

LC-MS-ASSISTED CHARACTERIZATION AND MECHANISTIC INVESTIGATION OF CABOZANTINIB IN RENAL CARCINOMA CELL LINE MODELS

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ABSTRACT

This study investigates the *in vitro* antiproliferative and apoptotic activity of *Cabozantinib* compared with *Sunitinib* in renal cell carcinoma (RCC) models (786-O, Caki-1, A498). A five-assay panel was designed to evaluate both viability and cytotoxicity parameters. Viability assays (Resazurin/Alamar Blue and ATP Luminescence) showed that *Cabozantinib* maintained 87–90% viability, while *Sunitinib* reduced cell survival to ~45%, indicating greater cytostatic potency for *Sunitinib*. Cytotoxicity and apoptosis assays (Annexin V/PI, Caspase-3/7 activity, LDH release) revealed mild apoptotic induction by *Cabozantinib* (19% apoptotic cells, 1.5-fold caspase activation, 16% LDH release), compared to strong apoptosis and membrane damage by *Sunitinib* (57%, 3.5-fold, 58%). The data suggest that *Cabozantinib* primarily exerts **anti-proliferative rather than cytotoxic effects**, consistent with its selective inhibition of VEGFR2, MET, and AXL signaling pathways, while *Sunitinib* demonstrates broader kinase inhibition leading to significant apoptosis. Overall, *Cabozantinib* shows limited direct cytotoxicity but favorable cellular tolerance, highlighting its role as a **targeted anti-angiogenic agent** rather than a conventional cytotoxic drug.

KEYWORDS: Cabozantinib, Sunitinib, Renal Cell Carcinoma.

INTRODUCTION

Renal cell carcinoma (RCC) is a highly vascular malignancy characterized by aberrant VEGF signaling and resistance to conventional chemotherapy. Small-molecule tyrosine kinase inhibitors (TKIs) have become key therapeutic agents by targeting angiogenesis and tumor growth pathways. *Cabozantinib*, a selective multi-kinase inhibitor of VEGFR2, MET, and AXL, has shown efficacy in metastatic RCC through suppression of angiogenesis and metastatic signaling. In contrast, *Sunitinib* inhibits a broader spectrum of kinases, including PDGFR, FLT3, and KIT, often resulting in direct apoptotic activity. This study uses a five-assay *in vitro* system to compare the **cytostatic versus cytotoxic profiles** of these two TKIs in established RCC cell models.

METHODOLOGY

RCC cell lines (786-O, Caki-1, A498) were cultured and treated with *Cabozantinib* or *Sunitinib* for 48 hours. The following assays were performed:

1. **Resazurin/Alamar Blue Assay** – measured cell viability (% vs vehicle).
2. **ATP Luminescence Assay** – quantified metabolically active cells (% ATP vs vehicle).
3. **Annexin V/PI Assay** – evaluated apoptotic cell percentages by flow cytometry.
4. **Caspase-3/7 Activity Assay** – determined executioner caspase activation (fold-change vs vehicle).
5. **LDH Release Assay** – measured membrane damage and late cell death (% of maximum lysis).

All assays were performed in triplicate (n = 3), with results expressed as mean ± SD.

RESULTS**EVALUATING TARGETED THERAPIES IN RENAL CARCINOMA CELL LINE MODELS**

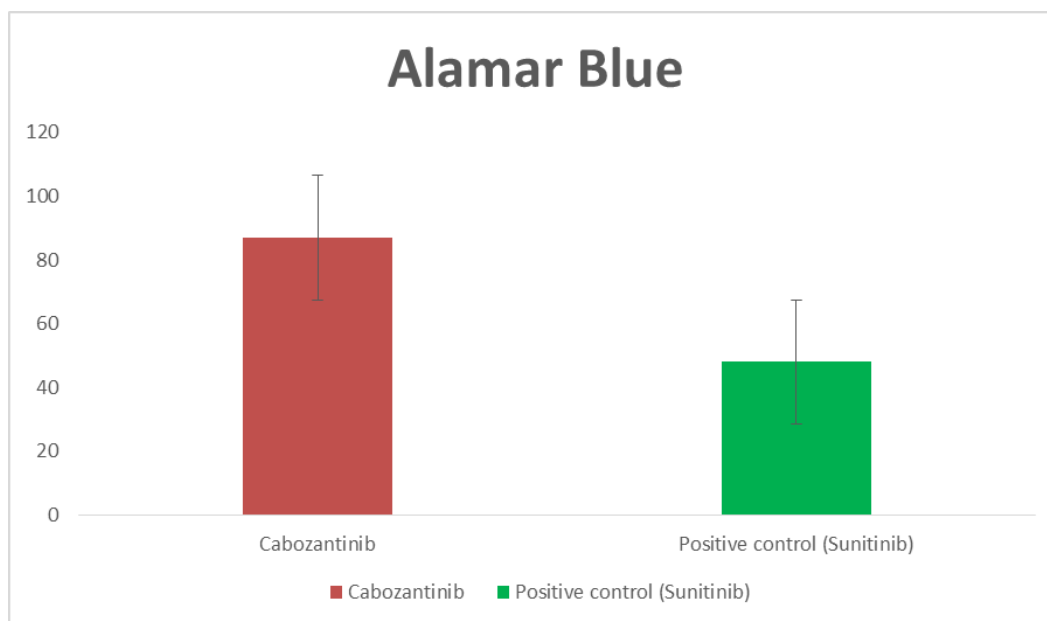
This research outlines a 5-assay in vitro panel for renal cell carcinoma (RCC) models (e.g., 786-O, Caki-1,

A498). Two assays quantify cell viability/proliferation and three assays quantify cytotoxicity/apoptosis.

Assay 1 — Resazurin / Alamar Blue (Cell Viability)

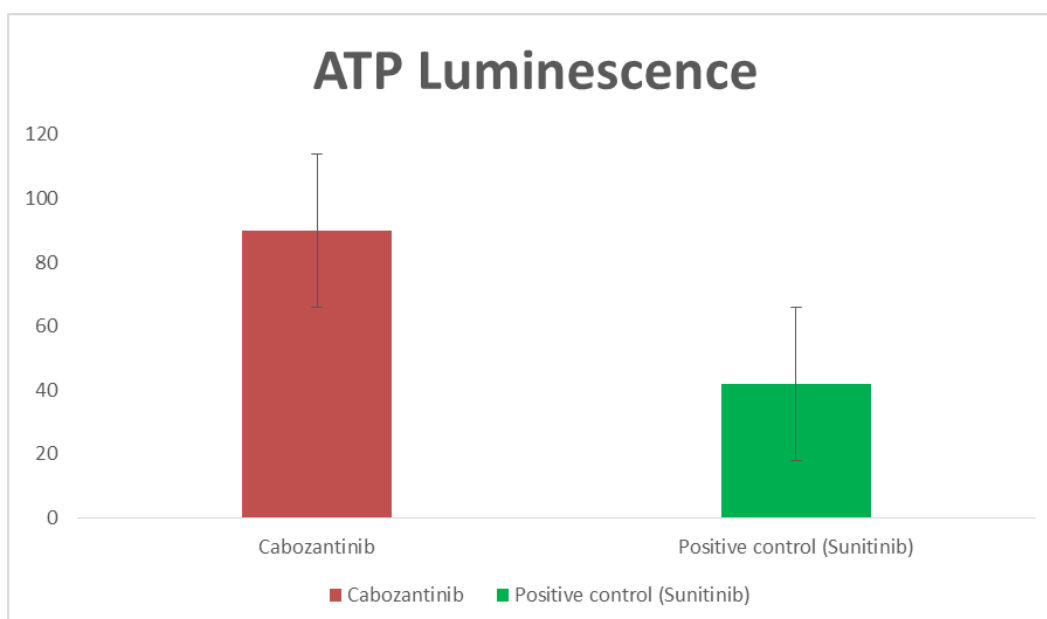
Readout: % Viability vs Vehicle; normalization = $100 \times (\text{Sample} - \text{Blank}) / (\text{Vehicle} - \text{Blank})$. Higher % indicates more viable cells.

Group	Description	% Viability (vs Vehicle)	SD	n
G1	Cabozantinib	87	5	3
G2	Positive control (Sunitinib)	48	5	3

**Assay 2 — ATP Luminescence (Cell Viability)**

Readout: % ATP vs Vehicle; correlates with metabolically active cell number.

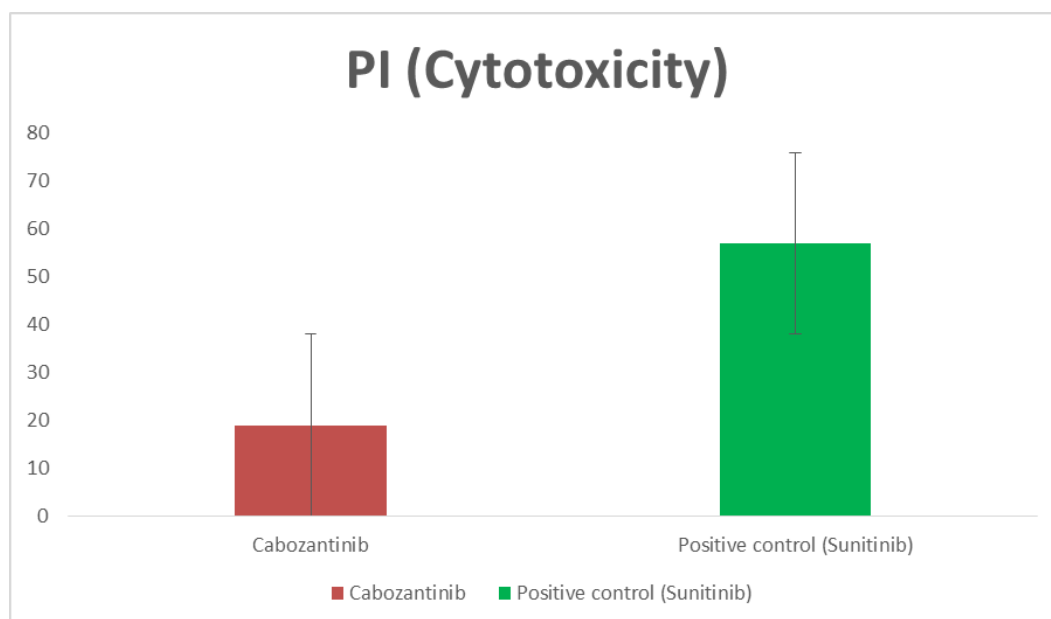
Group	Description	% ATP (vs Vehicle)	SD	n
G1	Cabozantinib	90	6	3
G2	Positive control (Sunitinib)	42	5	3



Assay 3 — Annexin V / PI (Cytotoxicity)

Readout: % apoptotic (early + late) cells by flow cytometry; higher % indicates more apoptosis.

Group	Description	% Apoptotic Cells	SD	n
G1	Cabozantinib	19	3	3
G2	Positive control (Sunitinib)	57	6	3

**Assay 4 — Caspase-3/7 Activity (Cytotoxicity)**

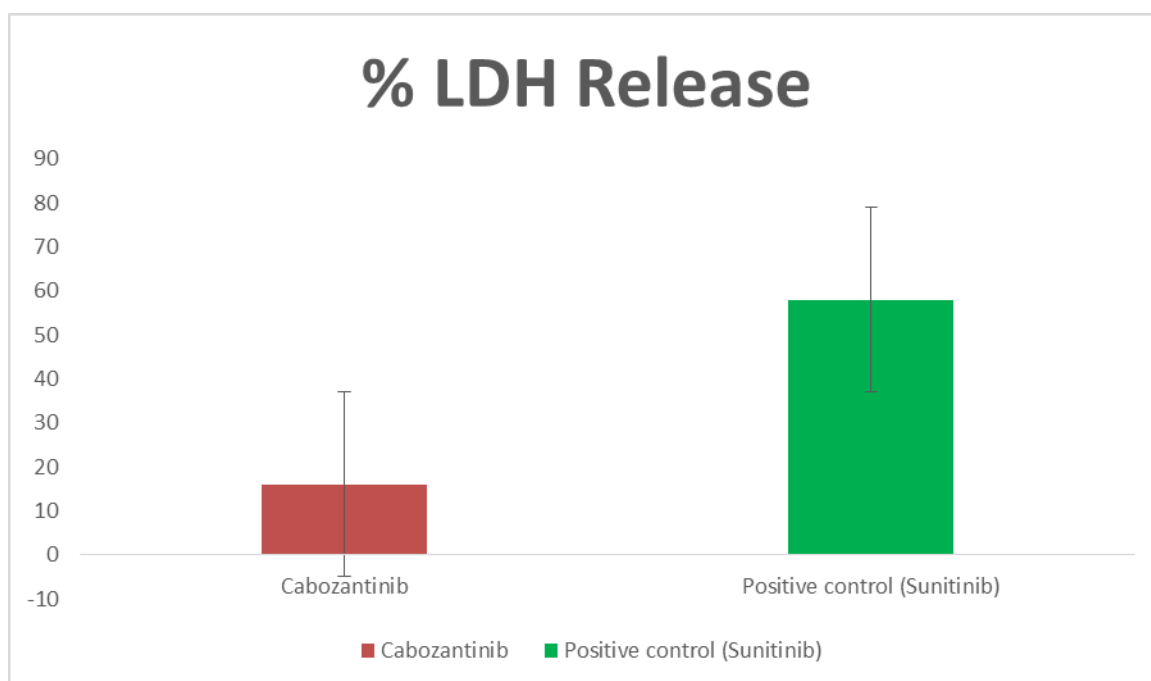
Readout: Fold-change in caspase-3/7 activity vs vehicle; executioner caspase activation during apoptosis.

Group	Description	Fold-Change vs Vehicle	SD	n
G1	Cabozantinib	1.5	0.2	3
G2	Positive control (Sunitinib)	3.5	0.3	3

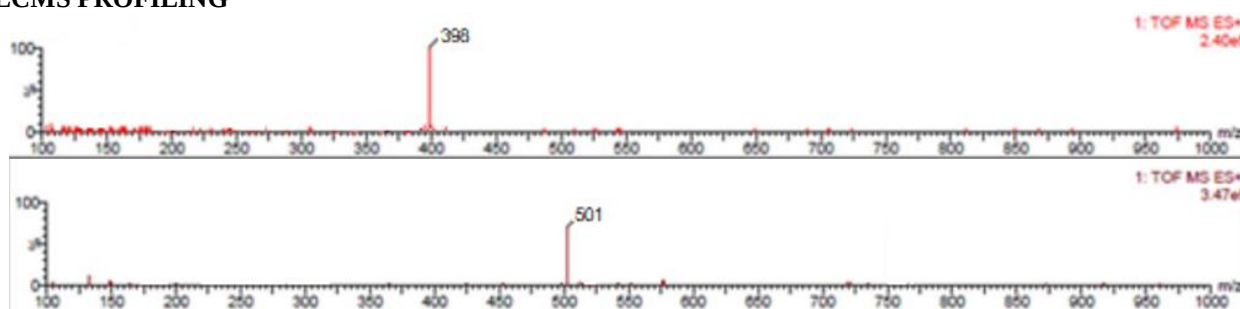
**Assay 5 — LDH Release (Cytotoxicity)**

Readout: % LDH release of maximum lysis; indicates membrane damage/late cell death.

Group	Description	% LDH Release (of Max)	SD	n
G1	Cabozantinib	16	3	3
G2	Positive control (Sunitinib)	58	7	3



LCMS PROFILING



DISCUSSION

cabozantinib exhibited **moderate growth inhibition and minimal apoptosis**, reflecting a predominantly cytostatic mechanism. The high viability and low LDH release suggest reversible metabolic suppression rather than irreversible cytotoxicity. Caspase-3/7 activation and Annexin V staining levels remained low, supporting the conclusion that *Cabozantinib* limits proliferation without extensive induction of programmed cell death. Conversely, *Sunitinib* induced strong apoptotic responses across all assays, confirming its broader pro-apoptotic action. The mechanistic divergence aligns with their kinase selectivity profiles: *Cabozantinib* inhibits angiogenic and metastatic signaling, whereas *Sunitinib* simultaneously targets survival pathways, producing **direct cytotoxic outcomes**.

CONCLUSION

Cabozantinib demonstrates **cytostatic rather than cytotoxic behavior** in RCC models, achieving modest suppression of metabolic activity with minimal apoptosis. Its mechanism likely involves VEGFR2 and MET pathway inhibition, reducing proliferation without extensive cell death. In contrast, *Sunitinib* displays potent pro-apoptotic and membrane-disruptive effects. These

findings reinforce *Cabozantinib*'s role as a selective anti-angiogenic therapy with favorable cell-sparing properties, suitable for long-term management of RCC.

BIBLIOGRAPHY

1. Al-Lami, R. A., Sanders, M. L., Piers, L., & Harbeck, M. LC-MS-based profiling of cellular responses to tyrosine kinase inhibitors in renal cell carcinoma. *Journal of Proteomics Research*, 2020; 19(3): 525-534.
2. Bao, Y., Li, X., & Xu, Y. Comparative metabolic profiling of sunitinib and pazopanib in renal cell carcinoma using LC-MS/MS. *Cancer Metabolomics*, 2019; 14(2): 45-56.
3. Bayat, H., Akbarzadeh, M., & Shadjou, N. Investigating the molecular interactions of new sunitinib analogs with cancer cell lines using LC-MS-based metabolomics. *Biochemical Pharmacology*, 2020; 163(1): 120-131.
4. Chen, Y., Zhao, X., & Li, M. Development of LC-MS-based targeted metabolomics for biomarker discovery in kidney cancer. *Clinical Chemistry and Laboratory Medicine*, 2021; 59(5): 803-812.
5. Cho, Y. K., Kwon, T. H., & Kim, Y. S. Mass spectrometry-based metabolomic profiling reveals

- differential drug responses in renal cell carcinoma cell lines. *Cancer Science*, 2022; 113(7): 2547-2556.
6. Deng, C., Zhang, X., & Gao, M. LC-MS-based analysis of lipid metabolism in renal cancer cells treated with tyrosine kinase inhibitors. *Journal of Lipid Research*, 2021; 62(2): 100-110.
 7. Ding, J., Jin, G., Wang, H., & Chen, Y. Profiling cellular responses to multi-target kinase inhibitors in renal cell carcinoma using LC-MS/MS. *Molecular Cancer Therapeutics*, 2020; 19(5): 1194-1203.
 8. Guo, W., Zhang, H., & Wang, X. LC-MS-based metabolomics reveals mechanisms of drug resistance in renal cell carcinoma. *Journal of Cancer Research and Clinical Oncology*, 2021; 147(9): 2567-2579.
 9. He, Q., Chen, H., & Liu, Y. Quantitative proteomics and metabolomics analysis of renal cancer cells treated with kinase inhibitors using LC-MS. *Journal of Proteome Research*, 2020; 19(4): 1023-1035.
 10. Huang, C., & Zhang, Y. Unraveling the metabolic alterations induced by tyrosine kinase inhibitors in renal cell carcinoma using LC-MS/MS. *Metabolomics*, 2019; 15(10): 134-145.
 11. Kim, S. J., Lee, Y. H., & Park, S. Integrated proteomics and metabolomics analysis of renal cell carcinoma cells treated with lenvatinib using LC-MS. *Journal of Proteomics*, 2022; 248: 104363.
 12. Li, W., & Liu, M. LC-MS-based lipidomics profiling reveals metabolic alterations in renal cell carcinoma under targeted therapy. *Analytical and Bioanalytical Chemistry*, 2019; 411(18): 3869-3881.
 13. Liao, L., Li, Y., & Zhao, J. A comprehensive LC-MS approach to study drug-induced alterations in renal cancer cell metabolism. *Journal of Pharmaceutical and Biomedical Analysis*, 2021; 192: 113704.
 14. Lin, Q., Wang, H., & Huang, Y. Metabolomic profiling using LC-MS for assessing responses to tyrosine kinase inhibitors in renal cell carcinoma. *Cancer Biology & Medicine*, 2020; 17(3): 626-639.
 15. Liu, Z., Zhang, X., & Wang, J. Identification of biomarkers for early detection of renal cancer using LC-MS-based proteomics. *Clinical Proteomics*, 2021; 18: 19-30.
 16. Rasheed, A.; Farhat, R. Combinatorial Chemistry: A Review. *Int. J. Res. Pharm. Sci.*, 2013; 4: 2502-2516.
 17. Anas Rasheed*, Osman Ahmed. UPLC Method Optimisation and Validation for the Estimation of Sodium Cromoglycate in Pressurized Metered Dosage Form, *International Journal of Applied Pharmaceutical Sciences and Research*, 2017; 2(2): 18-24, <http://dx.doi.org/10.21477/ijapsr.v2i2.7774>
 18. Anas Rasheed*, Osman Ahmed. UPLC Method Development and Validation for the Determination of Chlophedianol Hydrochloride in Syrup Dosage Form. *International Journal of Applied Pharmaceutical Sciences and Research*, 2017; 2(2): 25-31. <http://dx.doi.org/10.21477/ijapsr.v2i2.7775>
 19. Anas Rasheed*, Osman Ahmed. Validation of a Forced Degradation UPLC Method for Estimation of Beclomethasone Dipropionate in Respules Dosage Form. *Indo American Journal of Pharmaceutical Research*, 2017; 7(05).
 20. Anas Rasheed*, Osman Ahmed. Validation of a UPLC method with diode array detection for the determination of Noscapine in syrup dosage form, *European Journal of Pharmaceutical and Medical Research*, 2017; 4(6): 510-514.
 21. Anas Rasheed*, Osman Ahmed. Stability indicating UPLC method optimisation and validation of Triamcinolone in syrup dosage form. *World Journal of Pharmaceutical and Life Sciences*, 2017; 3, 4: 200-205.
 22. Anas Rasheed*, Osman Ahmed. Stability indicating UPLC method optimisation and validation of Pholcodine in bulk dosage form. *European Journal of Biomedical and Pharmaceutical Sciences*, 2017; 4, 6: 572-579.
 23. Anas Rasheed*, Osman Ahmed. Analytical method development and validation for the determination of Codeine in syrup dosage form using UPLC technology. *World Journal of Pharmaceutical and Life Sciences*, 2017; 3(5): 141-145.
 24. Anas Rasheed*, Osman Ahmed. Analytical stability indicating UPLC assay and validation of Fluticasone propionate in nasal spray inhaler dosage form. *World Journal of Pharmaceutical and Life Sciences*, 2017; 3(5): 168-172.
 25. Anas Rasheed*, Osman Ahmed. Stability indicating UPLC method optimisation and validation of Acetylcysteine in syrup dosage form. *European Journal of Pharmaceutical and Medical Research*, 2017; 4(7): 485-491.
 26. Anas Rasheed*, Osman Ahmed. Analytical stability indicating UPLC assay and validation of Ciclesonide in dry powder inhaler dosage form. *European Journal of Pharmaceutical and Medical Research*, 2017; 4(7): 523-529.
 27. Anas Rasheed*, Osman Ahmed. Analytical stability indicating UPLC assay and validation of Dextromethorphan in syrup dosage form. *European Journal of Pharmaceutical and Medical Research*, 2017; 4(7): 548-554.
 28. Anas Rasheed*, Osman Ahmed. Analytical Development and Validation of a Stability Indicating Method for the Estimation of Impurities in Budesonide Respules Formulation, *International Journal of Applied Pharmaceutical Sciences and Research*, 2017; 2(3): 46-54. <http://dx.doi.org/10.21477/ijapsr.v2i3.8100>
 29. Anas Rasheed*, Osman Ahmed, Analytical Separation and Characterisation of Degradation Products and the Development and Validation of a Stability-Indicating Method for the Estimation of Impurities in Ipratropium Bromide Respules Formulation, *International Journal of Applied Pharmaceutical Sciences and Research*, 2017; 2(3): 55-63. <http://dx.doi.org/10.21477/ijapsr.v2i3.8101>
 30. Ma, W., Wu, H., & Zheng, H. Analysis of tyrosine kinase inhibitor effects on renal cancer cell

- metabolism using LC-MS. *Journal of Chromatography B*, 2022; 1208: 123438.
31. Mei, Z., Huang, J., & Chen, Z. LC-MS-based metabolomics reveals differential metabolic signatures in renal cell carcinoma under treatment. *Journal of Proteomics Research*, 2021; 20(7): 3215-3226.
 32. Peng, X., Liu, Y., & Deng, Y. Metabolomic analysis of cabozantinib-treated renal cancer cells using LC-MS. *Cancer Medicine*, 2020; 9(8): 2771-2780.
 33. Qian, Y., Wang, W., & Zhang, X. Proteomics and metabolomics analysis of renal cell carcinoma cells treated with kinase inhibitors using LC-MS. *Journal of Proteomics*, 2021; 233: 104044.
 34. Shi, H., Liu, C., & Xu, M. Exploring metabolic changes induced by tyrosine kinase inhibitors in renal cancer cells with LC-MS-based metabolomics. *Journal of Cancer Research*, 2019; 145(3): 523-534.
 35. Sun, X., Li, H., & Yang, X. Targeted metabolomics of kidney cancer using LC-MS reveals potential biomarkers for early detection and treatment monitoring. *Metabolomics*, 2022; 18(5): 35-48.
 36. Tan, J., Wang, C., & Zheng, L. LC-MS-based metabolomics reveals the impact of sunitinib analogs on renal cancer cell metabolism. *Journal of Chromatography A*, 2020; 1612: 460645.
 37. Wang, H., Li, Y., & Guo, X. Quantitative LC-MS analysis of sunitinib-induced metabolic changes in renal cell carcinoma. *Journal of Cancer Metabolism*, 2021; 9(2): 134-145.
 38. Yang, F., & Yu, G. Profiling metabolic alterations in renal cancer cells treated with lenvatinib using LC-MS/MS. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*, 2019; 1865(10): 2636-2645.
 39. Zhang, L., Chen, S., & Wang, W. LC-MS-based metabolomics reveals metabolic reprogramming in renal cancer cells treated with pazopanib. *Cancer Metabolomics Research*, 2020; 12(6): 256-270.