable UPLC technique and then evaluate its analytical performance. To determine whether the technique is suitable for regular pharmaceutical analysis, the validation procedure takes into account a number of factors, including specificity, precision, accuracy, linearity, robustness, and system appropriateness. Findings from this study have implications for both midostaurin analytical technique and pharmaceutical analysis more generally, highlighting the need for proven procedures to guarantee the security and efficacy of pharmaceuticals.

Drug Profile

| Therapeutic category | Antineoplastic Agent |
|----------------------|---------------------------|
| CAS Registry number | 120685-11-2 |
| Chemical name | 4'-N-benzoylstaurosporine |
| Molecular formula | $C_{35}H_{30}N_4O_4$ |
| Molecular Weight | 570.649 |
| Solubility | \geq 2.5 mg/mL |
| pka | 13.45 |
| λ_{\max} | 292 nm |

MIDOSTAURIN Chemical Structure of Midostaurin



Experimental Methodology Method Validation

What we mean when we talk about "the analytical technique" is the method by which the analysis is carried

RESULTS

Validation of Forced Degradation for Midostaurin Evaluation of Methods Assay Studies

| Recovery level | Set No. | Midostaurin | | |
|----------------|---------|-------------------|----------------------|--|
| | | Wt. Taken (μg/ml) | Amount found (µg/ml) | |
| 50% | Set 1 | 02.04 | 02.02 | |
| | Set 2 | 02.06 | 02.03 | |
| | Set 3 | 02.08 | 02.07 | |
| 100% | Set 1 | 04.12 | 04.11 | |
| | Set 2 | 04.14 | 04.13 | |
| | Set 3 | 04.16 | 04.15 | |
| 150% | Set 1 | 06.14 | 06.13 | |
| | Set 2 | 06.16 | 06.15 | |
| | Set 3 | 06.18 | 06.17 | |

Accuracy study for Analytical Method Validation (Midostaurin)

System Precision

Procedure

The parameters, retention time (RT), theoretical plates (N), tailing factor (T), peak asymmetry (As) and repeatability were evaluated at a concentration of $4\mu g/mL$ (Midostaurin).

| Parameters | Midostaurin |
|------------------------------|--------------------|
| Retention time (min) ± % RSD | 8.775 ± 0.04 |
| Theoretical plates ± % RSD | 2377.56 ± 0.50 |
| Asymmetry ± % RSD | 1.08 ± 0.05 |
| Repeatability (% RSD) | 0.11 |

System Precision (Midostaurin)

out. All of the analytical procedures should be spelled out in great detail. The sample, the reference standard, and the reagents, as well as their preparations, the use of the equipment, the development of the calibration curve, the application of the formulas for the calculation, etc. There has been comprehensive validation of the disclosed technique for its specificity, system appropriateness, linearity, accuracy, precision, limit of detection, limit of quantification, and robustness.

Robustness

Procedure

- 1. Change in flow rate
- 2. Change in temperature
- 3. Change in wave length

The robustness was studied by evaluating the effect of small but deliberate variations in the chromatographic conditions. The conditions studied were flow rate (\pm 0.05), column temperature (\pm 2°C) and wavelength of detection (\pm 5 nm). The result of robustness study of the developed assay method was established. The result shown that during all variance conditions, assay value of the test preparation solution was not affected and it was in accordance with that of actual. System suitability parameters were also found satisfactory; hence the analytical method would be concluded as robust.

| Robustness Studies | | | | | |
|--------------------|--------|-----------|-------|--|--|
| Parameter | Value | Peak Area | % RSD | | |
| | Low | 27548 | | | |
| Flow Rate | Actual | 27592 | 0.13% | | |
| | Plus | 27619 | | | |
| | | | | | |
| | Low | 27563 | | | |
| Temperature | Actual | 27646 | 0.16% | | |
| _ | Plus | 27629 | | | |
| | | | | | |
| Wavelength | Low | 27548 | | | |
| | Actual | 27593 | 0.14% | | |
| | Plus | 27626 | | | |

Robustness Studies (Midostaurin)

Ruggedness

Intraday precision (Repeatability): Intraday Precision was performed and % RSD for Midostaurin was 0.11%."

Inter day precision: Inter day precision was performed with 24 hrs time lag and the %RSD Obtained for Midostaurin was 0.15%.

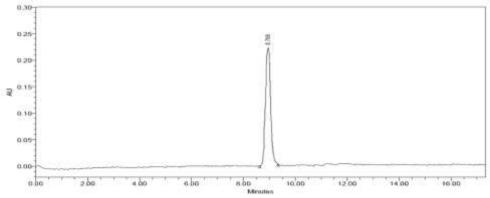
| Midostaurin | | | | |
|---------------------------|-----------|---------|--------|--|
| Ruggedness | | | | |
| Parameter | Peak Area | % RSD | %LC | |
| Intraday precision | 27158 | | 99.77% | |
| | 27212 | 0.11% | 99.97% | |
| | 27164 | U.11 70 | 99.79% | |
| Inter day precision | 27145 | | 98.72% | |
| | 27229 | 0.15% | 99.96% | |
| | 27182 | 0.15% | 99.86% | |
| T / / / | 27118 | | 99.62% | |
| Instrument:1 | 27139 | 0.04% | 99.70% | |
| Acquity UPLC Waters,2695H | 27121 | 0.04 70 | 99.63% | |
| Instrument:2 | 27117 | | 99.62% | |
| | 27134 | 0.04% | 99.68% | |
| Agilent Technologies,1290 | 27117 | 0.04 70 | 99.62% | |
| | | | | |
| Average | | • | 99.66 | |
| Std.Dev | | | 0.322 | |
| %RSD | | | 0.32% | |

Ruggedness Parameters (Midostaurin)

Forced Degradation Studies

Sample Control: An accurate 10 ml of the prepared pure drug stock solution of working standard was transferred to a clean and dry RBF. The volume of the sample was

 $100\mu g/ml$. It was injected into the UPLC system against a blank of Methanol and Acetonitrile in the ratio of 80:20 % v/v after optimizing the mobile phase composition, chromatogram was recorded.

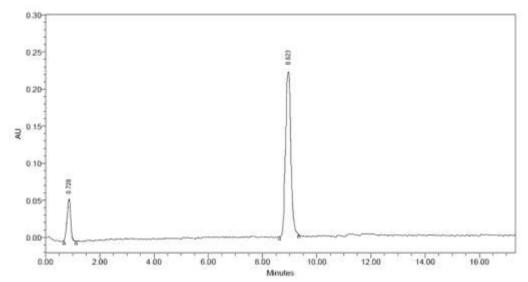


Assay of Midostaurin (Sample Control)

a. Acidic Degradation

An accurate 10 ml of pure drug sample solution was transferred to a clean and dry round bottom flask (RBF). 30 ml of 0.1 N HCl was added to it. It was refluxed in a water bath at 60°C for 4 hours. Drug became soluble after reflux which was insoluble initially. Allowed to cool at room temperature. The sample was then

neutralized using 2N NaOH solution and final volume of the sample was made up to 100ml with water to prepare 100ppm solution. It was injected into the UPLC system against a blank of Methanol and Acetonitrile in the ratio of 80:20 %v/v after optimizing the mobile phase composition, chromatogram was recorded and shown in Chromatogram."

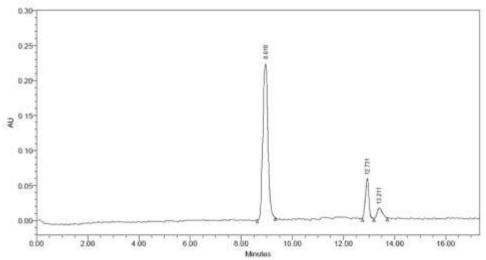


Chromatogram showing the degraded products in Acidic degradation

b. Basic Degradation

An accurate 10 ml of pure drug sample solution was transferred to a clean and dry RBF. 30 ml of 0.1N NaOH was added to it. It was refluxed in a water bath at 60°C for 4 hours. Drug became soluble after reflux which was insoluble initially. It was allowed to cool at room temperature. The sample was then neutralized using 2N

HCl solution and final volume of the sample was made up to 100ml with water to prepare 100ppm solution. It was injected into the UPLC system against a blank of Methanol and Acetonitrile in the ratio of 80:20 %v/v after optimizing the mobile phase composition, chromatogram was recorded and shown in Chromatogram."



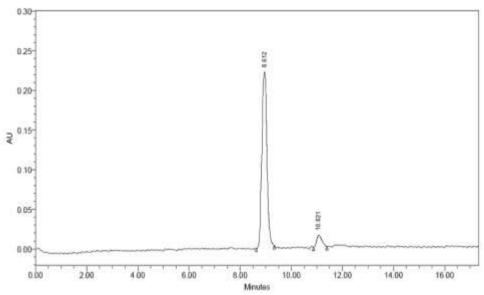
Chromatogram showing the degraded products in Basic degradation

c. Wet heat degradation

Accurate 10 ml of pure drug sample was transferred to a clean and dry RBF. 30 ml of HPLC grade water was added to it. Then, it was refluxed in a water bath at 60°C for 6 hours uninterruptedly. After the completion of

reflux, the drug became soluble and the mixture of drug and water was allowed to cool at room temperature. Final volume was made up to 100 ml with HPLC grade water to prepare 100 ppm solution. It was injected into the UPLC system against a blank of Methanol and

Acetonitrile in the ratio of 80:20 %v/v after optimizing the mobile phase composition, chromatogram was recorded and shown in Chromatogram.

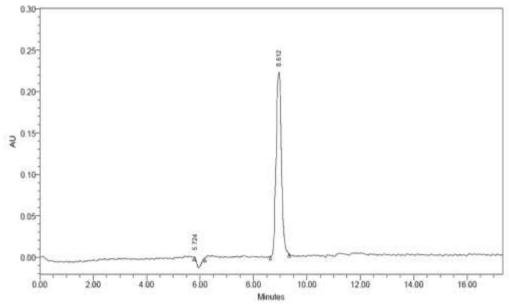


Chromatogram showing the degraded products in Wet heat degradation

d. Oxidation with (3%) H₂O₂

Approximately 10 ml of pure drug sample was transferred in a clean and dry 100 ml volumetric flask. 30 ml of $3\%~H_2O_2$ and a little methanol was added to it to make it soluble and then kept as such in dark for 24

hours. Final volume was made up to 100 ml using water to prepare 100 ppm solution. The above sample was injected into the UPLC system. The chromatogram was recorded and shown in Chromatogram."



Chromatogram showing the degraded products in Oxidative degradation

| Nature of Stress | Degradation condition | Time(h) | Number of degradation products (Rt) |
|------------------|-----------------------|---------|-------------------------------------|
| Acidic | 60°C | 3 | 1 (0.728) |
| Basic | 60°C | 9 | 2 (12.731, 13.211) |
| Oxidative | RT | 48 | 1 (5.724) |
| Wet Heat | 105°C | 24 | 1 (10.821) |

Summary of Forced Degradation Studies (Midostaurin)

Forced Degradation Studies Analysis of Midostaurin

| Conditions | Sample Amount (µg/ml) | Peak Area | % claim | % Degradation |
|-----------------------|-----------------------|-----------|---------|---------------|
| Sample Control | 04.15 | 26139 | 91.69% | - |
| Acidic Degradation | 04.08 | 24721 | 86.73% | 4.96% |
| Basic Degradation | 04.05 | 23581 | 82.72% | 8.97% |
| Oxidative Degradation | 04.03 | 24357 | 85.44% | 6.25% |
| Wet Heat | 04.06 | 25832 | 90.62% | 1.07% |

Results of Forced Degradation Assays (Midostaurin) Calculation formula for Midostaurin

$$\% \, Assay = \frac{AT}{AS} \times \frac{W1}{100} \times \frac{1}{25} \times \frac{100}{W2} \times \frac{25}{1} \times \frac{AW}{LC} \times P$$

Whereas,"

AT = Average area of test preparation, 26139"

AS = Average area of standard preparation, 28358"

W1 = Weight taken of reference standard (μg), 04.15"

W2 = Weight taken of test sample (µg), 04.25

AW = Average weight of sample (μg), 3057"

LC = Label claim (µg), 3000"

P = Potency of reference standard (%), 99.98%"

$$\% Assay = \frac{AT}{AS} \times \frac{W1}{100} \times \frac{1}{25} \times \frac{100}{W2} \times \frac{25}{1} \times \frac{AW}{LC} \times P$$

Sample Control (Midostaurin)

% Assay =
$$\frac{26139}{28358} \times \frac{04.15}{100} \times \frac{1}{25} \times \frac{100}{04.25} \times \frac{25}{1} \times \text{Error!} \times 99.98$$

= 91.69%

Acidic Degradation (Midostaurin)

% Assay =
$$\frac{24721}{28358} \times \frac{04.15}{100} \times \frac{1}{25} \times \frac{100}{04.25} \times \frac{25}{1} \times \text{Error!} \times 99.98$$

= 86.73%

Basic Degradation (Midostaurin)

% Assay =
$$\frac{23581}{28358} \times \frac{04.15}{100} \times \frac{1}{25} \times \frac{100}{04.25} \times \frac{25}{1} \times \text{Error!} \times 99.98$$

= 82.72%

Oxidative Degradation (Midostaurin)

% Assay =
$$\frac{24357}{28358} \times \frac{04.15}{100} \times \frac{1}{25} \times \frac{100}{04.25} \times \frac{25}{1} \times \text{Error!} \times 99.98$$

= 85.44%

Wet Heat (Midostaurin)

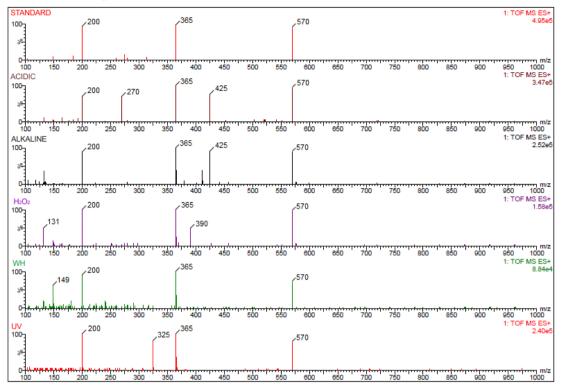
% Assay =
$$\frac{25832}{28358} \times \frac{04.15}{100} \times \frac{1}{25} \times \frac{100}{04.25} \times \frac{25}{1} \times \text{Error!} \times 99.98$$

= 90.62%

Elemental compositions of Midostaurin in MS/MS spectra

| Analyte | Observed ion mass (Da) | Proposed formula | Calculated mass (Da) | Error (ppm) |
|-------------------|------------------------|---------------------------|----------------------|-------------|
| Unknown | 131.38 | $C_7H_{16}NO$ | 130.21 | 1.17 |
| Unknown | 149.54 | $C_8H_{18}NO$ | 144.23 | 5.31 |
| Unknown | 200.02 | $C_8H_9NO_5$ | 199.16 | 0.86 |
| Unknown | 325.42 | $C_{14}H_{19}N_3O_4S$ | 325.38 | 0.04 |
| Unknown | 365.46 | $C_{16}H_{19}N_3O_5S$ | 365.40 | 0.06 |
| Unknown | 390.36 | $C_{17}H_{16}N_3O_6S$ | 390.39 | -0.03 |
| Penicilloic Acids | 425.72 | $C_{16}H_{20}N_3O_6SNa_2$ | 428.39 | -2.67 |
| Midostaurin | 570.60 | $C_{35}H_{30}N_4O_4$ | 570.649 | 0.049 |

Compositions in MS/MS spectra



CONCLUSION

In the pursuit of pharmaceutical quality assurance, this study aimed to develop and validate an analytical technique utilizing Ultra-Performance Liquid Chromatography (UPLC) for the estimation of midostaurin in bulk dosage forms. Midostaurin, a tyrosine kinase inhibitor with significant therapeutic potential in the treatment of certain blood cancers, necessitates precise quantification for both therapeutic efficacy and regulatory compliance.

The UPLC method was meticulously validated, covering parameters such as specificity, system suitability, linearity, accuracy, precision, limit of detection, limit of quantification, and robustness. The robustness studies demonstrated the method's resilience to small variations in chromatographic conditions, affirming its reliability under diverse circumstances.

Forced degradation studies provided insights into the stability of midostaurin under various stress conditions, including acidic and basic degradation, oxidative degradation, and wet heat. The method displayed a remarkable ability to identify and quantify degradation products, allowing for a comprehensive understanding of midostaurin's stability profile.

Assay studies post-degradation indicated the impact of different stress conditions on midostaurin content in bulk dosage forms. While the sample control exhibited a favorable assay result of 91.69%, indicating a stable formulation, variations were observed in acidic

(86.73%), basic (82.72%), oxidative (85.44%), and wet heat (90.62%) degradation, reflecting the sensitivity of the method in detecting changes in midostaurin concentration under stress.

The calculated elemental compositions of midostaurin in MS/MS spectra further supported the accuracy and reliability of the developed UPLC method, with minimal errors observed.

In conclusion, the comprehensive evaluation and validation of the UPLC method for midostaurin estimation in bulk dosage forms have yielded promising results. The method demonstrated robustness, sensitivity, and suitability for routine pharmaceutical analysis. The findings of this study not only contribute to the specific analytical methodology for midostaurin but also underscore the broader importance of validated techniques in pharmaceutical analysis for ensuring the safety, quality, and efficacy of pharmaceutical products.

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