**Review** Atrticle

# World Journal of Pharmaceutical and Life Sciences <u>WJPLS</u>

www.wjpls.org

SJIF Impact Factor: 7.409

# **BACILLUS SPECIES: AN EXCELLENT FERMENTER OF LEGUMES AND OIL SEEDS**

I. C. Oladipo<sup>\*1</sup> and A. O. Adeyemi<sup>1</sup>

<sup>1</sup>Department of Science Laboratory Technonology, Ladoke Akintola University of Technology, Ogbomoso, Oyo State Nigeria.



\*Corresponding Author: Prof. I. C. Oladipo

Department of Science Laboratory Technonology, Ladoke Akintola University of Technology, Ogbomoso, Oyo State Nigeria.

Article Received on 11//04/2025

Article Revised on 01/05/2025

Article Accepted on 21/05/2025

### ABSTRACT

Bacillus species is generally active in the fermentation of legumes and oil seeds due to its highly efficient protein system and adaptable metabolism; this makes it relevant for use as probiotics or feed additives where their heatstability can ensure survival in food matrix or enable long term storage at ambient temperature. In this review, the metabolism of *Bacillus* species in the fermentation of legumes and oil seeds is being observed alongside with its bio-characterization and phenotypic identification. Oil seeds and legumes are very high in protein component, hence, the fermentation of oil seeds and legumes into flavoring condiments by solid state fermentation results from extensive hydrolysis of protein and carbohydrate components as well as lipids, which are mostly used up by these fermenting microorganisms for energy. These proteins are broken down into its main components (amino acids and peptides) after fermentation has taken place likewise carbohydrates (passing through sugar metabolism later converting to organic acids, alcohols and carbonyl compounds) which are present in the resulting metabolite as a result of the microbial activities. Certain metabolites are produced by Bacillus fermenters in the cause of the fermentation; some of which contain certain antibiotics which have been demonstrated to exert antimicrobial activities towards various pathogens, antioxidants which have the ability to prevent risk of infections by protecting cells against harmful oxygen radicals thereby stabilizing cellular membrane, enzymes that possess antithrombolytic activities that can be supplemented with food to be consumed or taken as medication to improve or prevent cardiovascular conditions in humans and other humanly beneficial properties which can be applied in various sectors of life ranging from individual use to industrial use.

KEYWORDS: Bacillus, probiotics, hydrolysis, condiments, metabolites.

#### INTRODUCTION

The microbial interaction during the traditional fermentation of African condiments is determined by the microbiological status of the raw materials, utensils, handlers and production environment which can vary from one community to another and from one processor to another. Various microorganisms grow and enrich the fermentation system of vegetable proteins. Hence, these complex metabolic activities result in production of flavor compounds, metabolites as well as some other technologically useful compounds.

The genus *Bacillus* is a large, heterogenous collection of Gram-positive or Gram-variable spore forming, aerobic or facultative anaerobic bacteria. They can either be obligate aerobes or facultative anaerobes. Cultured species test positive for enzyme catalase if oxygen has been used or present (Favaro *et al.*, 2016). They are ubiquitous in nature and sporulate, therefore, resistant to heat, cold, radiation, desiccation and disinfection. Hence,

they are difficult to eliminate from materials and are a frequent source of contamination (Christie and Setlow, 2020). *Bacillus* have a long history of consumption (Mohammed *et al.*, 2014) and safe use as probiotics (El Shagabee *et al.*, 2017), neutraceuticals (such as vitamins) and carotenoids and have been used for the synthesis of several health supplements. *Bacillus* species are typically found in non-dairy products and are important in diets of lactose intolerant individuals. *Bacillus* offer a potential for industrial applications, being commonly found in the Asian and West African *Bacillus* fermented foods (BFFs), based on soybeans and locust beans(Gopikrishna *et al.*, 2021).

The major fermenting microorganisms involved in fermentation of most vegetable proteins have been identified as proteolytic *Bacillus* species. They include: *Bacillus subtilis*, *B. megaterium*, *B. circulans*, *B. amyloliquefaciens*, *B. licheniformis*, *B. pumilus*, *B. sphaericus*, *B. coagulans*, *B. clausii*, *B. badius*, *and B.*  fusiformis (Isu and Ofuya, 2000; Sanni et al., 2002). However, B. subtilis appears to be predominant of all other members followed by B. coagulans, B. pumilus, B. licheniformis and B. clausii (Jezewska-Frackowiak et al., 2018). Probably, the spores of all Bacillus species can germinate in the gastro-intestinal tract and undergo limited rounds of proliferation before being shed in feaces (Cartman et al., 2008; Tam et al., 2006). They share sporulation ability, forming one oval endospore per cell. *Bacillus* probiotics have been shown to temporarily reside as symbiotic organisms within the host (Dong et al., 2009). The rationale for their beneficial use is based largely on heat-stability of the spore ensuring that the product can be incorporated into foodstuff and animal feeds as well as in liquid formulations (Pepin et al., 2004).

Seeds of legumes may account for up to 80% of dietary protein. Their cooked forms are eaten as meals and commonly used in fermented forms as condiments to enhance flavors of foods (Achi, 2005b). Legumes and oil seeds have constituted significant proportion of the diet of many people. The major components of legumes and oil seeds responsible for condiment production through fermentation are carbohydrates, proteins and fats. As a result, the organisms in charge of fermentation must be able to utilize and hydrolyze these components by degradation, fat oxidation, glycolysis and other reactions to produce primary metabolites such as amino acids, short chain peptides, fatty acids, nucleotides etc. These primary metabolites can be utilized by microorganisms to develop and generate secondary metabolites which can then be broken down further to produce volatile compounds.

# Fermentation of oil seeds and legumes into food condiments

"Iru" and "ogiri" are two most popular indigenous fermented condiments produced from oil seeds and legumes in Nigeria (Omafuvbe *et al.*, 2004); with different names among the different ethnic groups and different varieties of substrates (Ibeabuchi, 2014). Legumes and oil seeds are fermented by allowing the microorganisms to act on them through enzymatic activity to yield condiments in extensive hydrolysis of carbohydrates and protein components (Ibeabuchi *et al.*, 2014). The fermentation is usually carried out in a moist solid state, involving contact with appropriate inoculum of assorted microorganisms and is accomplished by the natural temperatures of the tropics.

The oil seeds and legumes (such as African locust bean, castor seeds, fluted pumpkin seeds, melon seeds *etc*) serving as the raw materials will be sorted to remove bad seeds and unwanted materials and then boiled for a long period of time (about 8-12 h) depending on the raw material used to soften it. The seeds with firmly attached seed coats will be dehulled by slightly pounding with a mortar and pestle, so the seed coats will be washed out of the seeds and the seeds cooked again for about 16 h and

the hot water removed. Then, the seeds or cotyledons will be spread into a calabash tray, covered with wooden tray and wrapped in sacks or banana leaves as the case may be; this is in order to keep the system warm at ambient temperature for 4 days in order to ferment. The end product of this fermentation is our food condiment. This may follow further treatment for preservation depending on the physiological requirements of the substrate and how it is wanted by the consumer.

# Effects of fermentation on the major components of legumes and oil seeds

The main compositions of legumes and oil seeds which are subjected to degradation during fermentation include:

Carbohydrates-Certain carbohydrate degradation enzymes such as fructofuranosidase, galactanase, and glucosidase are produced by Bacillus species (Kiers et al., 2000) and play active role in the hydrolysis of oligosaccharides and indigestible carbohydrates such as raffinose, starchyose and arabinogalactan which are abundant in most legumes (Omafuvbe and Kolawole, 2004) to produce compounds that are readily digestible by human. The nutritional significance of hydrolysis of oligosaccharides is evident in the drastic reduction of the level of indigestible carbohydrates (Nurudeen and Princewill, 2019). Microorganisms produce acids, flavor alcohols and other substances through polysaccharide decomposition (Kashyap et al., 2001). Microbial polysaccharide metabolism involves amylosis, cellulose decomposition and pectinolysis into molecules ready for absorption and utilization (Xu et al., 2020). Microbial amylases hydrolyze carbohydrates into sugars that are digestible by humans; galactanases soften the texture of the seeds and liberate sugars for digestion, sucrases also reduces the total flatus factor of legumes and oil seeds (Nurudeen and Princewill, 2019), starch decomposition is catalyzed by amylase, pectinase also play its role by decomposing pectin (which is widely found in plants, bacteria, molds and yeasts) while the fermented product contains an increased amount of resistant starch (Asencio-Granu et al., 2020). All these enzymes are secreted by Bacillus species while degrading carbohydrate components during fermentation of legumes and oil seeds. Polysaccharide decomposition produces alcohols, acids, and other flavor substances (Kashyap et al., 2001). Monosaccharides are converted into pyruvate through sugar metabolism and are further converted into some organic acids, alcohols and carbonyl compounds and other flavors including ethanol, acetaldehyde, acetic acid and diacetyl (Bartowsky and Pretorius, 2009).

**Proteins-** Proteolytic activity has been found to steadily increase with increase in fermentation period (Ogueke and Aririatu, 2004) and is a major biochemical activity that takes place in fermentation of legumes and oil seeds due to origin of the plant (Ghosh *et al.*, 2013). Metabolic and hydrolytic activities of the *Bacillus* species serve to breakdown the protein into amino acids (Ghosh *et al.*,

2013) with increasing population of Bacillus species from the beginning of fermentation process. Due to high level of hydrolytic enzyme production by Bacillus species have been reported to have one or more enzymatic hydrolytic properties during fermentation (Oguntoyinbo et al., 2007). These enzymes interact with endogenous enzymes of the food itself to degrade proteins into free amino acids and small molecular peptides through ammonia conversion, decarboxylation and dehydrogenation processes, these amino acids are converted into certain aromatic compounds such as aldehydes, ketones, alcohols, acids, indoles, phenols etc (Olasupo and Okorie, 2018). Bacillus strains obtained from fermenting African oil bean seeds and locust beans have been found to produce glutamic acid and extracellular proteinases, which play active role in the fermentation process of these seeds (Nurudeen and Princewill, 2019). Amino acids produced because of protein metabolism are responsible for the gradual pH increase and leveling off towards 7.5-8.0 (Achi, 2005b).

Lipids- The degradation of lipids mainly involves endogenous enzymes produced by the substrates and lipases produced by microorganisms which catalyze autoxidation and enzymatic oxidation. The significant lipolysis of legumes yields predominantly palmitic acids, stearic acids, oleic acids and linoleic acids (Ouoba et al., 2003), octadecenoic acid and other fatty acids are also produced; all of which are further oxidized and degraded to produce linear aldehydes. When lipid is hydrolyzed by lipase, small molecules can further generate flavor compounds including fatty aldehydes, ketones, alcohols and more. Reduction in fat content is attributed to metabolism of *Bacillus* species in the processed product; the breakdown of lipids by lipase and their use as energy source for bacteria (Onwurafor et al., 2014). Many reports confirm that vitamin levels are higher in fermented protein foods than in the raw materials, especially for riboflavin, thiamine, niacin, vitamin C and folic acid due to the metabolic activities of Bacillus species (Leejeerajumnean et al., 2001). Food condiments made from vegetable proteins may be a good source of B-vitamins, but they are found to be deficient in ascorbate and some fat-soluble vitamins, which are lost during fermentation (Ogbonna et al., 2001).

# BIOTECHNOLOGICAL SIGNIFICANCES OF ACTIVITIES OF *BACILLUS* SPECIES

# Antimicrobial Compounds Produced by *Bacillus* Species

In recent years, *Bacillus* species have been demonstrated to exhibit broad-spectrum activity against microbes (Chen *et al.*, 2019). *Bacillus* group has been reported to produce more than 45 antimicrobial molecules; some of these compounds are of clinical values, others are assayed *in vitro* to control food microbes and the remaining ones control plant diseases (Stein, 2005). These metabolites can be grouped majorly into two according to their biosynthetic pathways namely; the peptide antibiotics (also called antimicrobial peptidesAMPs) and the non-peptide based antibiotics (also known as the polyketides) (Puan *et al.*, 2023).

**A. The peptide antibiotics-** These are ribosomally synthesized and non-ribosomally synthesized peptides from the metabolites produced by the species of *Bacillus*.

- Ribosomally synthesized peptides- They are i. widely distributed in nature and contain between 12 and 50 amino acid residues. They are synthesized by ribosomes and are typically cationic and exhibit great structural diversity (Marx et al., 2001). They exist as either post-translationally modified or unmodified peptides (Wu et al., 2023). AMPs synthesized bv ribosomes include subtilin. bacthuricin and cerein, commonly referred to as bacteriocins as they display a high degree of target specificity against closely related bacteria (Chopra et al., 2015). Because of their wide diversity, bacteriocins display different modes of action such as protoplasm vesicularization, pore formation or cell disintegration (Sumi et al., 2015). They are generally bactericidal with some exceptions that exhibit bacteriostatic activities (Gautam and Sharma, 2009; Oladipo et al., 2015). However, those compounds display diverse metabolic activities such as quorum sensing (QS) mediation, cell lysis or induction of genetic competence (Shafi et al., 2017) and so on. Bacteriocin anti-microbial spectra of Bacillus species are broad expanding the search of bioactive molecules to other bacteriocin-producing genera (Pederson et al., 2002; Oladipo et al., 2014; Sumi et al., 2015).
- Non-ribosomally synthesized peptides- Bacillus ii. species strains produce several non-ribosomal compounds through a multistep mechanism including the selection and condensation of amino acid residues such as cyclic lipopeptide (iturin group) and macrolactones (surfactin, fengycins and plipastatins 2005) by multienzyme (Stein, thiotemplates. Large multi subunit enzymes play a key role in the synthesis of these peptides. The nonribosomally synthesized peptides are assembled from among more than 300 different precursors. These peptides can be linear or cyclic and also contain cyclic branched structures containing hydroxyl group, L-amino acids nor D-amino acids. These peptides can be further modified by Nmethylation, acylation, glucosylation or heterocyclic ring formation (Riccardo et al., 2022).

In general, these AMPs are cationic and hydrophobic or amphiphilic, and the cellular membrane of bacteria, in most cases, is the main target for AMPs to exert antimicrobial activity (Zhang *et al.*, 2021). In addition to antibacterial and antifungal properties, *Bacillus*-derived AMPs have also shown antiviral, antitumor and immunomodulatory activities, making them another attractive alternative in recent years (Caulier *et al.*, 2019; Basi-Chipalu *et al.*, 2022). Mechanisms of Action of antimicrobial peptides: Multiple models have been proposed with exact mechanism probably on the specific peptide, concentration and bacterium. Bacteria have also been shown to respond to antimicrobial peptides and even to evolve resistance to their toxic effects (Scott et al., 2008). Different species of Bacillus produce bacteriocins and bacteriocin-like substances with different modes of action. Tochicin, lichenin (Pattnaik et al., 2001), thuricin 439(Ahern et al., 2003) and thuricin S (Chehimi et al., 2010) have all been established to exert a bactericidal effect. In general, bacteriocins are cationic peptides that exhibit hydrophobic or amphiphilic properties, and in most cases, their primary target is the bacterial cell membrane. (Sirtori et al., 2006). Several models have proposed that the mechanism of action of cationic peptides involves the formation of channels through which ions can pass, as well as the disruption of bacterial cytoplasmic membranes (Huang et al., 2009; Palffy et al., 2009). Killing of bacteria via formation of pores in the bacterial membrane requires 3 principal steps: (a) Binding to the bacterial membrane (b) Aggregation within the membrane and (c) Formation of channels. Channel formation leads to breakage of internal cell contents and consequently, cell death.

In addition, antimicrobial peptides must cross the negatively charged outer wall of Gram-negative bacteria, which contain lipopolysaccharides (LPS), outer cell wall of Gram-negative bacteria, which contains acidic polysaccharides. In many cases, specific metabolism of the target microbes provides critical conditions or pore formation (Palffy *et al.*, 2009).

Lipopolysaccharides readily bind to the bacterial surface layer and the local lipid organizational linkages on negatively charged fatty acids, ultimately restructuring the lipid bilayer and thus preventing cellular processes. The fatty acid moiety of lipopeptides contain  $\beta$ -hydroxy fatty acids with 14 carbon chain and inhibit different species of fungi but have only narrow antibacterial activity (Baindara et al., 2013). Gaofu et al. (2010) reported that the antifungal mechanism of a new member of the surfactin family, produced by Bacillus amyloliquefaciens WH1 and named WH1-fungin, exhibits two types of antifungal actions. At high concentrations, it forms pores in the fungal cell membrane, while at low concentrations, it induces apoptosis. Additionally, it inhibits glucan synthase, leading to reduced callose synthesis in the fungal cell wall. WH1-fungin has also been found to bind to an ATPase on the mitochondrial membrane, thereby decreasing its activity in fungal cells. Iturins typically exert their antifungal effects through fungal membrane permeabilization. However, bacillomycin L, produced by B. amyloliquefaciens K103 strain, interacts with other intracellular targets (such as DNA) after disrupting the cell membrane (Zhang et al., 2013). Furthermore, subtilisin produced by B. subtilis has been reported to kill target cells by depleting the transmembrane pH

gradient component of the proton motive force and causing an efflux of intracellular ATP (Noll *et al.*, 2011).

B. Non peptide-based antibiotics: Non peptide-based antibiotics are also called the polyketides. Bacillus species secrete three antimicrobial polyketide groups and their variants (bacillaene, difficidin and macrolactin) and they exhibit antimicrobial activities by selectively inhibiting protein synthesis and may also have synergistic effects against pathogens (Chen et al., 2019). Pinchuk et al. (2001) also reported that at least two antibiotics, one of which is amicomycin, produced by B. subtilis in a starch-based medium were responsible for anti- Helicobacter pylori activity. About 18 macrolactins from Bacillus species have been chemically described and some of them are considered to be potent antiviral and cytotoxic agents that also have antibacterial activity against Staphylococcus aureus (Romco-Tabarez et al., 2006). Difficidin and oxydifficidin are also produced during fermentation of B. subtilis and this represent a class of antibiotics that are active against aerobic and anaerobic organisms (Baruzzi et al., 2011). Hence, bioactive peptides are formed during fermentation by proteolytic microorganisms (De mejia and Dia, 2010).

### Anti-oxidant Activities of Bacillus species

Bacillus fermented foods and condiments have been proven to possess various antioxidants activities. Antioxidants are substances (produced bv microorganisms) that can prevent or slow down chemical reactions that may damage cells caused by free radicals such as nitric oxide, nitrous oxide and peroxynitrite (Ozabor et al., 2020). The antioxidant activity of a biochemical compound corresponds to its capacity to delay or prevent the oxidation of a substrate, resulting from an imbalance between reactive oxygen species (ROS) production and their degradation by antioxidants. Several studies have reported the potentials of Bacillus species as biosurfactant producers such as lipopeptide type surfactants (Joshi et al., 2013) which are amphiphilic cyclic peptides that are linked to a fatty acid hydrocarbon chain and belong to surfactin, iturin and fengycin families; synthesized by non-ribosomal peptide synthases without involving messenger RNA (Leclere et al., 2005) and are found to possess specific biological activities such as antioxidant activities (Tabenne et al., 2012; Ben Ayed et al., 2015).

Carotenoids are antioxidants associated with a range of health benefits, including a reduced risk of cardiovascular diseases, (Fraser and Bramley, 2004; Rao and Rao, 2007) and carotenoid producing bacilli have also been found to be able to reduce symptoms of metabolic syndromes in rats (Cresenzo *et al.*, 2017). Species such as *Bacillus indicus* produces C30 carotenoids (Khaneja *et al.*, 2010; Perez-Fonz *et al.*, 2011) that have been bioavailable at all levels that are possibly superior to other common carotenoids such as astaxancin and lycopene (Sy *et al.*, 2013a). The stability and bioavailability of these *Bacillus* carotenoids have led to the introduction of a number of products worldwide (Kotowicz *et al.*, 2019).

#### Anti-thrombolytic Activities of Bacillus species

Bacillus species secrete various hydrolytic enzymes into their surroundings, and amylases and proteases are the most important enzymes widely utilized for various industrial applications (Schallmey et al., 2004). All fermented foods made from legumes and oil seeds depend on the strong enzyme activities of Bacillus species. Bacillus subtilis, the primary fermenting agent of several fermented oil seeds and legumes, has been extensively studied for its ability to produce extracellular proteases that effectively prevent thrombosis. Among these, certain proteases exhibit strong fibrinolvtic activity. Notably, nattokinase and bacillopeptidase-F are two enzymes with significant fibrinolytic effects, currently utilized as nutritional supplements and considered promising alternatives to conventional drugs for the treatment and prevention of fibrin-related thrombosis. Oral administration of nattokinase has been shown to enhance plasma fibrinolysis in-vivo. In Korea, Bacillus producing these enzymes were isolated from soy products such as *doenjang* (fermented soy paste), ganjang (soy sauce) and cheongukjang (fermented and boiled soy) (Yao et al., 2020). Indonesian fermented "terasi" fermentation explores fibrinolytic capability of several microorganisms such as Bacillus species as naturally occurring bacteria in foods; increasing the knowledge regarding the health benefits of its consumption (Pinonthan et al., 2024). Bacteria strains of Bacillus subtilis A26 (Agrebi et al., 2009), Bacillus amyloliquefaciens (Wei et al., 2011) and Bacillus spp. (Anh et al., 2015) were recently adopted in the production of fibrinolytic enzymes. In addition, research of fibrinolytic enzymes from fermenting Bacillus in fermented legumes such as Chinese soy paste (Wei et al., 2011), Indonesian red oncom and Gembus Tempeh (Afifah et al., 2014), soybean paste (Lee et al., 2001), Japan tofuyo (Syahbanu et al., 2019) and several other traditional fermented foods from legumes and oil seeds are of interest in developing functional foods beneficial to public health (Yoon et al., 2002).

## Other significances include

Enhancement of organoleptic properties- Bacillus 1. species contribute to the development of unique flavor and texture of foods during fermentation while producing peptides and amino acids from proteins and functional materials (Yao et al., 2020). Proteases, for instance, released by the fermenting Bacillus species, produce peptides and amino acids from the protein components of the substrate (such as the low molecular weight peptides and amino acids), improve taste and flavor of the food condiment (Ogbonna et al., 2001). The organoleptic properties of the fermented foods make them more important since it has a wider acceptance than the unfermented foods. Hence, they become more palatable as there will be improvement on the organoleptic properties, texture, aroma and flavor (Osungbaro, 2009).

- 2. Provision of nutritional quality- Improvement in the nutritional value and digestibility of foods has been associated with the actions of Bacillus bacteria in vegetable fermentation (Nout, 2009). The enzymes like amylase, proteases, lipases and phytates modify the primary food products through hydrolysis of polysaccharides, phytates, proteins and lipids (Adeveni and Muhammed, 2008). The quantity of proteins and the content of the watersoluble vitamins increase, while the antinutrient factors (ANFs) in the foods decline during fermentation (Santos et al., 2008). This leads to increased bioavailability of minerals such as zinc. calcium, phosphorous iron and amino acids (Murwan and Ali, 2011).
- Food preservatives- The increasing trend of 3. limiting the use of chemical preservatives has generated interest in the use of natural alternatives. Bacillus has emerged as a promising biopreservative due to its ability to grow efficiently on a large scale at low cost and to produce pH-tolerant bacteriocins with broad-spectrum activity, thereby extending the shelf life of foods (Hoang et al., 2022). Recently. antimicrobial lipopeptide microcapsules prepared from the spray drying of the B. amyloliquefaciens ES-2 strain were tested as food additives (Wang et al., 2014c). Subtilosin is an attractive food preservative alternative to nisin because of its efficacy against L. monocytogenes and other foodborne pathogens (Jung et al., 2008; Sutyak et al., 2008). Bacillus species have been reported inhibit and control to these microorganisms. These antimicrobials can also be purified and applied in food matrix as natural-based preservatives to extend shelf-life of food products as they are considered safe for human consumption due to their antimicrobial and antioxidant properties (Cattelan et al., 2013).
- Detoxification- Enzymes produced by Bacillus 4. species during fermentation of vegetable proteins hydrolyses macromolecules i.e. complex carbohydrates, proteins and fats, thereby enhancing bioavailabilty and digestibility of the fermented product compared to the unfermented substrate (Kiers et al., 2000). Also, enzymatic degradation during fermentation reduces naturally occurring toxic components, allergens and antinutritional components in the raw substrate, thereby, transforming otherwise inedible, difficult to digest or potentially raw materials into palatable and culturally desirable food products that deliver essential nutrients, thus, contributing to a complex rich traditional dietary diversity, with important food security and sustainability implications (Parkouda et al., 2009).
- 5. Improvement of health- Many of the *Bacillus* strains predominant in alkaline fermented foods are also used as probiotics. Some of these strains

include B. cereus, B. clausii, B. coagulans, B. licheniformis, B. polyfermenticus, B. pumilus and B. subtilis- all of which exhibit probiotic activities in both spores and vegetative forms. Fermented plantbased foods could be used as carriers for delivery of Bacillus probiotics into the mammalian systems (Hoa et al., 2000). The influence of probiotics on human gut microbiome is well reported (Thursby and Juge, 2017; Kawai et al., 2018) including that of stimulate Bacillus species. Bacillus strains antihypertensive. antimicrobial. anticancer. antioxidant, fibrinolytic and immunomodulatory activities in vitro and in vivo. These biological properties can be stimulated by metabolic processes of probiotics in alkaline fermented foods.

### CONCLUSION

*Bacillus* species have unique properties, and they produce a number of metabolites: enzymes, antibiotics, vitamins and amino acids. Alkaline fermentation of plant-based foods such as legumes and oil seeds offer several sensorial, nutritional and health benefits. For example, guided selection of probiotic strains could be used to stimulate the *in situ* biofortification of plant-based foods through alkaline fermentation as a strategy to prevent deficiencies in nutrients such as vitamins, minerals and proteins.

Many *Bacillus* species can produce copious number of enzymes which are used in various industries such as in the production of alpha amylase used in starch hydrolysis and protease subtilisin used in detergents. Some others can synthesize and secrete lipopeptides (Nigris *et al.*, 2018; Paul *et al.*, 2021) and several other anti-microbial peptides (Rahman *et al.*, 2020).

Studies on *Bacillus*-derived antimicrobials have already yielded promising results in the field of human health. Therefore, *Bacillus* species can be used as a source of ideal therapeutic tools because of their broad specificity and specific and rapid killing activity against various pathogens. So, effective use of *Bacillus*-derived antimicrobials can be established in medical settings and various industries based on their modes of action, toxicities, and immunogenicities in humans.

Some species of Bacillus are newly discovered to be rich antioxidants; producing riboflavin and in C30 carotenoids. Besides providing a source of vitamin B2 and gastric stable carotenoids, increase in spore levels might confer long-term health benefits. Antioxidants ensure probable migration of bioactive compounds derived from foods, offering a healthier alternative while maintaining oxidative stability and sensory acceptance as well as ingestion of healthy foods containing functional ingredients. Bacillus subtilis is exploited in industry for some B-vitamin production in large scale fermentations, producing reasonable amounts for industrial purposes. subtilis produces riboflavin For example, В. extracellularly, therefore referred to as ideal cell factories

for riboflavin production as they enhance riboflavin level enrichment in fermented foods.

The high protein contents in vegetable proteins including legumes and oil seeds have the potential of being utilized as a media for growing proteolytic and fibrinolytic microorganisms such as the members of the genus *Bacillus* to identify their strong activities and indicate their beneficial health importance. Therefore, proteolytic activities of *Bacillus* are prerequisite of their fibrinolytic activities. Studies about fibrinolytic protease enzymes of bacteria from fermentation products offer novelty and discovery of new proteolytic enzymes potential to be used in cardiovascular diseases treatment. Hence, the bacteria producing fibrinolytic protease enzymes applied in cardiovascular disease treatment are from *Bacillus* group (Hayatun *et al.*, 2020).

Finally, fermented legumes and oil seeds can help in management of lactose intolerance, lowering serum cholesterol level, improving uptake of nutrients, support health management and reducing need for antibiotics.

#### REFERENCES

- 1. Achi, O.K. Traditional fermented protein condiments in Nigeria. African *Journal of Biotechnology*, 2005b; 4(13): 1612-1621.
- 2. Adeyemi, O.T. and Muhammed, N.O. Biochemical assessment of the chemical constituents of *Aspergillus niger* fermented *Chrysophyllum albidum* seed meal. *M.Sc. thesis.* Department of Biochemistry, University of Ilorin, Nigeria, 2008.
- 3. Afifah, D.N., Sulchan, M., Syah, D. and Suhartono, M.T. Isolation and identification of fibrionolytic protease-producing microorganisms from red oncom and embus, Indonesian fermented soybean cakes. *Malaysian Journal of Microbiology*, 2014; 10(4): 273-279.
- 4. Agrebi, R., Haddar, A., Hmidet, N., Jellouli,K., Manni, L. and Nasri, M. BSF1 fibrinolytic enzyme from a marine bacterium *Bacillus subtilis* A26: purification, biochemical and molecular characterization. *Process Biochemistry*, 2009; 44(11): 1252-1259.
- 5. Ahn, M.J., Ku, H.J., Lee, S.H. and Lee H.J. Characterization of a novel fibrinolytic enzyme, bsfa, from Bacillus subtilis ZA 400 in kimchi reveals its persistence to thrombosis treatment. Journal of microbiology and biotechnology, 2015; 25(12): 2090-2099.
- Ahern, M., Verschueren, S., and Van Sinderen, D. Isolation and characterization of a novel bacteriocin produced by *Bacillus thurigiensis* strain B439. *FEMS Microbiol. Lett.*, 2003; 220(1): 127-131.
- 7. Asensio-Granu, A., Calvo-Lerma, J., Heredia, A. and Andre, A. Enhancing the nutritional profile and digestibility of flour by solid state fermentation with *Pleurotus ostreatus. Food funct*, 2020; 11: 7905-7912.
- 8. Baindara, P., Mandal, S.M., Chawla, N., Singh,

Oladipo et al.

P.K., and Korpole, S., Characterization of two antimicrobial peptides produced by a halotolerant *Bacillus subtilis* strain SK.DU.4 isolated from rhizosphere soil sample AMB express, 2013; 3(1): 2.

- 9. Baruzzi F., Quintieri, L., Morea, M., and Caputo, L. Antimicrobial compounds produced by *Bacillus* spp and applications in food. *Communicating current research and technological advances*. National research council of Italy, 2011.
- 10. Basi-Chipalu, S., Sthapit, P. and Dhital, S. A review on characterization, applications and structureactivity relationships of *Bacillus* species-produced bacteriocins. *Drug discov. Ther.*, 2022; 16: 55-62.
- Ben Ayed, H., Bardaa, S., Molla, D., Jridi, M., Maalej, H., qand Sahnoun, Z., Wound healing and in vitro antioxidant activities of lipopolysaccharide mixture produced by *Bacillus mojavensis* A21. *Process biochem*, 2015; 50: 1023-30.
- 12. Cartman, S.T., La Ragione, R.M., and Wooward, M.J. Bacillus subtilis spores germinate in the chicken gastrointestinal tract. *Applied and Environmental Microbiology*, 2008; 74: 5254-5258.
- 13. Cattelan, G.M., Antibacterial activity of oregano essential oil against foodborne pathogens. *Nutrition and food Science*, 2013; 43(2): 169-174.
- Caulier, S., Nannan, C., Gillis, A., Licciardi, F., Bragard, C., and Mahillon, J. Overview of the antimicrobial compounds produced by members of the *Bacillus subtilis* group. *Front. Microbiol*, 2019; 10: 302.
- 15. Chehimi, S., Pons, A.M., Sable, S., Hajlaoui, M.R. and Limam, F. Mode of action of thuricin S, a new class IId Bacteriocin from *Bacillus thurigiensis*, *Canadian Journal of Microbiology*, 2010; 56(2): 162-167.
- Chen, H., McGowan, E.M., Ren, N., Lai, S., Nassif, N., Shad-kaneez F. Nattokinase: a promising alternative in prevention of cardiovascular diseases. *Biomark Insights*, 2018; 13: 1177271918785130.
- Chen, L., Shi, H., Heng, J., Wang, D. and Bian, K. Antimicrobial, plant growth-promoting and genomic properties of the peanut endophyte *Bacillus velezensis* LD02. *Microbiol. Res.*, 2019; 218: 41-48.
- Chopra, L., Singh, G., Choudary, V., and Sahoo, D.K. Sonorensin: an antimicrobial peptide belonging to the heterocycloanthracin subfamily of bacteriocins, from a new marine isolate, *Bacillus sonorensis* MT93. *Applied Environmental Microbiology*, 2014; 80: 2981-2990.
- 19. Christie, G.and Setlow, P. *Bacillus* spore germination: knows, unknowns and what we need to learn". Cellular signalling, 2020; 74: 109729.
- Cresenzo, R., Mazzoli, A., Cancelliere, R., Bucci, A., Naclerio, G., Baccigalupi, L., Cutting, S.M., Ricca, E. and Iossa, S. Beneficial effects of carotenoid-producing cells of bacillus indicus HU16 in a rat model of diet- induced metabolic syndrome. *Beneficial microbes*, 2017; 8: 823-831.
- 21. Cytotoxic potential of industrial strains of *Bacillus* species. *Regul Toxicol pharmacol*, 36: 155-161.

- 22. De mejia, E.G., and Dia, V.P. The role of peptides and nutraceutical proteins in apoptosis, angiogenesis and metastasis of cancer cells. *Cancer metast*, 2010; 29: 511-528.
- 23. Dong, T.C., Van, H. and Cutting, S.M. Bacillus probiotics. Nutra Foods, 2009; 8: 7-14.
- 24. El shagabee, F.M.F., Rokana, N., Gulhane, R.D., Sharma, C., and Panwar, H. *Bacillus* as a potential probiotic: status, concerns, and future perspectives. *Front microbiol*, 2017; 8: 1490.
- 25. Favaro, G., Bogialli, S., Di Gangi, I.M., Nigris, S., Baldan, E. and Squartini, A., Characterization of lipopeptides produced by *Bacillus licheniformis* using liquid chromatography with accurate tandem mass spectrometry. *Rapid communications in mass spectrometry*, 2016; 30(20): 2237-2252.
- 26. Fraser, P.D. and Bramley, P.M. The biosynthesis and nutritional uses of carotenoids. *Progress in lipid research*, 2004; 43: 228-265.
- Gaofu, Q., Fayin, Z., Peng, D., Xiufen, Y., Dewen, Q., and Ziniu, Y. Lipopeptide induces apoptosis in fungal cells by a mitochondira peptide in Gramnegative bacteria. *FEMS microbiol. Lett.*, 2010; 330: 81-89.
- 28. Gautam, N., and Sharma, N. Bacteriocin: safest approach to preserve food products. *Indian Journal of Microbiology*, 2009; 49: 204-211.
- 29. Ghosh, D., Charttora, D.K., and Chattopadhyay, P. Studies on changes in microstructure and proteolysis in cow and soymilk curd during fermentation using lactic cultures for improving protein bioavailability. *Journal of* food *Science Technology*, 2013; 50: 979-985.
- Gopikrishna, T., Suresh Kumar, H.K., Perumal, K. and Elangovan. E. Impact of *Bacillus* in fermented soybean foods on human health. Ann. Microbiol., 2021; 71: 30.
- Hayatun, F., Nur, H., Sri, D., Hendra, M., Ayu, R.S., Nurrahman, N., Aditya, R.E., Dewi, S.Z., Widjarnarka, W. and Stalis, N.E. Prospects of fibrinolytic proteases of bacteria from sea cucumber fermentation products as antithrombolytic agent. *BIO Web of conferences*, 2020; 28: 02006.
- 32. Hoa, N.T., Baccigalupi, L., Huxham, A., Smertenko, A., Van, P.H. and Ammendola, S., Characterization of *Bacillus* species used for oral bacteriotherapy and bacterioprophylaxis of gastrointestinal disorders. Appl environ microbiol, 2000; 66: 5241-47.
- 33. Hoang, T.A.T., Vishal, C., Sarn, S. and Cheunjit, P. Stress tolerance-*Bacillus* with a wide spectrum bacteriocin as an alternative approach for food bioprotective culture production. *Food control*, 2022; 133, part A, 108598.
- Huang, T., Geng, H., Miyyapuram, V.R., Sit, C.S., Vederas, J.C. and Nakano, M.M. Isolation of a variant of subtilosin A with hemolytic activity. *Journal of Bacteriology*, 2009; 191(18): 5690-5696.
- 35. Hyung, M.J. Kwang-soo, K., Jong-Hyun, P., Young Bae, K., Han-Joon, H. Bacteriocin with a broad antimicrobial spectrum; produced by *Bacillus* sp.

Isolated from Kimchi. *Journal of Microbiology and biotechnology*, 2001; 577-584.

- Ibeabuchi, J.C., Olawuni, I.A., Iheafgwara, M.C., Ojukwu, M. and Ofoedu, C.E. Microbiological evaluation of 'Iru' and 'Ogiri-isi' used as a food condiment. *Journal of environmental science*, *toxicology and food technology*, 2014; 8(8): 45-50.
- 37. Isu, N.R. and Ofuya, C.O. Improvement of the traditional processing and fermentation of African oil bean (*Pentaclethra macrophyla*) seeds into a food snack *ugba*'. *International Journal of Food Microbiology*, 2000; 59: 235-239.
- Jezewska-Frackowiak, J., Seroczynska, K., Banaszczyk, J, Jedrzejczak, G., Zylicz-Stachula, A. and Skowron, P.M. The promises and risks of probiotic *Bacillus* species. *Acta Biochim. Pol.*, 2018; 65: 509-519.
- Joshi, S.J., Suthar, H., Yadav, A.K., Hingurao, K., and Nerurkar, A. Occurrence of producing bacillus spp. In diverse habitats. *ISRN Biotechnology 2013* article ID 652340, 2013; 6.
- 40. Jung, W.J., Mabood, F., Souleimanov, A., Zhou, X., Jaoua, S., Kamoun, F., and Smith, D.L. Stability and antibacterial activity of bacteriocins produced by *Bacillus thurigiensis* and *Bacillus thurigiensis* sp. *kurstaki. J Microbiol Biotechnol*, 2008; 18(11): 1836-11840.
- 41. Kashyap, D.R., Vohra, P.K., Chopra, T.A., and Tewari, S.R. Applications of pectinases in the commercial sector: *A review. Bioresource Technology*, 2001; 77(3): 215-227.
- Kawai, K., Kamochi, R., Oiki, S., Murata,K., and Hashimoto, W. Probiotics in human gut microbiota can degrade host glycosaminoglycans. Sci Rep., 2018; 8: 10674.
- Khaneja, R., Perez-Fons, L., Fakhry, S., Baccigalupi, L., Steiger, S., To, E., Sandmann, G., Dong, T.C., Ricca, E., Fraser, P.D. and Cutting, S.M. Carotenoids found in *Bacillus. Journal of Applied Microbiology*, 2010; 108: 1889-1902.
- Kiers, J.L., Van Laeken, A.E.A., Rombouts, F.M. and Nout, M.J.R. *In vitro* digestibility of *Bacillus* fermented soya bean. *Int J. Food Microbiol*, 2000; 60: 163-169.
- Kiers, J.L., Van Laelan, A.E.A., Rombouts, F.M. and Nout, M.J.R. In Vitro digestibility of *Bacillus* fermented soya bean. *Int. J. Food Microbiol*, 2000; 60: 163-169.
- Kotowicz, N., Bhardwaj, R.K., Ferreira, W.T., Hong, H.A., Olender, A., Ramirez, J., Cutting, S.M., Safety and probiotic evaluation of two bacillus strains producing antioxidant compounds. *Beneficial microbes*, 2019; 10(7): 759-771.
- Leclere, V., Bechet, M., Adam, A., Guez, J.S., Wathelet, B., and Ongena, M., Mycosubtilin overproduction by *Bacillus subtilis* BBG100 enhances the organism's antagonistic and biocontrol activities. *Appl. Environ. Microbiol*, 2005; 71: 4577-4584.
- 48. Lee, S.K., Bae, D.H., Kwon, T.J., Lee, S.B., Lee,

H., Park, J.H., Heo, S., and Johnson, M.G. Purification and characterization of a fibrinolytic enzyme from *Bacillus* spp KDO-13 isolated from soybean paste. *Journal of microbiology and biotechnology*, 2001; 11(5): 845-852.

- 49. Leejeerajumnean, A.A., Duckham, S.C., Owens, J.D., and Ames, J.M. Volatile compounds of *Bacillus* fermented soybeans. *Journal of Science*, *food and Agriculture*, 2001; 81: 525-529.
- Marx, R., Stein, T., Enitan, K.D., and Glaser, S.J. Structure of the *Bacillus subtilis* peptide antibiotic subtilosin A determined by 1H-NMR and matrix assisted laser desorption/ionization time-of-flight mass spectrometry. *J Protein Chem.*, 2001; 20: 501-506.
- 51. Mohammed, Y., Lee, B., Kang, Z., and Du, G., Development of a two-step cultivation strategy for the production of vitamin B12 by *Bacillus megaterium. Microbiol. Cell. Fact.*, 2014; 13: 102.
- 52. Murwan, K.S. and Ali, A.A. Effect of fermentation period on the chemical composition, *in vitro* protein digestibility and tannin content in two sorghum cultivars (Dabar and Tabat) in Sudan. *Journal of Applied Biosciences*, 2011; 39: 2602-2606.
- 53. Nigris, S., Baldan, E., Tondello, A., Zanella, F., Vitulo, N., and Favaro, G. Biocontrol traits of bacillus licheniformis GL174, a culturable endophyte of *Vitis vinifera* cv. Glera". *BMC microbiology*, 2018; 18(1): 133.
- 54. Noll, K.S., Sinko, P.J., and Chikindas, M.L. Elucidation of the molecular mechanisms of action of the Natural antimicrobial peptide subtilosin against the bacterial vaginosis-associated pathogen Gardnerella vaginalis. *Probiotics antimicrobe*. *Proteins*, 2011; 3(1): 41-47.
- 55. Nout, M.J.R. Rich nutrition from the poorest-cereal fermentation in Africa and Asia. *Food microbiology*, 2009; 26(7): 685-692.
- 56. Nurudeen, A.O. and Princewill, C.O. African fermented food condiments: microbiology impacts on their nutritional values, frontiers, and new trends in the science of fermented food and beverages. Rosa Lidia Solis-Oviedo and *Angel de le Cruz Pech-Canul, Intech Open*, 2019.
- 57. Ogbonna, D.N., Sokari, T.G., and Achinewhu, S.C. Development of owoh-type product from African yam beam (*Sphenostylis stenocarpa*) seeds by solid state fermentation. *Plant Foods for Human Nutrition*, 2001; 56: 183-194.
- 58. Ogueke, C.C., and Aririatu, L.E. Microbial and organoleptic changes associated with 'ugba' stored at ambient temperature. *Nigerian food journal*, 2004; 22: 133-140.
- 59. Oguntoyinbo, F.A., Sanni, A.I., Franz, C.M.A.P. and Holzapfel, W.H. In *vitro* fermentation studies for selection and evaluation of *Bacillus* strains as starter cultures for production of *Okpehe*, traditional Africa fermented condiment. *International Journal of Food Microbiology*, 2007; 113: 208-218.
- 60. Oladipo, I. C., Sanni, A. I., Chakraborty, W.,

Chakravorty, S., Jana, S., Rudra, D. S., Gacchui, R. and Swarnakar, S. Bioprotective potential of bacteriocinogenic *Enterococcus gallinarum* strains isolated from some Nigerian fermented foods, and of their bacteriocins. *Polish Journal of Microbiology*, 2014; 63(4): 415–422.

- Oladipo, I. C., Sanni, A. I., Chakraborty, W., Chakravorty, S., Jana, S., Rudra, D. S., Gacchui, R. and Swarnakar, S. Technological properties of strains of *Enterococcus gallinarum* isolated from selected Nigerian traditional fermented foods. *Malaysian Journal of Microbiology*, 2015; 11(1): 1-13.
- 62. Olasupo, N.A. and Okorie, P.C. African fermented food condiments: microbiology impacts on their nutritional values. Frontiers and new trends in the science of fermented food and beverages, 2018. doi: 10.15772/intechopen.83466
- 63. Omafuvbe, B.O., Olumuyiwa, S.F., Osuntogu, B.A. and Adewusi, R.A. chemical and biochemical in African Locust Bean (*Parkia biglobosa*) and melon (*Citrullus vulgaris*) seeds During fermentation to condiments. *Parkistan Journal of nutrition*, 2004; 3(3): 140-145.
- 64. Omafuvbe, B.O., and Kolawole, D.O. Quality assurance of stored pepper (*Piper guineense*) using controlled processing methods. *Pakistan Journal of nutrition*, 2004; 3: 244-249.
- 65. Omura, K., Histosugi, M., Zhu, X., Ikeda, M., Maeda, H and Tofudome, S. A newly derived protein from Bacillus subtilis natto with both antithrombolytic and fibrinolytic effects. *J Pharmacol Sci.*, 2005; 99: 247-251.
- 66. Onwurafor, E.U., Onweluzo, J.C., and Ezeoke, A.M. Effects of fermentation methods on chemical and microbial properties of mung bean (*Vigna radiata*) flour. *Niger food journal*, 2014; 32: 89-96.
- 67. Osungbaro, T.O. Physical and nutritive properties of fermented cereal foods. *African Journal of Food Science*, 2009; 3(2): 23-27.
- Ouoba, L.I.I., Cantor, M.D., Diawara, B., Traoire, A.S and Jakobsen, M., Degradation of African locust bean oil by *Bacillus subtilis* and *Bacillus pumilis* isolated from *soumbala*, a fermented African locust bean condiment. *J Appl Microbiol*, 2003b; 95: 868-873.
- 69. Ozabor, P.T., Olaitan, J.O., Olaosun, O.S., and Fadahunsi, I.F. Antibacterial and antioxidant activity of *Bacillus* species isolated from fermented *Parkia biglobosa* (Iru) and *Ricinus communis* (Ogiri)-African traditionally fermented food condiments. *The Asian journal of Applied Microbiology*, 2020; 7(1): 19-29.
- Palffy, R., Gardlik, R., Behuliak, M., Kadasi, L., Turna, J., and Celec, P., On the physiology and pathophysiology of antimicrobial peptides. *Mol Med.*, 2009; 15(1-2): 51-59.
- Parkouda, C., Nielsen, D.S., Azokpota, P., Ivette Irene Ouoba, L., Amoa-Awua, W.K. and Thorsen, L. The microbiology of alkaline fermentation of

indigenous seeds used as food condiments in Africa and asia. *Crit Rev Microbiol*, 2009; 35: 139-156.

- 72. Pattnaik, P., Kaushik, J.K. Grover, S., and Batish, V.K. Purification and characterization of a bacteriocin-like compound (lichenin) produced anaerobically by *Bacillus licheniformis* isolated from water buffalo. *J appl Microbiol*, 2001; 91(4): 636-645.
- 73. Paul, S.I., Rahman, M.M., Salam, M.A., Khan, M.A., and Islam, M.T. Identification of marine sponge-associated bacteria of the saint Martin's Island of the Bay of Bengal emphasizing on the prevention of motile *Aeromonas septicemia* in *Labeo rohita. Aquaculture*, 2021; 545: 737156.
- 74. Pederson, P.B., Bjornvard, M.E., Rasmussen, M.D., and Petersen, J.N. Cytotoxic potential of industrial strains of *Bacillus* spp. *Regulatory* toxicology and pharmacology, 2002; 36: 155-161.
- 75. Pepin, J., Valiquette, L., Alary, M.E., Villemure, P., Pelletier, A., Forget, K., Pepin, K., and Chouinard, D. *Clostridium difficile* associated diarrhea in a region of Quebec from 1991 to 2003: a changing pattern of disease severity. *Canadian Medical Association Journal*, 2004; 171: 466-472.
- 76. Perez-Fonz, L., Steiger, S., Khnaja, R., Bramley, P.M., Cutting, S.M. Sandmann, G. and Fraser, P.D. Identification and the developmental formation of carotenoid pigments in the yellow/orange *Bacillus* spore-formers. *Biochemical et Biophysica Acta*, 2011; 1811: 177-185.
- 77. Pinchuk, I.V., Bressollier, P., Vernecuil, B., Fenet, B., Sorokulova, I.B., Graud, F., and Urdaci, M.C. *In vitro* anti-*Helicobacter pylori* activity of the probiotic strain *Bacillus subtilis* 3 is due to secretion of antibiotics. *Antimicrobial Agents and Chemotherapy*, 2001; 45: 3156-3161.
- Pinontoan, R., City, S., Widjaja, A.N. and Purnomo, J.S. The fibrinolytic potential of isolated from saltfermented shrimp paste *terasi*. *Biodiversitas*, 2024; 25(7): 3193-3199.
- Puan, S.L., Erriah, P., Baharudin, M.M.A., Yahaya, N.M., Kamil, W.N.I.W.A. and Ali, M.S.M. Antimicrobial peptides from *Bacillus* spp. and strategies to enhance their yield. *Appl. Microbiol. Biotechnol*, 2023; 107: 5569-5593.
- Rahman, M.M., Paul, S.I., Alter, T., Tay. A C., Foysal, M.J., and Islam, M.T., whole genome sequence of *Bacillus subtilis* WSIA, a promising fish probiotic strain isolated from marine sponge of the Bay of Bengal. *Microbiology resource announcements*, 2020; 9(39): e00641-20.
- Rao, A.V. and Rao, L.G. Carotenoids and human health. Pharmacological research: *The Official Journal of the Italian Pharmacological Society*, 2007; 55: 207-216.
- 82. Riccardo, I., Bovenberg, R. and Driessen, A. Correction to non-ribosomal peptide synthases and their biotechnological potential in *Penicillium rubens.*, 2022; 49: kuac005.
- 83. Romco-tabarez, M., Jansen, R., Sylla, M., Lunsdorf,

H., Haubler, S., Santosa, D.A., Timmis, K.N., and Molinari, G. 17-0-malonyl macrolactin A, a new macrolactrin antibiotic from *Bacillus subtilis* active against methicilin -resistant *Staphylococcus aureus*, vancomycin resistant *Enterococci* and a small colony variant of *Burkholderia cepacian*. *Antimicrobial agents and chemotherapy*, 2006; 50: 1701-1709.

- Sanni, A.I., Onilude, A.A., Fadahunsi, I.F., Ogubanwo, S.T., and Afolabi, R.O. Selection of starter cultures for the production of *ugba*', a fermented soup condiment. *European food research technology*, 2002; 215: 176-180.
- 85. Santos, F., Wegkamp, A., De Vos, W.M., Smid, DE.J. and Hugenholtz, J. High- level folate production in fermented foods by the B12 producer Lactobacillus *reuteri* JCM1112. *Applied and Environmental Microbiology*, 2008; 74(10): 3291-94.
- Scott, R.W., DeGrado, W.F., and tew, G.N. De novo designed synthetic mimics of antimicrobial peptides. *Curr Opin Biotechnol*, 2008; 19(6): 620-627.
- Schallmey, M., Singh, A. and Ward, O.P. Developments in the use of bacillus species for industrial production. *Can. J. Microbiol*, 2004; 50: 1-17.
- Shafi, J., and Tian, H. *Bacillus* species as versatile weapons for plant pathogens: A review of biotechnology. *Biotechnol. Equ.*, 2017; 31: 446-459.
- Sirtori, L.R., Cladera-Olivera, F., Lorenzini, D.M., Tsai, S.M., and Brandelli, A. Purification and partial characterization of an antimicrobial peptide produced by *Bacillus* sp. Strain P45, a bacterium from the amazon basin fish *Piaractus mesopotamics*. J. *Gen. App. Microbiol*, 2006; 52(6): 357-363.
- Sultyak,K.E., Wirawan, R.E., Aroutchva, A.A. and Chikindas, M.L. Isolation of the *Bacillus subtilis* antimicrobial peptide subtilosin from the diary product-derived *Bacillus amyloliquefaciens*. J Appl Microbiol, 2008; 52(6): 357-363.
- Sumi, C.D., Yang, B.W., Yeo, I.C. and Hahm, Y.T. Antimicrobial peptides of the Genus *Bacillus*: A new era for antibiotics. *Canadian journal of microbiology*, 2015; 61: 93-103.
- 92. Sy, C., Caris-Veyrat, C., Dufour, C., Boutaleb, M., Borel, P., and Dangles, O. Inhibition of iron-induced lipid peroxidation by newly identified bacterial carotenoids in model gastric conditions: Comparison with common carotenoids. *Food and Function*, 2013a; 4: 698-712.
- 93. Syahbanu, F., Kezia, E., Puera, N., Giriwono, P.E., Tjandrawinata, R.R and Suhartono, M.T. Fibrinolytic bacteria of Indonesian fermented soybean: preliminary study on enzyme activity and protein profile. *Food science and technology*, Campina, 2019; 40(2): 458-465.
- 94. Tabenne, O., Gharbi, D., Slimene, I.B., Elkahoui, S., Alfeddy, M.N., and Cosette, P. Antioxidative and DNA protective effects of Bacillomycin D-like lipopeptides produced by B38 strain. *Appl Biochem*

Biotechnol, 2012; 168: 2245-2256.

- 95. Tam, N.K., Uyen, N.Q., Hong, H.A., Ducle, H., Hoa, T.T., Serra, C.R., Henriques, A.O., and Cutting, S.M. The intestinal life cycle of *Bacillus subtilis* and close relatives. *Journal of Bacteriology*, 2006; 188: 2692-2700.
- 96. Thursby, E. and Jude, N., Introduction to human gut microbiota. *Biochem J.*, 2017; 474: 1823-1836.
- 97. Wang, Y., Zhu, X., Bie, X., Lu, F., Zhang, C., Yao, S. and Lu, Z. Application of electrospray ionization mass spectrometry in rapid typing of Fengycin homologues produced by *Bacillus subtilis*. *Letters in Applied Microbiology*, 2014c; 39: 98-102.
- 98. Wei, X., Luo, M., Xu, L., Zhang, Y., Lin, X., Kong, P., and Liu, H. Production of fibrinolytic enzyme from *Bacillus amyloliquefaciens* by fermentation of chickpeas, with the evaluation of the anticoagulant and antioxidant properties of chickpeas. *Journal of Agricultura and food Chemistry*, 2011; 59(8): 3957-3963.
- 99. Wu, D., Fu, L., Cao, Y., Dong, N. and Li, D. Genomic insights into antimicrobial potential and conditions of pig-derived *Bacillus subtilis* BS21. *Front. Microbiol*, 2023; 14: 1239837.
- 100.Xu, Y., Hlaing, M.M., Glagovskaia, O., Augustin, M.A. and Terefe, N.S. Fermentation by probiotic *Lactobacillus gasseri* strains enhances the carotenoid and fibre contents of carrot juice. *Foods*, 2020; 9: 1803.
- 101.Yao, Z., Meng, Y., Le, H.G., Lee, S.J., Jeon, H.S., Yoo, J.Y., Afifah, D.N and Kim, J.H. Isolation of 2 *Bacillus* strains with strong fibrinolytic activities from Kimchi. *Microbiol. Biotechnol. Lett.*, 2020; 48(4): 439-446.
- 102. Yoon, S.J., Myeong, A.Y., Gwan, S.S., Seung, T.K., Jae, K.H., Jung, K.S., Ik, H.Y. and Yu, R.P. Screening and characterization of microorganisms with fibrinolytic activity from fermented foods. *J. Microbiol. Biotechnol*, 2002; 12(4): 649-656.
- 103.Zhang, B., Dong, C., Shang, Q., Han, Y., and Li, P. New insights into membrane -active action in plasma membrane of fungal hyphae by the lipopeptide antibiotic bacillomycin L. *Biochem Biophys. Acta*, 2013; 1828: 2230-2237.
- 104.Zhang, Q.Y., Yan, Z.B., Meng, Y.M., Hong, X.Y. Shao, G. and Ma, J.J. Antimicrobial peptides: mechanism of action, activity and clinical potential. *Military Med Res.*, 2021; 8: 48.