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EVALUATION OF BIOCHEMICAL PARAMETER AND HYPOGLYCERMIC POTENTIAL OF COMBRETUM HISPIDUM.LAW ROOT EXTRACT ON ALLOXAN INDUCED DIABETES IN RATS

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ABSTRACT

Antidiabetic activity of *Combretum hispidum*, was checked. Twenty-five rats were used for the research and were grouped into five of five rats each. Groups 1 was the untreated diabetic group while groups 2, 3 and 4 were the treatment which received 200, 400 and 800 mg/kg body weight of the *C. hispidum* extract respectively. Group 5 was the positive control and was administered known antidiabetic drug Glibenclamide. Diabetes was induced in the rats with alloxan monohydrate. *C. hispidum*, extract was administered to rats for 14 days orally by intubation, thereafter were sacrificed and blood collected from heart for analysis. Effect of *C. hispidum*, extract was checked on blood glucose level for possible hypoglycermic potential and biochemical parameters. All results in treatment groups were compared with the untreated diabetic group at statistical confidence of p<0.05. This significant effect was recorded at Day 1, Day 3 and Day 7 of the treatment. Result shows that *C. hispidum* extract reduced blood glucose level in the test groups as dose of extract increased. *C. hispidum*, demonstrated hypoglycermic effect. Biochemical indices indicated safety of liver, kidney cardiac cells.

KEYWORD: Alloxan, Blood glucose, Combretum hispidum, Diabetes, Glucose oxidase, Root.

INTRODUCTION

The disease diabetes mellitus, usually just 'diabetes' is a metabolic disease characterized by raised levels of blood glucose and elevated fat and protein metabolism. It is one of the most prevalent human metabolic defects ADS (2007), Guyton (2006). It is caused by failure of the β cell, of islet of Langerhans of the pancreas to produce insulin. Lenzen, (2008), Ezeja et al (2015). It can be treated successfully by regulating the diet alone otherwise it is necessary to give daily injection of insulin or treat with sulphonylurea drugs. Dietary management is very important in the control of diabetes Bantle et al (2006). Diabetes is a risk factor for cardiovascular diseases such as hypertension, heart failure and nephropathy Bantle et al 2006. Neutritional therapy, counseling and the use of specialized nutritional supplement are recommended for diabetic cases Pastor et al (2002). Chronic complications of diabetes results from elevated blood glucose levels and associated with impairment of lipid and other metabolic pathways Sheetz and King (2003), Diabetic Control Trial (1993). Diabetic nephropathy is the leading cause of chronic kidney

disease/failure, ADS (2007), US Renal Data System (1998). The term diabetic nephropathy is used to describe the combination of lesions that occur concurrently in the diabetic kidney, Sheetz and King (2000).

Diabetic retinopathy which is a leading cause of blindness is closely linked to elevation in blood glucose and hyperlipidemia seen in people with uncontrolled diabetes. Seen in cataracts and glaucoma ADS 2004.

Diabetic neuropathies which can affect the somatic and autonomic nervous systems, results from uncontrolled diabetes Boulten 2004, ACCE 2003.

Macrovascular disorder such as coronary heart disease, stroke and peripheral vascular disease reflect the combined effects of unregulated blood glucose levels, elevated blood pressure and hyperlipidemia due to metabolic disorder Martfin (2007), Grundy and Panel (2001).

Diabetic foot ulcers has been associated with diabetic patients, causing ulceration, infection and eventually the need for amputation, ADS (2004).

The chronic complications of diabetes are best prevented by measures aimed at tight control of glucose levels, maintenance of normal lipid levels and control of hypertension. ADS (2004). Hypoglycermia occur when an agent, drug or plant extract causes lowering of blood glucose level below normal reference range Masharani and Gitelman (2007), ADS Workgroup 2005.

Plants play a vital role in the treatment and prevention of diseases. They help in the prevention and reduction of the adverse side effects of conventional drugs Bachrach sources of biological (2012).Thev are pharmacological important chemicals. It has been reported that plants are sources of successful drugs, and will continuously be in the front line for screening novel lead compounds Atanasov et al (2015). An essential part of organic chemistry and biochemistry of plant is the identification of the novel bioactive compounds present plant leading to further biological pharmacological studies Momin et al (2014), Farid et al (2015), Guo et al (2013).

Combretum hispidum (Laws) (Combretaceae) is a common climbing weed of exist in the forest and savanna region. It regrows rapidly after forest and grass fires. It has trailing branches. It produces from seeds and vegetatively from basal stumps. The leaves are opposite, oblong, elliptic, 10 - 25 cm long and 5 - 11 cm wide. It has a cylindrical woody stem that is covered with short bristly hairs. The outer part of the root is fleshy which covers the inner wooden centre Figure 1b. The pharmacological use of plants of the family Combretaceae is widely reported in the scientific literature Atindehou et al (2004), Muthu et al (2006), Gansane et al (2010). Combretaceae families exist predominately in tropical and subtropical areas, for example, in Africa and Brazil. Pictorial view of the root is shown in Figure 1a.

Phytochemical analysis on the genus Combretum have revealed the presence of triterpenes, flavonoids and nonprotein amino acids, Pietrovsky et al (2006). In the past few decades, numerous unusual phytocompounds have been isolated from Combretum species. It has been that reported 9,10-dihydrophenanthrenes substituted bibenzyl was isolated and characterized from C. molle, Rogers and Verotta (1996). Isolation of eleven triterpenes and their glycosides from C. laxum were reported by Bisoli and co-workers, Bosoli et al (2008). Cycloartane dienone lactone and alkaloids (combretine and betonicine) were isolated from C. quadrangulare, Banskota et al (2000), and C. micranthum, Ogan, (1972). Flavonoids such as rhamnoctrin, quercetin-5,3'dimetylether, ramnazin and kaempferol were isolated from C. erythrophyllum, Martini et al (2004).

Analysis of bioactive phytochemicals present in the leaves of *C. hispidum* was carried out by Ikpeazu *et al* (2020) and revealed presence of antidiabetic compounds. Bioactive compounds in plants have been identified by many researchers using Gass Chromatography-Mass Spectrometry analysis (Igwe *et al* 2016, Otuokere *et al* 2016, Ikpeazu *et al* 2017) There are no published literatures that determine the hypoglycermic potentials of ethanol extracts of *C. hispidum* root hence the research.

This study is aimed to investigate the hypoglycermic potentials and biochemical indices of root of *C. hispidum*. and to disprove or otherwise the natives claim that *C. hispidum* root extract can be used to reduce the blood glucose level of diabetic patients.

MATERIALS AND METHODS



Fig. 1: C. hispidum root.

Plant sample

Fresh roots of *C. hispidum* were collected from Obi-Ngwa, Abia State Nigeria on 24th May, 2020. Sample of plant roots was identified by a Botanist at the Department of Plant Science and Biotechnology, College of Natural Sciences, Michael Okpara University of Agriculture, Umudike, Nigeria.

Extraction of crude extracts

The identified root of *C. hispidum* was shade dried for 10 days and pulverized to a coarse powder using manual grinder [Corona-Landers YC 1A SA]. The plant root extract was prepared using Soxhlet method described by Jensen (2007). Forty-five grams (45g) of coarse powdered sample was introduced into the extraction chamber of the Soxhlet extractor using ethanol as solvent. Temperature was maintained at 70°C throughout the extraction period of 48 hours. At the end of the extraction period the extract was concentrated using oven at 30°C to obtain dried extract which was weighed and kept in a well labelled sterile specimen bottle.

LD₅₀ and Dose selection

A preliminary acute toxicity test was done using rats to determine the LD_{50} (lethal dose that kills 50% of the rats) Acute toxicity (LD_{50}) was determined according to Lorke

method, Lorke (1983). At 5000 mg/kg body weight of administered root extract, the treated rats were still healthy and active. This observation shows that the root extract is safe at dose below 5000 mg/kg b.w. Based on this safety determination, different doses of 200, 400 and 800 mg/kg body weight was prepared and administered to rats in group 2, 3, and 4 respectively. These doses were calculated from a stock solution dissolved in distilled water.

Chemicals

Alloxan was used in this study and was obtained from Sigma and Alderich USA. Other reagents/chemicals used were obtained within Nigeria and were of analytical grade.

Experimental Animals

Adult albino rats (148 to 253 g) were purchased from University Farm. Approval was obtained from College of Vet Medicine, Michael Okpara University of Agriculture Umudike, Nigeria, in line with the guidelines for the care and use of laboratory animals as given by the National Research Council (NRC, 1985). The rats were acclimatized and fed *ad libitum*.

Experimental Design

Twenty-five rats were used for the research were grouped into five of five rats each. Groups 1 was the untreated diabetic group, 2, 3 and 4 were the treatment groups which received 200, 400 and 800 mg/kg body weight of the *C. hispidum*, extract. Group 5 was the positive control and was administered known antidiabetic drug Glibenclamide. Diabetes was induced in the rats with alloxan monohydrate. *C. hispidum*, extract was administered to rats for 14 days orally by intubation, thereafter were sacrificed and blood collected from heart for analysis.

Experimental Diabetes Induction

The method of Lenzen (2008) was adopted. The animals were fasted for 16–18 hours with free access to water

before the induction of diabetes. Induction of diabetes was carried out by single intraperitoneal injection of alloxan monohydrate (Sigma St Louis, M.O., USA) dissolved in 0.9% v/v normal saline solution at a dose of 150 mg/kg body weight (Katsumat *et al.*, 1999). The diabetes was assessed in alloxan induced rats by determining the blood glucose concentration using one touch glucometer and Accu-check strips at day 1 and day 3 after injection of alloxan. The rats that recorded elevated blood glucose level above 240 g/dL were considered diabetic and were selected for the study.

Blood Glucose Levels determination

The procedure of Aziz (1983), Bergman (1984) based on the glucose oxidase principle was adopted in the determination of blood glucose level of the experimental rats. The enzyme glucose oxidase reacts with glucose, water and oxygen to form gluconic acid and hydrogen peroxide. The hydrogen peroxide can then be used to oxidize a chromogen or the consuption of oxygen measured to estimate the amount of glucose present. Glucose oxidase is specific for B-D-glucose so cross reaction with other sugars is not a problem Aziz (1983), Bergman (1984), Howanitz and Howanitz (1984). The blood samples were collected by cutting the tip of the tail artery of the rats, and a drop allowed touching the sensor part of one touch glucometer strips. The values obtained were recorded in mg/dl. The blood glucose levels were sampled at intervals of day 1, day 3 and day 7 of treatment.

Statistical Analysis

All the data were expressed as mean \pm SEM. The data was analysed using SPSS vision 20 Statistical comparisons were performed by one way analysis of variance (ANOVA) followed by Duncan's multiple range tests to separate the mean. The results were considered statistically significant at p<0.05.



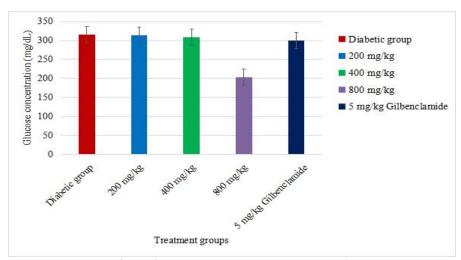


Figure 2: Represent glucose level at Day 1.

Graph in Fig 2 represent hypoglycermic result of root extract of *C. hispidum* on alloxan induced diabetic rats at Day 1 of treatment. Values are presented as Mean \pm Standard Error of Mean (S.E.M.)

There was mild reduction of glucose in the treatment groups 313.40 ± 19.43 , $309.20.20 \pm 11.82$ and 283.00 ± 13.08 when compared to the diabetic untreated group 316.00 ± 20.31 . The reference standard drug was 299.60 ± 16.85 . There was no significant reduction of glucose level at p<0.05 on Day 1 treatment.

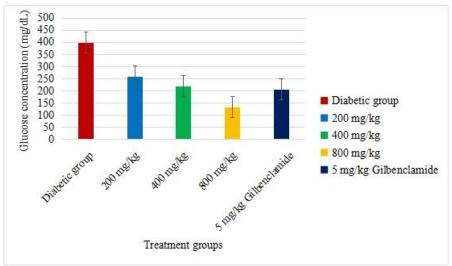


Figure 3: Represent glucose level at Day 3.

Graph in Fig 4 represent hypoglycermic result of root extract of *C. hispidum* on alloxan induced diabetic rats at Day 3 of treatment. Values are presented as Mean \pm S.E.M.

There was more reduction of glucose level in the treatment groups 258.60 ± 22.26 , 258.60 ± 22.26 and 133.60 ± 3.50 when compared to the diabetic untreated group 398.00 ± 25.05 . The reference standard drug was 206.40 ± 14.12 . The reduction was statistically significant at p<0.05 on Day 3 treatment.

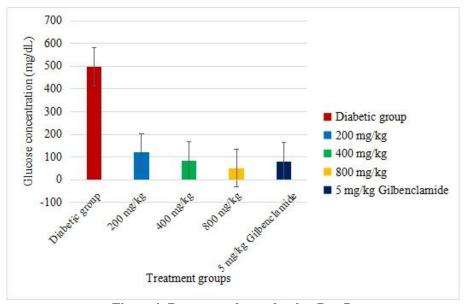


Figure 4: Represent glucose level at Day 7.

Graph in Fig 4 represent hypoglycermic result of root extract of C. hispidum on alloxan induced diabetic rats at Day 7 of treatment. Values are presented as Mean \pm S.E.M.

There was a very noticeable reduction of glucose in the treatment groups 120.40 \pm 3.07, 83.20 \pm 2.26 and 51.20 \pm 1.85 when compared to the diabetic untreated group 496.80 \pm 4.74. The reference standard drug was 81.80 \pm 1.52. The reduction was statistically significant at p<0.05 on Day 7 treatment.

RESULT

The result as presented in Table 1 showed the percentage protection of *C. hispidum* extract on blood glucose

concentration (mg/dl) of alloxan induced diabetes in Wistar rats after seven (7) days of treatment.

Table 1: Percentage Reduction of root extract of *C. hispidum* on mean fasting blood glucose concentration (mg/dl) of alloxan induced diabetic wistar rats after seven days of treatment.

Treatment groups	Value before induction	Day 1	Day 3	Day 7	Percentage protection after Day 7 (%)
5 mg/kg Gilbenclamide	60.60±2.24	290.02±3.27	235.20±5.47 ^b	81.80±1.52°	72.8
Diabetic group	60.80±1.39	315.17±3.16	398.00±25.05 ^a	496.80±4.74 ^a	0.0
200 mg/kg <i>C. hispidum</i> root extract	61.20±0.58	303.06±5.04	258.60±22.26 ^b	120.40±6.07 ^b	61.1
400 mg/kg <i>C. hispidum</i> root extract	61.60±0.92	311.13±2.41	253.40±14.52 ^b	83.20±2.26°	73.1
800 mg/kg <i>C. hispidum</i> root extract	64.40±0.93	302.16±3.65	197.60±41.04 ^b	59.40±6.31 ^d	80.6

Values are presented as mean \pm S.E.M. Different superscripts represent significant differences at p<0.05.

Table 2: Effect of C. hispidium root on lipid profile of alloxan-induced diabetic rats.

Treatment	Total cholesterol (mg/dl)	Triglycerides	LDL (mg/dl)	HDL (mg/dl)	VLDL (mg/dl)
Normal control	6.54 ± 0.37^{c}	0.72 ± 0.18^{b}	3.96 ± 0.47^{c}	1.81±0.05 ^a	0.11 ± 0.03^{c}
Untreated group	8.34±0.45 ^b	1.58±0.33 ^a	6.01 ± 0.58^{b}	1.47 ± 0.10^{b}	0.27 ± 0.06^{b}
200 mg/kg)	4.51 ± 0.20^{d}	0.35 ± 0.01^{b}	3.92 ± 0.03^{c}	1.84±0.01 ^a	0.04 ± 0.02^{c}
400 mg/kg	6.78 ± 0.16^{c}	0.54 ± 0.01^{b}	5.22 ± 0.34^{bc}	1.32±0.01°	0.11 ± 0.02^{c}
800 mg/kg	7.46 ± 0.17^{a}	1.48 ± 0.15^{a}	6.36 ± 0.35^{a}	1.16 ± 0.01^{d}	0.48 ± 0.02^{a}
Glibenclimide (5 mg/kg)	8.15±0.42 ^b	1.34±0.19 ^a	6.02±0.55 ^b	1.51±0.01 ^b	0.25±0.05 ^b

Values are presented as means \pm Standard Error of Mean. Means with different superscript along columns are significantly different at p<0.05.

Table 3: Effect of C. hispidium Root on the liver markers of alloxan-induced diabetic rats.

Treatment	ALP (IU/L)	AST (IU/L)	ALT (IU/L)	Total protein (g/dL)	Albumin (mg/dL)	Globulin (mg/dL)	Total Bilirubin (mg/dL)
Normal group	156.34±6.21 ^b	186.64 ± 7.60^{b}	66.08±3.78 ^b	6.12 ± 0.19^{c}	2.69 ± 0.13^{d}	3.80 ± 0.31^{a}	0.65 ± 0.02^{d}
Untreated group	156.70±8.53 ^b	236.52±14.60 ^a	78.72±4.39 ^a	7.00±0.21 ^b	3.61±0.22 ^b	3.38±0.08 ^{abc}	0.75±0.03°
200 mg/kg)	186.00±3.79 ^a	166.60±1.77 ^{bc}	56.40±0.92°	5.56 ± 0.14^{d}	2.60 ± 0.19^{d}	2.96±0.16°	0.48 ± 0.01^{e}
400 mg/kg	156.98±1.90 ^b	151.40±0.60 ^{cd}	42.00±1.18 ^d	6.23 ± 0.15^{c}	3.08 ± 0.16^{cd}	3.15 ± 0.12^{bc}	0.66 ± 0.01^{d}
800 mg/kg	131.10±0.71°	142.40 ± 1.46^{d}	36.90±2.45 ^d	7.73 ± 0.08^{a}	4.28 ± 0.06^{a}	3.39 ± 0.05^{abc}	0.95±0.01 ^a
Glibenclamide (5 mg/kg)	181.54±11.06 ^a	235.42±2.86 ^a	79.38±2.00 ^a	7.08±0.09 ^b	3.44±0.11 ^{bc}	3.63±0.05 ^{ab}	0.85±0.01 ^b

Values are presented as means ± Standard Error of Mean. Means with different superscript along columns are significantly different at p<0.05. AST: Aspartate Transaminase, ALT: Alanine Aminotransferase; ALP: Alkaline Phosphatase.

Table 4: Effect of C. hispidium root on the kidney markers of alloxan-induced diabetic rats.

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Treatment	Urea (mMol/L)	Creatinine (µMol/L)	Na ⁺ (mMol/L)	Cl (mMol/L)	K ⁺ (mMol/L)	HCO ₃ ⁻ (mMol/L)	
Normal group	32.20±1.85°	0.60 ± 0.02	137.54±5.44	95.12±3.09 ^b	4.57±0.29°	16.42±0.81 ^{bc}	
Untreated group	41.32±1.78 ^b	0.65±0.03	135.24±1.93	101.40 ± 2.76^{b}	5.21 ± 0.37^{bc}	18.58±1.09 ^{bc}	
200 mg/kg)	29.02±0.48°	0.44±0.01	111.00±1.84	86.20±1.68°	3.30 ± 0.15^{d}	14.42±0.14°	
400 mg/kg	38.22±2.13 ^b	0.55±0.02	127.14±0.78	96.50±0.63 ^b	5.15 ± 0.28^{bc}	24.90±2.24 ^a	
800 mg/kg	48.36±0.86 ^a	0.81±0.02	119.86±20.27	110.68±2.41 ^a	7.87 ± 0.09^{a}	25.72±2.07 ^a	
Glibenclamide (5 mg/kg)	38.90±0.53 ^b	1.86±1.19	134.28±0.65	101.66±0.73 ^b	5.54±0.37 ^b	19.26±0.31 ^b	

Values are presented as means \pm Standard Error of Mean. Means with different superscript along columns are significantly different at p<0.05.

DISCUSSION

In this study diabetes was induced in rats by a single intra-peritoneal injection of alloxan monhydrate at 150 mg/kg body weight. Alloxan is a cytotoxic agent known to induce diabetes in a wide variety of animal species by damaging insulin secreting β-cell, resulting in decrease of insulin release. This results in decrease utilization of glucose by the tissues leading to hyperglycermia (Lenzen, 2008). The findings indicate that administration of C. hispidum root extract at the graded dosage on alloxan-induced diabetic rats caused a significant (p < 0.05) reduction of the elevated glucose level. The extract from the root of C. hispidum caused a significant decrease of blood glucose in a dose dependent manner and hypoglycermic effect was highly pronounced at day 7, treatment (Fig 4). We suspect that the antihyperglycemic activity of this root extract may be partly due to insulin release from the existing cells of the pancreas, stimulation of insulin secretion and release, regeneration of β-cell of Langerhans islets or activation of enzymes responsible for glucose utilization (Spasov et al., 2008). This findings is not in isolation as it is in agreement with other studies reported by various researchers, Ezeja et al (2015) who demonstrated that antidiabetic activities of plant extract may be due to its multiple effects involving both pancreatic and extrapancreatic mechanisms. Figure 2 showed that reduction of blood glucose was not significant at Day 1 treatment. Figures 3 and 4 shows significant reduction of the root extract of C. hispidum at Day 3 and Day 7 treatment respectively. The hypoglycermic response was in a dose dependent manner when compared to the diabetic untreated group. The extract competed favourably with the reference drug Gilbenclamide. This suggest that the at higher doses the root extract of C. hispidum can compete favourably with known antidiabetic drugs, hence a good alternative for diabetic cases.

Acute toxicity test shows that ≤ 5000 mg/kg of C. hispidum root was safe and was used for the study. The hypoglycermic potential of C. hispidum was evaluated by checking its ability to reduce the fasting blood sugar (FBS) of rats induced with alloxan monohydrate (150 mg/kg). Elevated serum lipids observed in diabetes mellitus are suspected to cause coronary heart disease in diabetic cases, Murugan et al (2009). There was mild elevation in total cholesterol which was not significant. Triglyceride, LDL, HDL and VLDL showed reduction when compared with untreated diabetic rats. Though these reduction was not significant at p<0.05 (Table 2). This reduction could be beneficial in preventing diabetic complications. We suspect that the reduction is due to a control in lipid metabolism (Cho et al, 2002). Therefore C. hispidum root could be useful in preventing cardiac diseases associated with diabetes. Induction of diabetes with alloxan monohydrate can lead to leakage and elevated levels of liver enzymes ALT, AST, ALP into the blood, Edet et al (2011) as seen in untreated diabetic group (Table 3). The significant dose-dependant reduction in the elevated serum ALT, AST and ALP

after administration of C. hispidum root suggest hepatoprotective effect. This could be due to membrane stabilization and repair of tissue damage, Argawal et al (2012). There was mild reduction in total protein of treated rats when compared with the untreated diabetic rats. This effect was not significant and bilirubin level was not affected (Table 3). There was mild reduction in urea and creatinine levels which is an indication of kidney protective effect of C. hispidum leaf extract. Na+, Cl-, K+ and HCO3- were within normal reference range (Table 4). Diabetes mellitus is among the most common disorder in developed and developing countries (Makund et al., 2008). The disease is increasing rapidly in most parts of the world (Kumar et al., 2008). Hyperglycaemia in diabetic patients is associated with alterations in glucose and lipid metabolism and modification in liver enzyme levels (Jenson and Stender, 1998). Despite the presence of known anti diabetic medicines in the pharmaceutical market, screening for new anti-diabetic sources from plants is a good alternative because they contain substances which are safer for use on diabetes mellitus (Alli Smith, and Adanlawo, 2012). Other plant extracts have been known to show antidiabetic effects (Uhuegbu and Ogbuehi (2002, Nwanjo and Nwokoro (2004).

There is plan for further studies to find the bioactive compounds responsible for this antidiabetic activity in *C. hispidum* root.

CONCLUSION

Root extract of *C. hispidum* has a potent antidiabetic activity which is comparable with the known drug, Gilbenclamide in alloxan - induced diabetic rats and hence maybe a good alternative in the treatment of diabetes. Further studies are hereby recommended to isolate and characterize the active ingredients responsible for the hypoglycermic effect in *C. hispidum* root extract. Biochemical indices indicated liver, kidney and cardiac protective effect.

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DECLARATION OF INTEREST

The authors declare no conflict of interest.

REFERENCES

- 1. Nwanjo, H. H. Nwokoro, E. A. antidiabetic and Biochemical effects aqueous extract of *Vernonia amygdalina* leaf in normoglycaemic and diabetic rats. *J. Innov. Life Sci.*, 2004; 7: 6-10.
- Uhegbiu, E. O. and Ogbuehi, K. J. Effects of aqueous extract (crude) of leaves of *Vernonia* amygdalina Del. Blood glucose, serum albumin and cholesterol levels in diabetic albino rats. Global

- Journals of Pure And Applied Science, 2002; 10 (91): 189-194.
- 3. Alli Smith, Y.R. and Adanlawo, I.G. Hypoglycaemic effect of saponin from the root of *Garcinia kola* (bitter kola) on alloxan-induced diabetic rats. *Journal of Drug Delivery & Therapeutics*, 2012; 2(6): 9-12
- 4. Lenzen, S. The mechanism of alloxan and streptozotocin-induced diabetes. *Diabetologia*, 2008; 51: 216-226.
- 5. Spasov, A.A., Maxeiner, M.P. and Bulanov, A.E. Antidiabetic properties of *Gymnema sylvestre*. *Pharma. Chem. J*, 2008; 42(11): 626-629.
- 6. NRC Guide for the care and use of laboratory animals. National Research Council, National Institute of Health. Bethesta (MD), 1985; 8523.
- 7. Z.Y. Bachrach. Contribution of selected medicinal plants for cancer prevention and therapy. *Acta Fac Medicae Naissensis*, 2012; 29(3): 117-23.
- 8. A.G. Atanasov, B. Waltenberger, E.M. Pferschy-Wenzig T. Linder, C. Wawrosch and P. Uhrin P. Discovery and resupply of pharmacologically active plant-derived natural products: a review. *Biotechnol Adv*, 2015; 3(8): 1582-614.
- M.A. Momin, S.F. Bellah S.M. Rahman, A.A. Rahman, G.M. Murshid, T.B. Emran. Phytopharmacological evaluation of ethanol extract of *Sida cordifolia* L. roots. *Asian Pac J Trop Biomed*, 2014; 4(1): 18-24.
- M.M. Farid, S.R. Hussein, L.F. Ibrahim, M.A. Desouky, A.M. Elsayed and A.A. Oqlah. Cytotoxic activity and phytochemical analysis of *Arum* palaestinum Boiss. Asian Pac J Trop Biomed, 2015; 5(11): 944-947.
- 11. F. Guo, L. Feng, C. Huang, H. Ding, X. Zhang, and Z. Wang. Phenylflavone derivatives from Broussonetia papyrifera inhibit the growth of breast cancer cells *in vitro* and *in vivo*. *Phytochem Lett.*, 2013; 6(3): 331-6.
- 12. K.K. Atindehou, C. Schmid, R. Brun, M.W. Koné, D. Traore. Antitrypanosomal and antiplasmodial activity of medicinal plants from Côte d'Ivoire. *J. Ethnopharmacol*, 2004; *90*: 221–227.
- C. Muthu, M. Ayyanar, N. Raja, S. Ignacimuthu. Medicinal plants used by traditional healers in Kancheepuram District of Tamil Nadu, India. *J. Ethnobiol. Ethnomed.*, 2006; 2. doi:10.1186/1746-4269-2-43.
- 14. Gansané, S. Sanon, L.P. Ouattara, A. Traoré, S. Hutter, E. Ollivier, N. Azas, A.S. Traore, I.P. Guissou, S.B. Sirima. Antiplasmodial activity and toxicity of crude extracts from alternatives parts of plants widely used for the treatment of malaria in Burkina Faso: Contribution for their preservation. *Parasitol. Res.*, 2010; 106: 335–340.
- E.F. Pietrovski, K.A. Rosa, V.A. Facundo, K. Rios, M.C.A. Marques, A.R.S. Santos. Antinociceptive properties of the ethanolic extract and of the triterpene 3β,6β,16β-trihidroxilup-20(29)-ene

- obtained from flowers of *Combretum leprosum* in mice. *Pharmacol. Biochem. Behav*, 2006; 83: 90–99.
- 16. C.B. Rogers and L. Verotta, Chemistry and Biological Properties of the African Combretaceae. In Chemistry, Biological and Pharmacological Properties of African Medicinal Plants; K. Hostettman, F. Chinyanganga, M. Maillard, J.L. Wolfender, Eds; University of Zimbabwe Publications: Harare, Zimbabwe, 1996.
- 17. E. Bisoli, W.S. Garcez, L. Hamerski, C. Tieppo, F.R. Garcez, Bioactive pentacyclic triterpenes from the stems of *Combretum laxum*. *Molecules*, 2008; *13*: 2717–2728.
- 18. A.H. Banskota, Y. Tezuka, Q.T. Kim, K. Tanaka, L. Saiki, S. Kadota Thirteen novel cycloartane-type triterpenes from *Combretum quadrangulare*. *J. Nat. Prod.*, 2000; 63: 57–64.
- A.U. Ogan, The alkaloids in the leaves of Combretum micranthum. Studies on West African medicinal plants. VII. Planta Med., 1972; 21: 210– 217.
- 20. N.D. Martini, D.R.P. Katerere, J.N. Eloff. Biological activity of five antibacterial flavonoids from *Combretum erythrophyllum* (Combretaceae). *J. Ethnopharmacol.* 2004; 93: 207–212.
- 21. Katsumat, K.Y., Katsumat, T.O. and Katsumat, K. Hormonal Metabolism. *Res.*, 1999; 25: 125-126.
- 22. American Diabetic Society (2007). Diabetes fact and figures. [Online] Available: www.diabetes.org. Accessed April 7, 2008.
- 23. Guyton A., Hall J. E. *Medical Physiology* (11th ed., 2006; 961-977). Philadelphia: Elsevier Saunders.
- 24. Bantle J.P., Wylie-Rossett J., Albright A.L., et al. Nutritional recommendation and interventions for disease. A position statement of the American Diabetes Association. *Diabetes Care*, 2006; 29: 2140-2157.
- 25. Pastor J.G., Warshaw H., Daly A., et al. The evidence of the effectiveness of medical nutrition therapy in diabetes management. *Diabetes Care*, 2002; 25: 608-613.
- Masharani U., Gitelman S.E. Hypoglycermic disorders. In Gardner D.G., Shoback D.,(Eds.) Greenspan's basic and clinical endocrinology (8th ed., 2007; 748-769). New York: Lange Medical Books/McGraw-Hill.
- 27. American Diabetic Society Workgroup on Hypoglycermia. Defining and reporting hypoglycermia in diabetes: A report of the American Diabetic Society Workgroup on hypoglycermia. Diabetes Care, 2005; 28: 1245-1249.
- 28. Sheetz M.J., King G.L. Molecular understanding of hyperglycermia's adverse effects for diabetic complications. Journal of the American Medical Association, 2002; 288: 2579-2588.
- 29. The diabetic control and complications Trial Research Group. The effect of intensified treatment of diabetes on the development and progression of long-term complications in insulin-dependant

- diabetes mellitus. New England Journal of Medicine, 1993; 329: 955-977.
- 30. Boulton A.J.M., Malik R.A. Diabetic somatic neuropathies. *Diabetes Care*, 2004; 27: 1458-1486.
- 31. ACCE Male Sexual Disfunction Taskforce. AACE medical guidelines for clinical practice for the evaluation and treatment of male sexual dysfunction: A couple's problem- 2003 update. *Endocrine Practice*, 2003; 9: 77-95.
- 32. U.S. Renal Data System. *USRDS 1998 Annual Data Report*. National Institute of Diabetes and Digestive and Kidney Diseases. NIH publication no, 1998; 98: 3176. Bethesda, MD: National Institutes of Health.
- 33. American Diabetic Society. Diabetic retinopathy. *Diabetes Care*, 2004; 27(1): S84-S87.
- 34. American Diabetic Society. Preventive foot care in people with diabetes. *Diabetes Care*, 2004; 27(1): S63-S64.
- 35. Otuokere I. E., Amaku A.J., Igwe K.K., Chinedum G.C. Medicinal studies on the phytochemical constituents in *Justicia carnae* by GC-MS analysis. *American Journal of Food Science and Health*, 2016; 2(4): 71-77.
- 36. Igwe K.K., Nwankudu O.N., Ijioma S.N., Madubuike A.J., Achi N.K, Screening for Secondary Metabolites in *Huru crepitans* Bark Ethanol Extract Using GC-MS Analysis: a Preliminary Study Approach, *Journal of Science and Technology* Advances, 2016; 1(2): 64-71.
- 37. Ikpeazu O.V., Otuokere I.E., Igwe K.K. Preliminary Studies on the Secondary Metabolites of *Buchholzia Coriacea* (Wonderful Kola) Seed Ethanol Extract by GC-MS Analysis, *International Journal of Research in Engineering and Applied Sciences*, March-2017; 7(3): 17~26
- 38. Ikpeazu O.V., Otuokere I.E., Igwe K.K. Gas Chromatography–Mass Spectrometric Analysis of Bioactive Compounds Present in Ethanol Extract of *Combretum hispidum* (Laws) (*Combretaceae*) Root. Communication in Physical Sciences, 2020; 5(3): 325-337.
- Aziz S, Hsiang YH, Comparative study of home blood glucose monitoring devices: visidex, Chemstrip bH, Glucometer and Accu-Chek bG. *Diabetes Care*, 1983; 6: 529-532. [PubMed:6653308]
- 40. Bergman M, felig P. Self monitoring of blood glucose levels in diabetes, Principles and practice. *Arch Intern Med.*, 1984; 144: 2029-2034. [PubMed:6385899]
- 41. Howanitz PJ, Hawanitz JH. Carbohydrares. In: Henry JB,ed. *Clinical diagnosis and management by laboratory methods*. Philadelphia:W.B.Saunders, 1984; 168-169.
- 42. Lorke D. A new approach to practical acute toxicity testing. Arch. Toxicol. 1983; 54: 275-287. http://dx.doi.org/10.1007/BF01234480.
- 43. Ezeja M I., Anaga A O., Asuzu I U. Antidiabetic, antilipidemic, and antioxidant activities of *Gouanialongipetala* methanol leaf extract in

- alloxan-induced diabetic rats, *Pharmaceutical Biology*, 2015; 53(4): 605-614, DOI:10.3109/13880209.2014.935864. https://doi.org/10.3109/13880209.2014.935864
- 44. Alli Smith, Y.R. and Adanlawo, I.G. Hypoglycaemic effect of saponin from the root of garcinia kola (bitter kola) on alloxan-induced diabetic rats. *Journal of Drug Delivery & Therapeutics*, 2012; 2(6): 9-12.
- 45. Jensen, T. and Stender, D. T. Abnormalities in Plasma Concentration of Lipoprotein and Fibrinogen in Type 1 (Insulin Dependent) Diabetic Patients with Increased Urinal Albumin Excretion. *Diabetology*, 1998; 31: 142-145.
- 46. Kumar, A., Ilavarasan, R., Jayachandran, T., Deecaraman, M., Aravindan, P., Padmanabhan, N. and Krishan, M. R. V. Hypoglycemic and Hypolipidemic Effects of *Strobilanthes heyeneanus* in Alloxan Diabetic Rats. Journal of Medicinal Plants and Research, 2008; 9: 246-249.
- 47. Lenzen, S. The mechanism of alloxan and streptozotocin-induced diabetes. Diabetologia, 2008; 51: 216-226.
- 48. Makund, H., Rao, C. M., Srinivasan, K. K., Mamathadevi, D. S. and Satish, H. Hypoglycemic and hypolipidemic effects of *Strobilanthes heyeneanus* in alloxan induced diabetic rats. *Pharmacogenesis Magazine*, 2008; 15: 819-824.
- 49. Spasov, A.A., Maxeiner, M.P. and Bulanov, A.E. Antidiabetic properties of *Gymnema sylvestre*. *Pharma. Chem. J*, 2008; 42(11): 626-629.
- 50. Murugan M, Uma C, Reddy M. Hypoglycaemic and hypolipi demic activity of leaves of Mucuna pruriens in alloxan-induced diabetic rats. J Pharm Sci Technol, 2009; 1: 69–73.
- 51. Edet EE, Atangwho IJ, Akpanablatu MI, et al. Effect of Gongronema latifolium leaf extract on some liver enzymes and protein levels in diabetic and non-diabetic rats. J Pharm Biomed Sci., 2011; 1: 104–7.
- 52. Cho SY, Park TY, Park IM, et al. Alteration of hepatic antioxidant enzyme activities and lipid profile in STZ induced diabetic rats by supplementation of daridetion water extract. Clin Chem Alta, 2002; 317: 109–17.
- 53. Argawal V, Sharma AK, Upadhyay A, et al. Hypoglycaemic effects of Citullus colocynthis roots. Acta Pol Pharm., 2012; 9: 75–9.
- 54. Martfin G. Challenges in developing drug for the metabolic syndrome. *British Journal of Diabetes and Vascular Disease*, 2007; 7: 152-156.
- 55. Grundy S. M., Panel Chair. Third Report of the National Cholesterol Education Program (NCEP) Expart Pannel on Detection, Evaluation and Treatment of High Blood Cholerterol in Adult (Adult Treatment Panel III). NIH publication no. 01-3670. Bethesda. MD: National Institute of Health, 2001.
- 56. Alli Smith, Y.R. and Adanlawo, I.G. Hypoglycaemic effect of saponin from the root of *garcinia kola* (bitter kola) on alloxan-induced

- diabetic rats. *Journal of Drug Delivery & Therapeutics*, 2012; 2(6): 9-12.
- Jensen, T. and Stender, D. T. Abnormalities in Plasma Concentration of Lipoprotein and Fibrinogen in Type 1 (Insulin Dependent) Diabetic Patients with Increased Urinal Albumin Excretion. *Diabetology*, 1998; 31: 142-145.
- 58. Kumar, A., Ilavarasan, R., Jayachandran, T., Deecaraman, M., Aravindan, P., Padmanabhan, N. and Krishan, M. R. V. Hypoglycemic and Hypolipidemic Effects of *Strobilanthes heyeneanus* in Alloxan Diabetic Rats. Journal of Medicinal Plants and Research, 2008; 9: 246-249.
- Lenzen, S. The mechanism of alloxan and streptozotocin-induced diabetes. Diabetologia, 2008; 51: 216-226.
- 60. Makund, H., Rao, C. M., Srinivasan, K. K., Mamathadevi, D. S. and Satish, H. Hypoglycemic and hypolipidemic effects of *Strobilanthes heyeneanus* in alloxan induced diabetic rats. *Pharmacogenesis Magazine*, 2008; 15: 819-824.
- 61. Spasov, A.A., Maxeiner, M.P. and Bulanov, A.E. Antidiabetic properties of *Gymnema sylvestre*. *Pharma. Chem. J*, 2008; 42(11): 626-629.
- 62. Murugan M, Uma C, Reddy M. Hypoglycaemic and hypolipi demic activity of leaves of Mucuna pruriens in alloxan-induced diabetic rats. J Pharm Sci Technol, 2009; 1: 69–73.
- 63. Edet EE, Atangwho IJ, Akpanablatu MI, et al. Effect of Gongronema latifolium leaf extract on some liver enzymes and protein levels in diabetic and non-diabetic rats. J Pharm Biomed Sci., 2011; 1: 104–7.
- 64. Cho SY, Park TY, Park IM, et al. Alteration of hepatic antioxidant enzyme activities and lipid profile in STZ induced diabetic rats by supplementation of daridetion water extract. Clin Chem Alta, 2002; 317: 109–17.
- 65. Argawal V, Sharma AK, Upadhyay A, et al. Hypoglycaemic effects of Citullus colocynthis roots. Acta Pol Pharm., 2012; 9: 75–9.