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ABSTRACT

Microbial activity causes serious damage to living organisms. In this study, ethanolic extracts were prepared from leaves of *Persea americana* Mill. *Pimenta dioica* (L.) Merr., *Cinnamomum tamala* (Buch.-Ham.) T. Nees & Eberm. and *Cinnamomum verum* J. Presl. The ethanolic extracts prepared from these plants exhibited broad spectrum antibacterial activity when evaluated against gram-positive bacteria *Streptococci* spp. and *Staphylococci aureus* and gram-negative *Escherichia coli* and *Pseudomonas aeruginosa* by disc diffusion method and growth inhibition assay.

KEYWORDS: Staphylococcus aureus, Persea americana, Pimenta dioica, Pseudomonas aeruginosa and antimicrobial.

INTRODUCTION

Plants have become the center of interest for the cure of various diseases in recent past. The utilization of plants for treatment of any disease has been tremendously increased from the last few years, because of the wider belief of less adverse effects. The risk of infection is increasing progressively, due to growing environmental pollution and lifestyle. This also leads to mutation and resistance bacteria. Whereas, synthetic drug antimicrobial agents that are used are also having many side effects and cause harm to the human body. Therefore, several natural products based antimicrobials have emerged to tackle this problem.

Spices are flavored or aromatic substances originating from vegetables and are obtained from the topical plants and are used to enhance the taste of the food and as condiments. The spices contain enormous amount of volatile oil which gives a general notion of the antimicrobial potential of various spices. This made the spices to utilize as adjuvant as antimicrobial agents for controlling the risk of infection. In the present study, four different plant leaves were collected and investigated for anti-microbial activity. The plant details such as botanical names, family, common name and parts used in the study are given in table 1.

Botanical Name	Family	Common name	Part used
Persea americana Mill., & Eberm	Lauraceae	Avocado	Leaf
Pimenta dioica (L.) Merr.	Myrtaceae	Allspice	Leaf
Cinnamomum tamala (BuchHam.) T.Nees	Lauraceae	Bay leaf, tejpatta	Leaf
Cinnamomum verum J. Presl	Lauraceae	Cinnamon, dalchini	Leaf

Table 1: Plants Name and its description.

Persea americana (Avocado)

Avocado belonging to the family of Lauraceae and its botanical name is *Persea americana* is a tropical tree crop originating from tropics of the western hemisphere. The avocado originates in highlands of the Guatemalan, and central and east-central Mexico highlands. The height of the Avocado tree reaches 30 m (Guatemalan and West Indian species) and the diameter of the trunk between 30 - 60 cm. The leaves of the plant are darkgreenish in color with a glossy layer on the upper surface, lanceolate, elliptic, ovate, and are alternatively arranged on the stem. The plant contains fruits of pearshaped and the greenish yellow pulp of fruit is edible.^[1] The largest producer of avocado is Mexico with about 25% of the world production.^[2] The traditional system of medicine reported good medicinal properties of Avocado which include antiulcer effect, anticonvulsant activity, antihepatotoxic activity, hypoglycemic activity and antioxidant activity.^[3]

Pimenta dioica (Allspice)

Allspice scientifically known as Pimenta dioica is a member of family Myrtaceae. It is an evergreen, tall and bushy tree of height 6.6 - 10 m. It is more branched at the top and the color of the bark is greyish. Leaves are oppositely arranged, oblong, lanceolate, tapering toward petioles and the size of leaves varies to 11-11.5 cm X 5 -6 cm and it contains essential oils.^[4] Allspice is indigenous to West Indies and Central America. The essential oils of the plant are like that of clove, nutmeg, cinnamon, and black pepper, so the effect produce by allspice is somewhat similar to these. It contains bilocular berries which are hard, brown-reddish having a diameter of 4 - 7 mm. Berries have a characteristic odor because of the essential oils present in it. Allspice contains phytoconstituents like alkaloids, glycosides, flavonoids, amino acid, saponins, tannins, etc. The plant acts as a very strong bacteriocide because of the presence of spice components which are thymol, eugenol, menthol and anethole. It also acts as an insecticide and antioxidant and is used as a food preservative. Other effects of allspice are the same as that of clove and other spice. It is also used as a digestant and stomachic.^[5]

Cinnamomum tamala (Tejpatta)

Tejpatta botanically known as Cinnamomum tamala of family Lauraceae is a small or perennial evergreen tree of height 8 - 12 m, the stem is rough with a girth of 150 cm and produces mucilage. Leaves are long (about 12 -20 cm) and are 5 - 8 cm broad. Its shape is somewhat ovate-lanceolate, thick, leathery, shining green on top and is oppositely arranged. The flower of the plant blossoms at the end of March or at the starting of April. Its leaves are vastly used as a spice for enhancing the flavor of food. The oil of tejpatta obtained from leaves is used as a diuretic, carminative and in cardiac disorders. It has been used as a food preservative for pineapple juice. Its special aroma makes it useful in the food industry. The main constituents of C. tamala are camphene, pinene, limonene and eugenol. Traditionally, tejpatta leaves are used in bladder disease, diarrhea, mouth dryness and nausea. It also shows hypoglycemic and hypolipidemic activity.^[6]

Cinnamomum verum (Dalchini)

The botanical name of dalchini is *Cinnamomum verum* and to the family of Lauraceae. It is widely found in South India and Sri Lanka. It is the oldest tree spice that was used by mankind. It is an evergreen, medium-sized tree of height about 16 m. Its bark has a delicately fragrant aroma with warm and sweet flavor along with pungent taste. Cinnamon leaves are shiny green on the

upper side and have a leathery look. The lower side is somewhat dull than the upper. When rubbed in hands, its leaves emit some spicy odor. Leaves are oppositely arranged, oval or elliptic to lanceolate and length of the leaves varies from 8.7 - 22.7 cm. It contains bisexual flowers which are pale yellowish-green in color and flowering occurs in October or November and continues till March ^[7]. Cinnamon or dalchini is the most commonly used spices that enhance food's flavor and is also used in tea as a taste enhancer. The small quills of its bark have a huge market all over the world because of the presence of chemical constituent oleoresin. The dried dalchini leaves are also used for food flavoring. It shows hypoglycemic activity and can be used for treating type 2 diabetes mellitus. The presence of phenolics tcinnamaldehyde, eugenol and cinnamic acid are responsible due to its antioxidant and antimicrobial property, for which it is used as a food preservative. The plant is reported for its antifungal, antibacterial and insecticidal activity.^[8]

MATERIALS AND METHODS

In the current study plant extracts were prepared and evaluated for anti-microbial activity by the disc diffusion method and growth inhibition assay. The plants selected for the study were Persea americana (Avocado), Pimenta dioica (Allspice), Cinnamomum tamala (Tejpatta) and Cinnamomum verum (Dalchini) (Details are given in table 1).

Plant material collection

Fresh leaves of all four plants were collected from the Madan Mohan herbal garden of Jawaharlal Nehru Cancer Hospital and Research Centre, Idgah Hills, Bhopal, India.

Preparation of plant extract

The leaves collected from four plants were cleaned from both sides properly with tissue paper and kept aside for a few minutes to dry at room temperature. The leaves were minced in small pieces using sterilized scissors. The mined leaves were soaked in 40 ml of 50% ethanol (50 ml ethanol and 50 ml distilled water) in a clean beaker for 24 hours. The extracts were centrifuged after 24 hours for 15 minutes at 1000 rpm. The prepared extract was used for further experiments and the rest of the extract stored at 4°C in the refrigerator.

Antimicrobial Activity

The antimicrobial activity was tested in plant extracts by growth inhibition assay and disc diffusion method using different bacterial strains.

Bacterial strains

The bacterial strains tested for the antibacterial activity were isolated in the Department of Research, Jawaharlal Nehru Cancer Hospital and Research Centre. The strains used for the study were two gram-positive bacteria *Streptococci* spp. and *Staphylococci* *aureus,* whereas *Pseudomonas aeruginosa* and *Escherichia coli* were used as gram-negative bacteria.

Disc Diffusion Method

The plant extracts were examined for antimicrobial activity by using disc diffusion method. The discs were taken in different petri plates and loaded with extracts and air-dried and subsequent loading was done with drying. After this, the discs were taken in glass vials, labeled and autoclaved. The discs for negative control were also prepared in ethanol only. The bacterial strains were inoculated on Mueller Hinton plates using sterile stirrer of high-quality stainless steel. After inoculation, the extract discs were placed using sterile forceps. The negative and positive control discs were also put on plates and were incubated at 37°C. After 24 hours of treatment, the plates were observed and the zone of inhibition was calculated. For positive control, standard discs were placed separately for gram negative and gram positive bacteria.^[9]

Growth Inhibition Assay

Nutrient agar media was prepared and poured in plates with 500 μ l of ethanolic extract and mixed using stirrer. The media was allowed to solidify and after solidification bacterial strains were inoculated using swab and incubated at 37°C. The plates were examined after 24 hours of treatment and the number of bacterial colonies was counted.^[10]

RESULTS

In the present study four plants were selected which includes *P. americana*, *P. dioica*, *C. tamala* and *C. verum*. Disc diffusion method and growth inhibition assay was performed to examine antimicrobial potential

of ethanolic extract of all four plants. The phytochemical like flavonoids and tannins present in plants, were responsible for its antimicrobial activity.

Disc Diffusion Method

The ethanolic extracts of plants was loaded on discs and placed on Mueller Hinton plates showed potent antimicrobial activity against all strains used for the study. The strains used for the study were P. aeruginosa, Streptococci spp., S. aureus and E. coli. P. americana (Avacado) showed zone of inhibition of 7 mm, 8 mm and 12 mm against Streptococci spp., P. aeruginosa and S. aureus respectively. P. dioica (Allspice) were found more effective than other plant extracts and gave zone of inhibition of 6 mm, 8 mm, 8 mm and 18 mm against E. coli, P. aeruginosa, Streptococci spp., and S. aureus respectively. The extract (ethanolic) of C. tamala and C. verum was found inactive and no zone of inhibition was found against both the plant extracts. The zone of inhibition obtained in standard disc used for the study for gram positive (Table - 3) and gram negative (Table - 4) bacteria were given in tabular form for comparative study. The results of extracts by disc diffusion method found were also given in Table -2.

Growth Inhibition Assay

The nutrient agar plates were taken along with 500µl of extract and inoculated with bacterial strains. The extracts which inhibited the growth of bacteria showed less or no growth in culture plates (Table -5). The ethanolic extracts of *P. americana*, *P. dioica*, *C. tamala* and *C. verum* leaves show potent antibacterial activity against *P. aeruginosa*, whereas *P. americana* inhibited the growth of *Staphylococcus aureus* bacteria.

Table 2: The zone of inhibition of ethanolic plant extracts against *Streptococci, Staphylococcus aureus, Pseudomonas aeruginosa* and *Escherichia coli*. The diameter of zone of inhibition (mm). No zone of inhibition indicated by NZ.

S. No.	Plant Name	Zone of Inhibition (mm)			
5. 110.	r lant manne	Streptococci	Pseudomonas aeruginosa	Staphylococcus aureus	Escherichia coli
1.	Avocado	7 mm	8 mm	12 mm	NZ
2.	Allspice	8 mm	8 mm	18 mm	6 mm
3.	Tejpat	NZ	NZ	NZ	NZ
4.	Dalchini	NZ	NZ	NZ	NZ

Table 3: Standard Drug for Gram positive bacteria.

N	Drug Name	Zone of Ir	Zone of Inhibition (mm)			
No.		Streptococci spp.	Staphylococci aureus			
1.	Cefotaxine	NZ	-			
2.	Ceftriaxome	NZ	-			
3.	Ceftazidine	NZ	-			
4.	Nitrofurantoin	NZ	-			
5.	Gentamicin	NZ	-			
6.	Amikacin	NZ	-			
7.	Cefuroxime	20 mm	-			
8.	Cefixime	NZ	-			
9.	Nalidixic acid	NZ	-			

10.	Cefdinir	NZ	-
11.	Cephalexin	18 mm	-
12.	Tetracyclin	10 mm	-
13.	Piperacillin	30 mm	-
14.	Pinicillin G	16 mm	-
15.	Cefazolin	16 mm	-
16.	Chloramphenicol	14 mm	-
17.	Ciprofloxacin	7 mm	-
18.	Erythromycin	13 mm	-
19.	Azithromycin	12 mm	-
20.	Cotrimoxazol	18 mm	-
21.	Amoxicillin clavulanic acid	25 mm	-
22.	Amoxicillin	25 mm	-

Table 4: Standard Drug for Gram negative bacteria.

C No	Drug Name	Zone of Inhibition (mm)			
S. No.		Escherichia coli	Pseudomonas aeruginosa		
1.	Cefotaxine	-	23 mm		
2.	Ceftriaxome	-	20 mm		
3.	Ceftazidine	-	12 mm		
4.	Nitrofurantoin	-	18 mm		
5.	Gentamicin	-	12 mm		
6.	Amikacin	-	10 mm		
7.	Cefuroxime	-	18 mm		
8.	Cefixime	-	8 mm		
9.	Nalidixic acid	-	8 mm		
10.	Cefdinir	-	20 mm		
11.	Cephalexin	-	-		
12.	Tetracyclin	-	-		
13.	Piperacillin	-	-		
14.	Pinicillin G	-	-		
15.	Cefazolin	-	-		
16.	Chloramphenicol	-	-		
17.	Ciprofloxacin	-	-		
18.	Erythromycin	-	-		
19.	Azithromycin	-	-		
20.	Cotrimoxazol	-	-		
21.	Amoxicillin clavulanic acid	-	-		
22.	Amoxicillin	-	-		

 Table 5: Growth inhibition Assay of ethanolic plant extracts against Streptococci spp., Staphylococcus aureus,

 Pseudomonas aeruginosa and Escherichia coli.

S.	Plant Name	Bacterial Strains				
No.		Streptococci	Pseudomonas aeruginosa	Staphylococcus aureus	Escherichia coli	Streptococci
1	Avocado	Moderate growth of bacteria	No growth	No growth	Inhibition not present	Inhibition not present
2	Allspice	Inhibition not present	No growth	Inhibition not present	Inhibition not present	Inhibition not present
3	Tejpat	Moderate growth of bacteria	No growth	Inhibition not present	Inhibition not present	Inhibition not present
4	Dalchini	Inhibition not present	No growth	Moderate growth of bacteria	Inhibition not present	Inhibition not present



Figure 1: Plants common and botanical name. (A) *Persea americana* (Avocado), (B) *Pimenta dioica* (Allspice), (C) *Cinnamomum tamala* (Tejpatta) and (D) *Cinnamomum verum* (Dalchini).

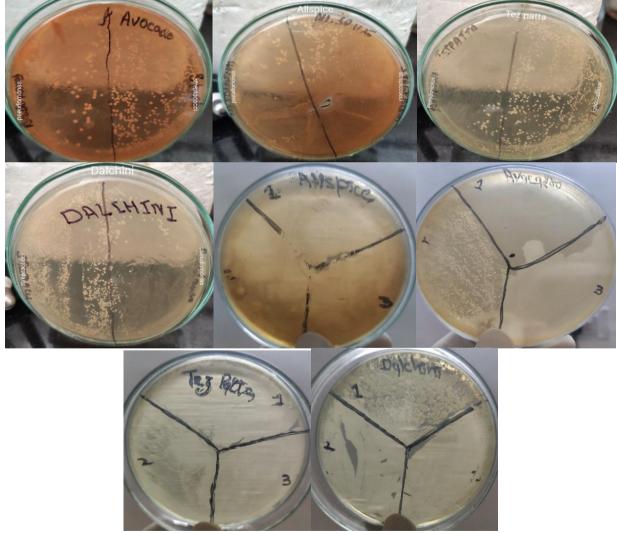


Figure 2: Growth inhibition of different plant extracts against *Escherichia coli* and *Staphylococcus aureus* (A) *Persea americana* (Avocado), (B) *Pimenta dioica* (Allspice), (C) *Cinnamomum tamala* (Tejpatta) and (D) *Cinnamomum verum* (Dalchini).

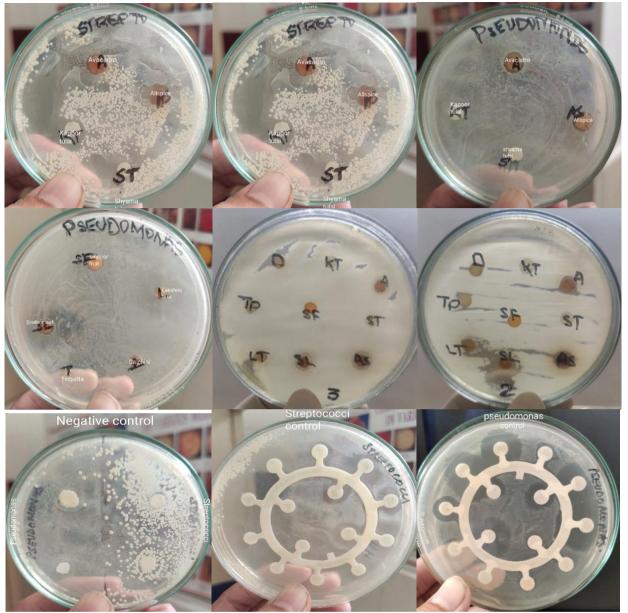


Figure 3: Zone of inhibition (mm) of different plant extracts against *Pseudomonas aeroginosa* and *Streptococcus* spp. by disc diffusion method.

DISCUSSION

Several researchers had analyzed antimicrobial activity on Persea americana, Pimenta dioica, Cinnamomum tamala and Cinnamomum verum. Our work includes the ethanolic extract of plants of above plants. In our study P. *americana* (Avocado) showed potent antimicrobial activity with a zone of inhibition of 7 mm, 8 mm and 12 mm against Streptococci spp., P. aeruginosa, and S. aureus respectively. The plant extract also inhibited the growth of bacteria in growth inhibition assay. Whereas, other researchers showed that methanolic extract of leaf gives highest zone of inhibition of 6.0 mm and for bark it gives the maximum zone of inhibition 12.0 mm against S. *aureus* using agar well diffusion method.^[11]

Most of the scientists had worked on fruit extract of P. americana (Avocado) and showed its antimicrobial activity. In a study, organic extracts prepared from seeds of two avocado species showed adequate antibacterial activity against gram positive S. aureus showing a zone of inhibition 19.03 mm by disc diffusion method.^[12] Another study on antibacterial activity of four varieties of avocado fruits was done by Amado and co-workers (2019) using E. coli, S. aureus, Salmonella and B. cereus. They found that peel and seed show less antimicrobial activity in gram negative bacteria than that of gram positive bacteria.^[13] The plant also showed antibacterial activity against a strain Porphyromonas gingivalis, isolated from primary endodontic infection with minimum bactericidal concentration (MBC) value at a concentration of 60% and MIC value at a concentration of 50%.^[14] The methanolic extracts of P.

americana showed potent antimicrobial activity against two MDR strains of *M. tuberculosis* H37Rvand H37Ra with MIC value of 62.5 μ g/ml and 125 μ g/ml respectively ^[15]. Another study showed antibacterial activity in methanolic extracts were more effective than acetone extract against clinical isolates which includes *Pseudomonas aeruginosa*, *Bacillus cereus*, *Salmonella typhi, Bacillus subtilis, Shigella flexneri, Escherichia coli* and *Staphylococci aureus*.^[16]

The aqueous and ethanolic seed extracts of P. americana suppressed the growth of gram-positive and gram-negative bacteria. The aqueous extracts have activity against Staphylococcus epidermidis and Listeria monocytogenes with MIC values of 354.2µg/mL and 93.8 - 375.0 μg/mL respectively.^[17] The antimicrobial potential of extracts of P. dioica (Allspice) were reported by several researchers and our study showed potent effect against P. aeruginosa, E. coli, Streptococci spp. and S. aureus with a zone of inhibition of 8 mm, 6 mm, 8 mm and 18 mm respectively. One study showed ethanolic leaves extract of P. dioica showed potent antibacterial activity against different bacterial strains in the order of *Staphylococcus aureus* < *Salmonella* typhimurium < Bacillus cereus < Escherichia coli.^[18] Whereas, some researchers gave significant activity in hexanes and alcoholic extracts with 32.1 ± 0.26 mm and 20.3 ± 0.16 mm zone of inhibition against grampositive B. megaterium and gram negative P. fluorescens respectively by disc diffusion method.^[19] The methanolic extract of leaf and bark of Allspice was found highly effective against S. mutans than S. aureus recovered from burn and dental caries patients^[20] Hari and coworkers (2010) studied antibacterial activity in essential oil and aqueous extract of P. dioica for against five different microbial species (including both gram negative and gram positive strains).^[21] The essential oils of leaves of P. dioica also exhibited potent antimicrobial activity against *B. cereus, S. typhimurium* and *S. aureus* by disc diffusion method.^[22] In another study MBC and MTC (maximally tolerated concentration) values of essential oil of P. dioica was obtained against P. putida was found to be 0.1% wt/vol and 0.05% wt/vol respectively.^[23]

Cinnamomum tamala is known for its medicinal property and used as a spice in India. The presence of major phytochemical constituents detected in methanolic extracts of C. tamala were tannins, alkaloids, flavonoids, terpenoids which were responsible for its and antibacterial activity ^[24]. Another study showed acetone, methanol and ethanolic extract of leaves of Tejpatta showed inhibitory action against both *Bacillus* cereus and Serratia. The zone of inhibition of acetone extract against *B*. cereus was found 26.6 ± 0.57 mm by agar well diffusion method.^[25] Whereas, the methanolic leaf extract of C. tamala showed activity in the sequence of P. aeruginosa < K. pneumonia < E. coli < S. aureus.^[26] In another study, methanolic extract of leaves of C. tamala exhibited potent antimicrobial activity against clinical samples of 8 different multidrug-resistant Gram negative bacteria namely Pseudomonas spp, K. pneumonia, Citrobacter spp., P. vulgaris, K. oxytoca, E. *coli*, Acinetobacter spp. and P. mirabilis $[^{\tilde{2}7]}$. Even essential oil taken from dried and fresh leaves of C. tamala showed potent antibacterial activity against M. luteus, B. subtilis, S. aureus, and P. aeruginosa^[28]. In present study, the ethanolic extracts of C. tamala and C. verum showed the growth inhibition of P. aeruginosa by growth inhibition assay whereas, both the extracts were found inactive when evaluated by disc diffusion method. Whereas other extracts showed moderate action against all the strains used for the study. The methanolic extracts of leaves and bark were tested against P. aeruginosa, S. aureus, E. coli, B. cereus by agar well diffusion method and agar dilution method and was found to be potential sources of antimicrobial agents ^[29]. The ethanolic extract of cinnamon bark exhibited an antibacterial effect against K. pneumonia and E. coli with zone of inhibition 25.50 ± 3.72 mm and 11.72 ± 1.86 mm respectively.^[30] In another study ethanolic extract and essential oil of C. verum were studied by Hamedo (2015) found it effective against E. coli and P. aeruginosa ^[31]. Cinnamaldehyde and eugenol present in the essential oil of C. verum are responsible for the bactericidal activity and found effective against Paenibacillus larvae and gave MBC and MIC of $125-250\mu$ g/ml and $25-100\mu$ g/ml respectively by tube dilution method.^[32] The ethanolic extracts of cinnamon bark were found potent against four bacterial strains aureus, E. coli, S. namely S. mutans, and Peptococcus isolated from orofacial infections [33]. The methanolic extract and its oil of C. verum also found to have a bacteriostatic action against E. coli, P. aeruginosa, and B. subtilis bacteria found in ketchup.^[34]

CONCLUSION

The ethanolic extracts prepared from all the four plants showed potent antimicrobial activity against two gram negative and gram positive bacteria. The extract of *Cinnamomum tamala* showed the highest effect as compared to other extracts. The plant can be further studied and investigated. The investigation and research on medicinal plants might bring to the scientific world many useful remedies for the treatment and cure of human sufferings.

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REFERENCES

- 1. Chanderbali AS, Soltis DE, Soltis PS, and Wolstenholme BN. Taxonomy and Botany. In Schaffer C, Wolstenholme BN, Whiley AW (eds.). The Avocado: Botany, Production and Uses, 2nd ed., Wallingford; CABI: 2013; 31-50.
- Gutiérrez-Díez A, Sánchez-González EI, Torres-Castillo JA, Cerda-Hurtado IM, and Ojeda-Zacarías MDC. Genetic Diversity of Mexican Avocado in Nuevo Leon, Mexico. In Caliskan M (eds.). Molecular Approaches to Genetic Diversity, 1st ed., London; InTechOpen, 2015; 140-159.
- 3. Yasir M, Das S, and Kharya M. The phytochemical and pharmacological profile of *Persea americana* Mill. Pharmacogn. Rev., 2010; 4(7): 77–84.
- Rao PS, Sheth NR, Jayaveera KN, and Rao SK. Pharmacognostic standardisation of the leaves of *Pimenta dioica* Linn. Int. J. Pharm. Sci. Res., 2010; 1(9): 110–115.
- Bagchi GD and Srivastava GN (2003). Spices and flavoring (flavouring) crops: Fruits and Seeds. In Caballero B, Finglas P, Toldra F (eds.). Encyclopedia of Food Sciences and Nutrition, 2nd ed., Oxford; Academic Press, 2003; 5465-5477.
- 6. Sharma G, and Nautiyal A. Cinnamomum tamala: A valuable tree from Himalayas. International Journal of Medicinal and Aromatic Plants, 2011; 1(1): 1–4.
- Rao BRR and Patel RP (2011). An Overview of Cinnamon (*Cinnamomum verum* Presl.) with Reference to Northeast India. In Baruah A, Nath SC. (eds.). Aromatic and spice plants: Utilization and Conservation, 1st ed., Jaipur; Aavishkar Publishers, 2011; 69-100.
- Nabavi SF, Di Lorenzo A, Izadi M, Sobarzo-Sánchez E, Daglia M, and Nabavi SM. Antibacterial effects of cinnamon: From farm to food, cosmetic and pharmaceutical industries. Nutrients, 2015; 7(9): 7729–7748.
- Tendencia EA Ruangpan L. Disk diffusion method. In Laboratory manual of standardized methods for antimicrobial sensitivity tests for bacteria isolated from aquatic animals and environment, Tigbauan, Iloilo, Philippines; Aquaculture Department, Southeast Asian Fisheries Development Center, 2004; 13-29.
- Loehrer M, Delventhal R, Weidenbach D, and Schaffrath U. Bacterial Growth Inhibition Assay for *Xanthomonas oryzae* pv. oryzae or *Escherichia coli* K12 Grown together with Plant Leaf Extracts. Bio-Protoc., 2016; 6(24): 1–5.
- 11. Ogundare AO, and Oladejo BO. Antibacterial Activities of the Leaf and Bark Extract of *Persea americana*. American Journal of Ethnomedicine, 2014; 1(1): 64-71.
- 12. Rodríguez-Carpena JG, Morcuende D, Andrade MJ, Kylli P, and Estevez M. Avocado (*Persea americana* Mill.) phenolics, in vitro antioxidant and antimicrobial activities, and inhibition of lipid and protein oxidation in porcine patties. J. Agric. Food Chem., 2011; 59(10): 5625–5635.

- Amado DAV, Helmann GAB, Detoni AM, de Carvalho SLC, de Aguiar CM, Martin CA, Tiuman TS, and Cottica SM. Antioxidant and antibacterial activity and preliminary toxicity analysis of four varieties of avocado (*Persea americana* Mill.). Braz. J. Food Technol., 2019; 22: 1–11.
- 14. Dennis, Nurliza C, and Savitri W. Antibacterial effect of ethanol extract of the avocado seed (*Persea americana* Mill.) as an alternative root canal Irrigants against *Porphyromonas gingivalis* (In Vitro). International Journal of Applied Dental Sciences, 2017; 3(1): 89–93.
- 15. Gomes-Flores R, Arzate-Quintana C, Quintanilla-Licea R. Tamez-Guerra P. Tamez-Guerra R. Monreal-Cuevas E, and Rodríguez-Padilla C. Antimicrobial Activity of Persea americana Mill (Lauraceae) (Avocado) and Gymnosperma glutinosum (Spreng.) Less (Asteraceae) Leaf Fractions Extracts and Active Against Mycobacterium tuberculosis. Am.-Eurasian J. Sci. Res., 2008; 3(2): 188-194.
- 16. Ajayi OE, Awala SI, Olalekan OT, and Alabi OA. Evaluation of Antimicrobial Potency and Phytochemical Screening of *Persea americana* Leaf Extracts against Selected Bacterial and Fungal Isolates of Clinical Importance. Microbiol. Res. J. Int., 2017; 20(1): 1–11.
- 17. Raymond Chia TW, and Dykes GA. Antimicrobial activity of crude epicarp and seed extracts from mature avocado fruit (*Persea americana*) of three cultivars. Pharm. Biol., 2010; 48(7): 753–756.
- Khandelwal P, Upendra RS, Raftaniamiri Z, and Ramachandra GG. Assessment of biotherapeutic potential of *Pimenta dioica* (allspice) leaf extract. Int. J. Pharm. Sci. Res., 2012; 3(09): 3379–3383.
- Boyd FAH, and Benkeblia N. In vitro evaluation of antimicrobial activity of crude extracts of *Pimenta dioica* L. (Merr.). Acta Hortic., 2014; 1047: 199– 206.
- 20. Asha MM, Chaithra M, Yashoda K, Vivek MN, and Prashith Kekuda TR. Antibacterial activity of leaf and bark extracts of *Pimenta dioica* (Linn.) Merill against clinical isolates of *Staphylococcus aureus* and *Streptococcus mutans*. World J. Pharm. Pharm. Sci., 2013; 2(5): 3207–3215.
- Kumar BH, Badarudin A, and Jose A. DPPH radical scavenging activity and antibacterial activity of *Pimenta dioica* (L.) Merr. Orient. J. Chem., 2010; 26(4): 1501–1505.
- Vazquez-Cahuich DA, Espinosa Moreno J, Centurion Hidalgo D, Velazquez Martinez JR, Borges-Argaez R, and Caceres Farfan M. Antimicrobial activity and chemical composition of the essential oils of *Malvaviscus arboreus* Cav, *Pimenta dioica* (L.) Merr., *Byrsonima crassifolia* (L.) Kunth AND *Psidium guajava* L. Trop. Subtrop. Agroecosyst., 2013; 16: 505-513.
- 23. Oussalah M, Caillet S, Saucier L, and Lacroix M. Antimicrobial effects of selected plant essential oils on the growth of a *Pseudomonas putida* strain

isolated from meat. Meat Sci., 2006; 73(2): 236-244.

- Hassan W, Zainab Kazmi SN, Noreen H, Riaz A, and Zaman B. Antimicrobial Activity of *Cinnamomum tamala* Leaves. Journal of Nutritional Disorders & Therapy, 2016; 6(2): 190.
- Dhiman R, Aggarwal NK, and Kaur M. Comparative evaluation of antimicrobial activities of commonly used Indian spices against microbes associated with juices. Res. J. Microbiol., 2015; 10(4): 170–180.
- Jain A, Dubey M, Gupta A, Mahajan S, and Chaudhari HS. Antimicrobial activity of *Cinnamomum tamala* (Tejpat) against some bacterial and fungal pathogens. J. Pharm. Res., 2011; 4(11): 3975–3977.
- 27. Thapa B, Singh A, and Tuladhar R. In vitro Antibacterial Effect of Medicinal Plants Against Multidrug Resistant Gram Negative Bacteria. Tribhuvan University Journal of Microbiology, 2018; 5(1): 25–31.
- Goyal P, Chauhan A, and Kaushik P. Laboratory Evaluation of Crude Extracts of *Cinnamomum tamala* for Potential Antibacterial Activity. Electronic Journal of Biology, 2009; 5(4): 75–79.
- 29. Mazimba O, Wale K, Kwape TE, Mihigo SO, and Kokengo BM. *Cinnamomum verum*: Ethylacetate and methanol extracts antioxidant and antimicrobial activity. Journal of Medicinal Plants Studies, 2015; 28(33): 28–32.
- Dhore MR, and Jha AR. Antimicrobial activity of *Allium cepa* and *Cinnamomum zeylanicum* against common bacteria causing urinary tract infections: in vitro study. International Journal of Basic & Clinical Pharmacology, 2019; 8(6): 1185.
- 31. Hamedo HA. Activity of *Cinnamomum zeylanicum* essential oil and ethanolic extract against extended-spectrum β lactamase-producing bacteria. Afr. J. Biotechnol., 2015; 14(4): 304–309.
- 32. Gende LB, Floris I, Fritz R, and Javier EM. Antimicrobial activity of cinnamon (*Cinnamomum zeylanicum*) essential oil and its main components against *Paenibacillus larvae* from argentine. Bulletin of Insectology, 2008; 61(1): 1–4.
- Abdulla E. H., Abdoun M. A., Mahmoud W. S., and Alhamdani F. Antibacterial Activity of Crude *Cinnamomum zeylanicum* Ethanol Extract on Bacterial Isolates from Orofacial Infections. ACTA SCIENTIFIC DENTAL SCIENCES, 2019; 3(8): 58–63.
- 34. Sharifan A, Shafiee M, and Tabatabaee A. Evaluation of Antimicrobial Effect of *Cinnamomum verum* Methanolic Extract and Essential Oil: A Study on Bio-preservative in Ketchup Sauce. J. Chem. Health Risks, 2016; 6(2): 113-124.