Research Artícle

World Journal of Pharmaceutical and Life Sciences WJPLS

www.wjpls.org

SJIF Impact Factor: 6.129

ISOLATION AND CHARACTERIZATION OF NOVEL ACTINOMYCETES STRAINS FROM THE SOIL OF DARJEELING TEA GARDEN

¹*Swagata Chakraborty and ²Dr. Maitree Bhattacharyya

¹Department of Biotechnology, Techno India University, Salt Lake, EM-4, Sector V, Kolkata- 700091, West Bengal,

India.

²Department of Biochemistry, University of Calcutta, 51/1, Hazra Road, Kolkata- 700019, West Bengal, India.

Corresponding Author: Swagata Chakraborty Department of Biotechnology, Techno India University, Salt Lake, EM-4, Sector V, Kolkata- 700091, West Bengal, India.

Article Received on 20/08/2021

Article Revised on 10/09/2021

Article Accepted on 30/09/2021

ABSTRACT

The organism Actinomycetes was isolated from the soil of the Darjeeling tea garden for carrying out the research. The organism Actinomycetes is a sub-group of Actinobacteria which are Gram- positive and are generally rodshaped or filamentous in nature. They are generally found in soil and also in the urogenital tract, gums and in the digestive system of humans. The bacteria was grown in starch-caesin media followed by the genomic DNA isolation. PCR was carried out for the amplification of the 16s rRNA gene which is unique in almost all bacterial communities and thus acts as a historical or chronological clock, followed by agarose gel electrophoresis. For bacterial identification Biochemical tests were carried out. The actinomycete strains were further screened for the production of secondary metabolites by cross-streak. Furthur purification was carried out and sequencing was done which was then matched with nucleotide sequences from a number of databases.

KEYWORDS: Actinomycetes, gram positive- bacteria, PCR, 16s r RNA, Biochemical tests, secondary metabolites.

INTRODUCTION

The Darjeeling tea garden is a home to variety of soil micro-organisms. Darjeeling tea is mainly grown in the Darjeeling district and the Kalimpong district. Unlike most Indian trees, Darjeeling tea is normally made from the small-leaved Chinese variety of Camellia sinensis. Traditionally Darjeeling tea is black, however Darjeeling oolong and green tea are becoming much more easier to find or produce. However the Darjeeling estates are producing white tea as well. Geography- Darjeeling is located in 27.0360 $\ensuremath{\mathbb{N}}$ and 88.2627 $\ensuremath{\mathbb{E}}$

Soils of Darjeeling

The soils in the Kalimpong area are predominantly red in colour. All the soils are acidic in nature with the tendency to slightly increase slightly in depth, in most cases indicating the lacking of bases from surface and accumulation in lower horizons. The basic soil types are yellow, red brown and deep brown forest soil.



Figure 1: Shows Darjeeling soil.

Actinomycetes

Actinomycetes are Gram-positive bacteria having a complex cell wall(containing muramic acid) that makes Gram staining unsuitable. They have prokaryotic nuclei and are susceptible to antibiotics. They generally form ring or chain like structures which are viewed under microscope.

This type of bacteria is mostly found in soil or in decomposing matter and they form symbiotic nitrogen

fixing associations with almost 200 species of plants. Apart from plants they can also be found on the human urogenital tract as well as the digestive system. Compared to the DNA of the organisms, the actinomycetes have a high percentage of guaninecytosine bases. In growth, this genus generally resembles the fungi but they are much smaller than the fungi. They are highly beneficial in human medicine, as half of the antibiotics such as chloromycetin, kanamycin, streptomycin etc came from soil actinomycetes.

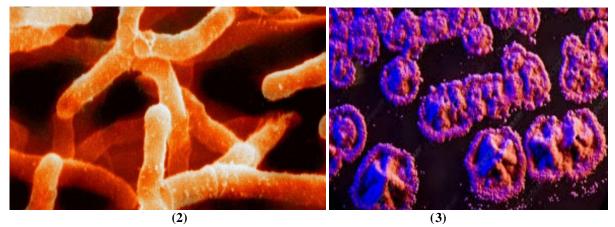


Figure 2 and 3 shows actinomycetes bacteria.

16s rRNA

16s r RNA or 16s ribosomal RNA is the component of the 30S small subunit of a Prokaryotic ribosome that binds to the Shine-Dalgarno sequence. The genes coding for it are referred to as 16s RNA gene. The 16s rRNA genes are ubiquitous i.e the ribosomes cannot translate mRNA without the 16s RNA component. The conserved regions allows the primer to be designed to target the bacteria, but they can amplify the 16s gene through a variable region in which differences in the sequence of the bases allow for the determination of various species. 16s rRNA genes are important because the phylogenetic trees are well developed and taxonomic information is readily

AIMS AND OBJECTIVES

- 1. To determine the uniqueness of the actinomycetes species
- 2. To determine the type of species
- 3. To determine the size of the DNA
- 4. 16S RNA sequencing for identification of the species
- 5. Biochemical Test to differentiate the bacterial type for determination of secondary metabolite synthesis
- 6. How anthropogenic influence shapes the characteristic of the bacteria
- 7. Characterization
- 8. Bioactive molecule screening

Literature Review

1-Definition

Actinomycetes- They are a order of actinobacteria which are Gram positive bacteria and generally aerobic in

nature which can be mainly found in soil

Ref- Actinomycetales- List of prokaryotic names with standing in nomenclature

Origin

The word actinomycetes were derived from a Greek word mukes or muketes. Actinomycete is a modern latin word.

Ref- Wikipedia

2-Definition

16s r RNA gene- 16s rRNA is the component of the 30S small subunit of a prokaryotic ribosome that binds to Shine-Delgarno sequence and the genes coding for it are referred to as 16s rRNA genes. Ref- Wikipedia

Origin

The large ribosomal subunit and the peptidyl transferase centre reconstructed the ancestral sequences of the tRNA's and they showed similarity between concatamers of proto-tRNAs with modern PTC.

Ref- Article in origin of the 16s Ribosomal molecule from ancestor tRNAs.

The gram positive bacteria actinomycetes are slow growing filamentous bacteria that are mainly found in the soil. Apart from this they are also found in the human's urogenital tract and their digestive tract. 16s rRNA genes are mainly found in all the bacterial species. Thus this gene acts as an evolutionary clock, the sequencing of which will aid in the identification of the organism. The 16s rRNA PCR and sequencing was done.

Ref- Identification by 16s rRNA Gene sequencing of Actinomycetes; NCBI

Analysis of Actinomycete communities by specific amplification of genes encoding 16s rRNA and gel electrophoretic separation in denaturing gradient; Heuner, Krsek, Baker, Smalla, Wellington; American society for Microbiology journal.

However, microbial pathogens are becoming increasingly resistant to available treatment and thus new antibiotics are needed, against antibiotic resistant strains of the organisms. The Actinomycetes provide a variety of antibiotics which are diverse in nature. The organism is needed to produce a variety of bioactive compounds and the 16s rRNA gene sequencing is unique to the bacterial community.

Ref- Diversity and Versaltility and its role in antibiotic production; Journal of applied pharmaceutical science.

MATERIALS AND METHODS

Isolation

Cultivable strains of bacteria were isolated using initial screening in normal saline solutio0.9%). Serial dilution was done(10 fold dilution) and the saline suspensions were spread spread on casein-starch medium containing starch, casein, potassium nitrate, potassium dihydrogen phosphate, sodium chloride, magnesium sulphate, calcium carbonate, ferrous sulphate with suitable antibiotics. The plates were incubated at 28 $^{\circ}$ for 5-7 days. Single colonies were selected and further streaked on casein-starch medium. The process was repeated 5-6 times to facilitate enrichment of pure cultures.

After isolation starch-caesin media was prepared again for the growth of the bacteria. The growth of the bacteria was determined in two ways: 1) growth in plate and 2) growth in broth.

Media preparation and components for plates

Starch-10gm Caesin-1gm KNO3-2gm KH2PO4-2gm NaCl-2gm MgSO4-0.5gm CaCO3-0.02gm FeSO4.7H2O-0.001 Agar-18gm

The pH was checked and maintained between 7.0-7.4(calibrated with 2M NaOH)

Pour plate method

The media was autoclaved and it was carefully poured into the plates. They were kept overnight to solidify.

Quadrant plate streaking

The rules for quadrant streaking was followed and the plates were streaked for isolating single bacterial colonies. The plates were kept at 28 °C for 5-7 days as this bacterial species is slow growing in nature.

Media preparation and components for broth Starch-10gm Caesin-1gm KNO3-2gm KH2PO4-2gm NaCl-2gm MgSO4-0.5gm CaCO3-0.02gm FeSO4.7H2O-0.001 pH was measured and kept between 7.0-7.4(calibrated with 2M NaOH)

The agar plates on which the bacteria has grown for several days are taken out. The broth media and toothpicks are autoclaved. The work was carried out in a laminar air flow chamber. Sterile toothpicks were used to pick up the bacterial colonies from the plate and put in the broth. After the procedure was over, the broth tubes were kept in 28 °C for the next 5-7 days. The clear solution turned turbid or yellowish which confirmed the growth of the bacteria.

Genomic DNA Isolation

2ml culture was taken and centrifuged at 14,000 rpm to collect the pellet. The culture media was removed and discarded

It was then resuspended into 300µl lysis buffer.

2µl RNAase solution was added and incubated at room temperature for 2mins. This helps in removing the RNA from the mixture.

20µl Proteinase K solution was added which helps in inhibiting the protein. The solution was mixed and the resuspended cells were transferred to the HiBead Tube and incubated for 30mins at 55 °C.

The HiBead Tubes were horizontally secured on a flat bed vortex pad with tape. It was then vortexed at maximum speed for 5-7 mins.

The mixture was then incubated for 10 mins at 95 \degree followed by pulse vortexing.

The tubes were centrifuged at 13,000rpm for 1min at room temperature.

The supernatant was then transferred to clean capped 2.0 ml collection tube.

200µl lysis buffer was added and vortexed thoroughly (15secs), followed by incubation for 10mins at 55 $^{\circ}$ C.

200µl ethanol was added to the lysate and mixed

thoroughly by vortexing for 15 secs.

The lysate was the loaded in HiEulute Mini prep spin column (capped) and centrifuged at 10,000rpm for 1min. The flow through was discarded and the spin column was placed in the same 2.0ml collection tube.

Prewash: 500µl of prewash solution was added to the column and centrifuged at 10,00rpm for 1min. The flow through was was discarded and thee same collection tube with the column was reused.

Wash: 500μ l of wash buffer was added to the column and centrifuged for 3mins at 14,000rpm. The flow through was discarded the same collection tube was used.

Additional 1 min of centrifugation was done for presence of ethanol in column.

DNA Elution: The flow through was discarded and the column was transferred to an uncapped collection tube. At first $70\mu l$ of Elution buffer was added directly to the

centre of the column and incubated at room temperature for 1 min. It was then centrifuged at 10,00rpm for 1 min. Now 30μ l Elution buffer was added and the same procedure was followed as above. The DNA was stored at -20 °C.

16S r DNA PCR

Primer designing was done to determine the suitable primers PCR reagents and amounts Water -16.25μ l Buffer -2.5μ l Forward primer -1μ l Reverse primer -1μ l dNTP -1μ l Taq pol -0.25μ l Template -3μ l Total- 25μ L

PCR conditions

The V1-V9 region of the 16s rRNA gene was amplified using the standard PCR protocol. The PCR conditions are mentioned below;

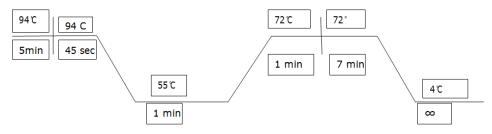


Figure 4: shows the PCR conditions.

Agarose gel electrophoresis

This is the most unique way of determining the size of DNA. The phosphate in the DNA is negatively charged so it will run towards the positive pole. The band was obtained in 1000bp range as compared with the DNA ladder.

PCR Purification

The protocol for PCR purification was followed and the products were purified for sequencing. The Qiagen gel extraction was used for purification.

Library Preparation

The amplicon library was purified and pooled amplicons were diluted as recommended and amplified by emulsion PCR on a thermal cycler.

Biochemical Test

Gram Staining

The Gram stain developed in 1884 by Danish physician Christian Gram is the most widely employed staining method in bacteriology. It is an example of differential staining because the use of Gram stain divides the bacteria into two classes- Gram positive and Gram

negative.

The procedure is as follows

After heat fixation, Smear is stained with basic dye crystal violet, the primary stain

Treatment with iodine solution functioning as a mordant. The iodine increases the interaction between the cell and the dye so that the cell is stained more strongly

Smear is decolourised by washing with ethanol or acetone, this step generated differential aspect of Gram staining, the gram-positive bacteria wil retain the crystal violet dye whereas the gram-negative bacteria will become colourless.

Finally the smear is stained with a counter stain called safranin. The gram negative bacteria as a result becomes pink as it retains the safranin stain.

Secondary Metabolites

Screening of microbial secondary metabolites is an important method to identify biologically active compounds. The procedure is as follows;

At first media was prepared (LB Agar broth) and autoclaved.

Then the media was poured into the plates and kept to solidify in the LAF for about 1 hour.

Strains of Actinomycetes were taken and streaked into the LB agar plates in a perpendicular manner and incubated at 28 °C.

Ater 7 days the growth was checked and it was streaked

RESULTS AND DISCUSSION

Quadrant Streak Plates

with other standard strains for determining the activity of secondary metabolites.

The secondary metabolites spreads into the middle of the plate to a particular distance. So upto that distance the other micro-organisms cannot grow which means that with respect to that organism, these secondary metabolites are potent antibiotics which inhibits their growth. So it can be said that the secondary metabolite synthesized from this compound is a potential source of new antibiotic.



Figure 5: Shows the first three strains used for quadrant streaking.

Quadrant Streak Plates



Figure 6: Shows the second part of the three strains used for quadrant streaking.

Gram-positive bacteria under microscope

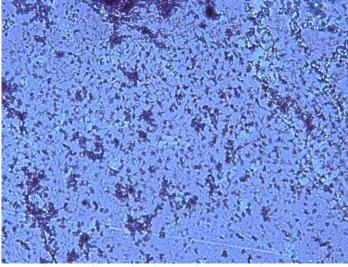


Figure 7: Shows Strain AB737 under microscope.

Gram positive bacteria under microscope

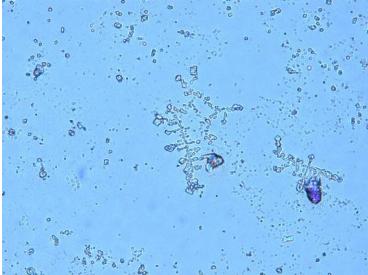


Figure 8: Represents Strain AB757 under microscope.

Gram positive bacteria under microscope

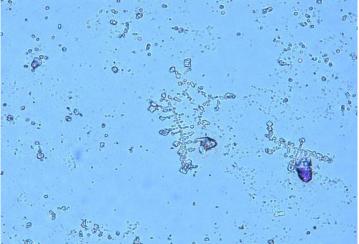


Figure 9: Represents Strain AB757 under microscope.

Gram-positive bacteria under microscope

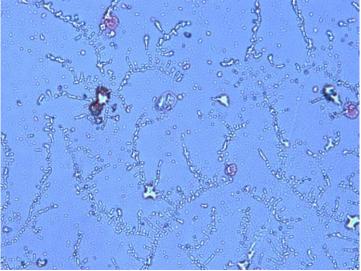
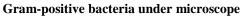


Figure 10: Shows Strain AB SCA5 under microscope.



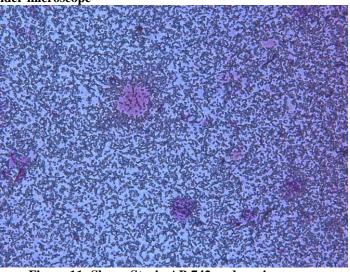


Figure 11: Shows Strain AB 742 under microscope.

Gram- positive bacteria under microscope

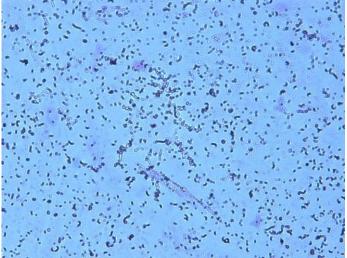


Figure 12: Shows Strain AB 847 under microscope.

Cross-streak for determining the activity of secondary metabolites



Figure 13: Shows the activity of the secondary metabolites. The mother strain streaked perpendicularly is the actinomycetes and the rest of the strains are as follows 1-E.coli, 2-B.subtilis, 3-P.aeruginosa 4-S.typhi 5-V.cholerae 6-E.aerogenes 7-S.aureus 8-S.typhimurium, 9-B.hemolyticus.

CONCLUSION

Actinomycetes are a group of gram positive bacteria which is hugely used in research due to its beneficial effect. These bacteria are a source of about 90% of potent antibiotics. Generally this group of bacteria uses variety of substrates found in soil and that is what makes it an important bacteria to study about. They are potent bio fertilizers and apart from being nitrogen fixing bacteria they produce geosmin which imparts earthy odor to soil.

In this project we mainly studied how actinomycetes behave as a potent source of antibiotics and how unique 16s r RNA genes are. These genes are ubiquitous i.e the cannot translate mRNA without their 16s RNA. This study clearly states how 16s r RNA acts as a signature sequence unique for a particular species thus giving us insight about the evolution of the organism through time.

Another main point of focus that the study iterates is the most challenging genre of research: antibiotic resistant. We used actinomycetes as the mother strain and streaked it against nine different strains to check the growth of potent secondary metabolite which will be a potent antibiotic with respect to that organism and thus we detected new strains which will be beneficial in the genre of antibiotic resistance.

More studies are done and are required because antibiotic resistance is a potential threat to mankind because there can be days when there will be no potent antibiotic to treat infections. In addition to this study, further studies are required on actinomycetes and antimicrobial resistance to further draw conclusion for antibiotic resistance which will most likely be another pandemic in near future and also for various reasons like bioremediation and natural resources.

ACKNOWLEDGEMENTS

I am honoured to have the privilege of rendering my profound gratitude and greatest obligation to my guide, Dr.Maitree Bhattacharyya, Professor, Department of Biochemistry, University of Calcutta and Director of Jagadish bose national science talent search; for giving me the opportunity to work in her lab.

I express my profound gratitude to Dr.Anish Bhattacharyya, SRF, Department of Biochemistry, University of Calcutta, for his advice, encouragement and continuous help, without which it would not have been possible to complete this project

I owe my wholehearted thanks and appreciation to all tha lab members for their guidance.

Last but not the least, I would like to thank my parents for their continuous support.

BIBLIOGRAPHY

- Ghosh A, Dey N, Bera A et al. "Culture independent molecular analysis of bacterial communities, 2010 Feb 17; 6.
- 2. Bhattacharyya A, Halder A, Bhattacharyya M, Ghosh A," Anthroprogenic influence shapes the distribution of antibiotic resistant bacteria in the sediment of Sundarban estuary in India, Science of total environment, 2019; 647: 1626-1639.
- Aminov RI." The role of antibiotics and antibiotic resistance in nature. Environ Microbiol, 2009 Dec; 11(12).
- 4. Dwivedi S, Thakkar R, Upadhay V, Kumar S," Isolation and characterization of Actinomycetes producing Antimicrobial substance against human pathogenic bacteria, Journal of pharmacy research, 2011.
- 5. Seikh G, Mab\qbul M, Shankar S," Isolation and

characterization of actinomycetes from soil of addawadmi, Saudi Arabia and screening their antibacterial activities.

- 6. LL.T.A.S Semedo, R.R.R Coelho," Isolation and characterization of actinomycetes from Brazilian tropical soils"
- 7. Jeffery, L.S.H, "Isolation and characterization and identification of actinomycetes from agricultural soils at Semongok, Sarawak.
- 8. Muthu M, Balasubramanium, "Isolation and Identification of Actinomycetes Isoptericola variabilis from Cauvery river soil sample.
- 9. X.T Jiang, X. Peng, G.-H. Deng et al., "Illumina sequencing og 16s rRNA Tag revealed spatial variations of Bacterial communities in a mangrove wetland" Microbial Ecology, 2013; 66: 96-104.
- 10. C. A Black, Methods of soil analysis, American Society of Agronomy, Madison, Wis, USA, 1965.
- 11. D. W. Nelson and L.E Somers, Organic Carbon, Academic Press, London, UL, 1975.
- 12. M. Knudsen, Hydrographical Tables, CEG GAD, Copenhagen, Denmark, 1901.