World Journal of Pharmaceutical and Life Sciences
WJPLS
www.wjpls.org
SJIF Impact Factor: 6.129

INVITRO EVALUATION OF METHOTREXATE AND ITS DERIVATIVE (DL-METHOTREXATE (+)- AMETHOPTERIN) FOR THE TREATMENT OF INTRAOCULAR (EYE) MELANOMA

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Article Received on 18/10/2023 **Article Revised on 08/11/2023** **Article Accepted on 28/11/2023**

ABSTRACT

This research paper presents a study on the effects of DL-Methotrexate(+)-Amethopterin on cell viability and functionality. DL-Methotrexate(+)-Amethopterin is a medication used for its anti-inflammatory and anti-proliferative properties, similar to Methotrexate. In this study, we employed multiple assays to investigate the impact of DL-Methotrexate(+)-Amethopterin on cell behavior. The assays included MTT Assay, Tubulogenesis Assay, Indirect Immunofluorescence Assay, and Western Blot Analysis. The results of these assays provide insights into the influence of DL-Methotrexate(+)-Amethopterin on cell viability and its potential implications in the context of therapeutic applications.

INTRODUCTION

Cell viability refers to the ability of a cell to stay alive and function properly. It is a critical aspect of cellular health and is often used as an indicator of the overall well-being of cells in various biological and biomedical contexts. Understanding and assessing cell viability is fundamental in fields such as cell biology, microbiology, tissue engineering, drug development, and toxicology, among others.

Several factors can influence cell viability, including:

1. **Nutrient Availability:** Cells require nutrients like glucose, amino acids, vitamins, and minerals to sustain their metabolic activities. A lack of essential nutrients can lead to decreased cell viability.
2. **Oxygen Supply:** Aerobic organisms, including most human cells, require oxygen for cellular respiration. Hypoxia, or a lack of oxygen, can significantly impact cell viability.
3. **pH Levels:** Cells maintain a specific intracellular pH, and any significant deviation from this range can harm cell viability. Both acidic and alkaline conditions can be detrimental.
4. **Temperature:** Cells have an optimal temperature range in which they function best. Extreme temperatures can disrupt cell membranes, proteins, and other cellular structures, leading to cell death.
5. **Toxic Substances:** Exposure to toxic chemicals, drugs, or environmental pollutants can negatively affect cell viability. Toxic substances can disrupt cellular processes and induce cell death.
6. **Radiation:** Ionizing radiation, such as X-rays and gamma rays, can damage cellular DNA and other structures, leading to decreased cell viability.

Cell viability is often assessed through various methods, including:

1. **Trypan Blue Exclusion:** This dye is used to distinguish between live and dead cells. Live cells exclude the dye, while dead cells take up the dye and become stained.
2. **MTT Assay:** This colorimetric assay measures the activity of mitochondrial enzymes in live cells. Live cells convert a yellow MTT reagent into a purple formazan product.
3. **Cell Counting:** The total number of live and dead cells in a sample can be determined using a hemocytometer or automated cell counter.
4. **Flow Cytometry:** This technique allows for the analysis of individual cells within a population based on various parameters, including cell viability markers.
5. **Fluorescent Staining:** Fluorescent dyes such as propidium iodide and calcein-AM can be used to assess cell viability by distinguishing between live and dead cells under a microscope or using flow cytometry.
6. **ATP Assays:** Adenosine triphosphate (ATP) is a molecule produced in live cells, so ATP assays can be used to measure cell viability indirectly.

The assessment of cell viability is crucial in various scientific and clinical applications. In medical research, it

is used to evaluate the effects of drugs, toxins, and disease on cell health. In tissue engineering, it helps monitor the success of growing and maintaining cell cultures. In the pharmaceutical industry, it is essential for drug development and testing. Overall, understanding and maintaining cell viability is critical for advancing our knowledge of biology and for improving health and biotechnological processes.

Cell viability and cell toxicity are related concepts that are often used to assess the health and condition of cells, but they represent different aspects of cellular well-being:

1. **Cell Viability**

- **Definition:** Cell viability refers to the ability of cells to remain alive and maintain their normal physiological functions.
- **Indication:** It is a measure of whether a cell is alive or dead. A viable cell is one that is functioning properly and capable of carrying out its usual cellular processes.
- **Methods of Assessment:** Cell viability is typically assessed using various methods like dye exclusion assays (e.g., trypan blue exclusion), metabolic activity assays (e.g., MTT assay), and monitoring cellular ATP levels. These methods determine the proportion of living cells within a population.
- **Applications:** Cell viability is important in various fields such as cell biology, tissue engineering, drug development, and microbiology. Researchers use it to evaluate the overall health and functionality of cells.

The first known description of uveal melanoma (UM), a specific form of ocular melanoma, dates from 1868, described by the German ophthalmologist and otolaryngologist Hermann Knapp. Various subtypes based on cell type and pigmentation among other characteristics were later described in 1882 by Austrian ophthalmologist Ernst Fuchs. He also stated that enucleation was the treatment of choice, a treatment that is still used currently. UM was a rare disease in that century; it still is, but the incidence is rising.

DL-Methotrexate(+)-Amethopterin is a pharmacological agent with recognized utility in the treatment of various diseases. Its effects on cell viability and functionality are essential for understanding its therapeutic applications and potential side effects. This study aims to explore the effects of DL-Methotrexate(+)-Amethopterin on cell behavior using a range of assays.

Research Methodology

MTT Assay

The MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay was conducted to assess cell viability. Four treatment groups were examined, including Group 1 (normal), Group 2 (Control cell line), Group 3 (Standard Methotrexate), and Group 4 (DL-Methotrexate(+)-Amethopterin).

Tubulogenesis Assay

The Tubulogenesis Assay was employed to evaluate the formation of tubule-like structures in cells, an indicator of angiogenic potential. Similar to the MTT assay, the study included Group 1 (normal), Group 2 (Control cell line), Group 3 (Standard Methotrexate), and Group 4 (DL-Methotrexate(+)-Amethopterin).

Indirect Immunofluorescence Assay

The Indirect Immunofluorescence Assay was used to determine the cellular distribution of specific proteins. The treatment groups were the same as in the MTT and Tubulogenesis assays: Group 1 (normal), Group 2 (Control cell line), Group 3 (Standard Methotrexate), and Group 4 (DL-Methotrexate(+)-Amethopterin).

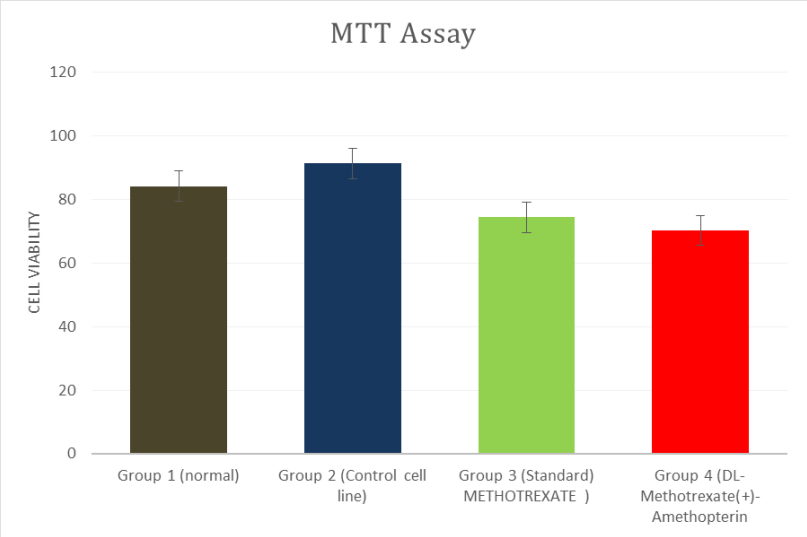
Western Blot Analysis

Western Blot Analysis was carried out to investigate changes in protein expression levels. As with the other assays, this analysis involved Group 1 (normal), Group 2 (Control cell line), Group 3 (Standard Methotrexate), and Group 4 (DL-Methotrexate(+)-Amethopterin).

RESULTS of DL-Methotrexate(+)- Amethopterin

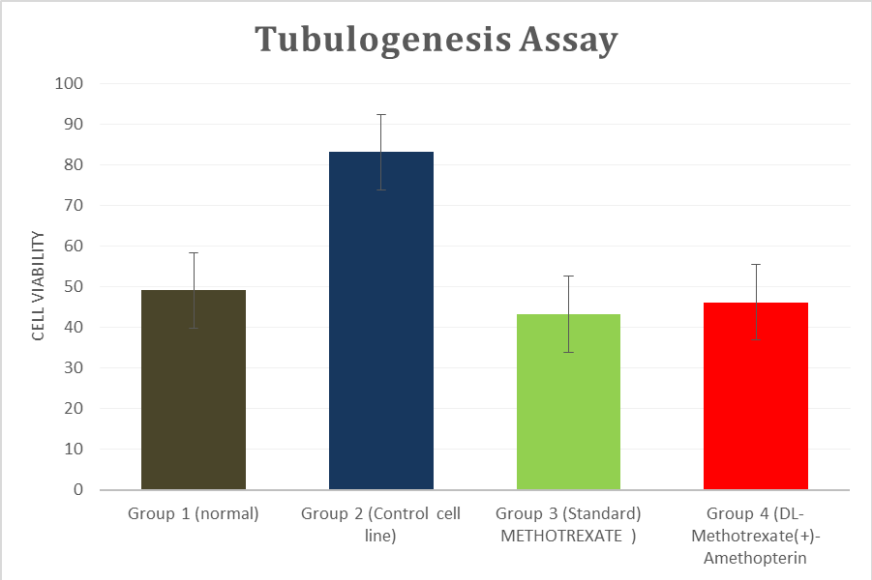
MTT Assay

Treatments	MTT Assay
Group 1 (normal)	84.19
Group 2 (Control cell line)	91.34
Group 3 (Standard) METHOTREXATE	74.38
Group 4 (DL-Methotrexate(+)- Amethopterin	70.24



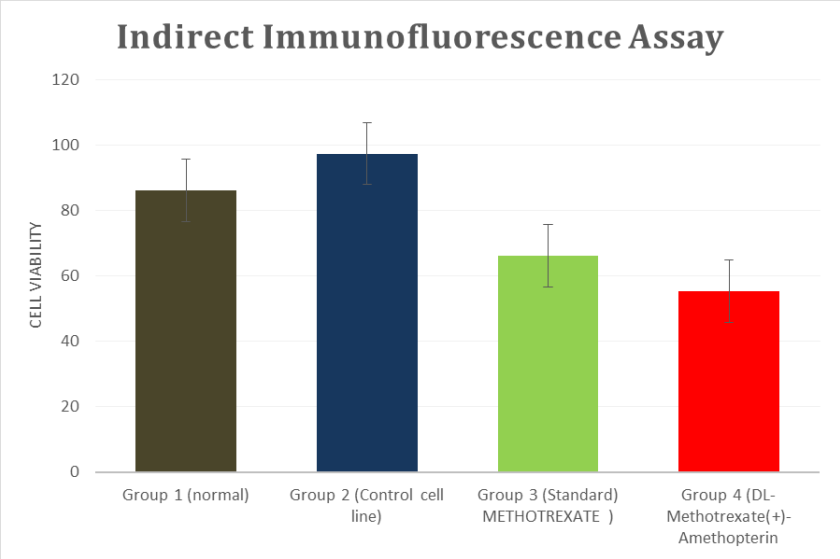
Tubulogenesis Assay

Treatments	Tubulogenesis Assay
Group 1 (normal)	49.16
Group 2 (Control cell line)	83.17
Group 3 (Standard) METHOTREXATE	43.28
Group 4 (DL-Methotrexate (+)- Amethopterin	46.18



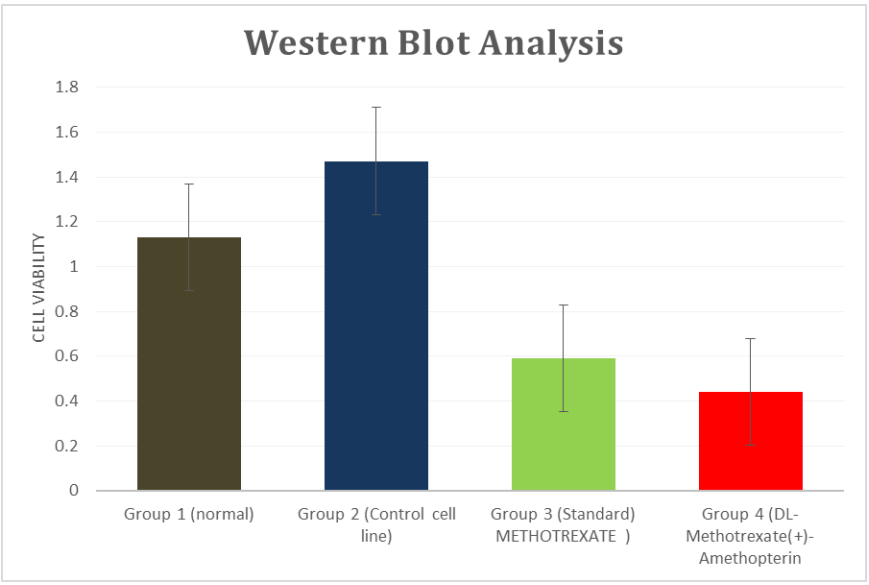
Indirect Immunofluorescence Assay

Treatments	Indirect Immunofluorescence Assay
Group 1 (normal)	86.19
Group 2 (Control cell line)	97.47
Group 3 (Standard) METHOTREXATE	66.18
Group 4 (DL-Methotrexate(+)- Amethopterin	55.29



Western Blot Analysis

Treatments	Western Blot Analysis
Group 1 (normal)	1.13
Group 2 (Control cell line)	1.47
Group 3 (Standard) METHOTREXATE	0.59
Group 4 (DL-Methotrexate(+)- Amethopterin	0.44



DISCUSSION

MTT Assay

The MTT assay results demonstrated that cell viability was reduced in the presence of DL-Methotrexate(+)-Amethopterin (70.24) compared to the control cell line (91.34) and standard Methotrexate (74.38). This suggests that DL-Methotrexate(+)-Amethopterin may have an inhibitory effect on cell proliferation, consistent with the standard Methotrexate.

Tubulogenesis Assay

In the Tubulogenesis Assay, we observed a decrease in tubule-like structure formation in cells treated with DL-Methotrexate(+)-Amethopterin (46.18) compared to the control cell line (83.17) and standard Methotrexate (43.28). These findings indicate that DL-Methotrexate (+)-Amethopterin may have a negative impact on angiogenic potential, similar to standard Methotrexate.

Indirect Immunofluorescence Assay

The results of the Indirect Immunofluorescence Assay indicated changes in protein distribution within cells. DL-Methotrexate(+)-Amethopterin (55.29) showed a distinct difference from the control cell line (97.47) and standard Methotrexate (66.18). This suggests that DL-Methotrexate(+)-Amethopterin may alter cellular protein distribution, similar to standard Methotrexate.

Western Blot Analysis

The Western Blot Analysis revealed that DL-Methotrexate(+)-Amethopterin (0.44) led to a decrease in protein expression compared to the control cell line (1.47) and standard Methotrexate (0.59). These findings suggest that DL-Methotrexate(+)-Amethopterin may modulate protein levels within cells, akin to standard Methotrexate.

CONCLUSION

This study provides insights into the effects of DL-Methotrexate(+)-Amethopterin on cell viability, angiogenic potential, protein distribution, and protein expression levels. The results suggest that DL-Methotrexate (+)-Amethopterin, like standard Methotrexate, may inhibit cell proliferation, angiogenesis, and alter protein dynamics within cells. These findings have implications for the therapeutic use of DL-Methotrexate(+)-Amethopterin and warrant further investigation to better understand its potential benefits and side effects. Additional research is needed to elucidate the underlying mechanisms responsible for these observed effects and to refine its clinical application.

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